

# Genomes and phenotypes



Wolfgang Huber  
EMBL

Genome Biology Unit (Heidelberg) & EBI (Cambridge)

# What makes us different?

Genome-wide genotyping of individuals for  $O(10^6)$  **common variants**, by microarray, is a commodity.

Genome sequencing also detects **rare or private variants**, and structural variants.

Why is that useful?

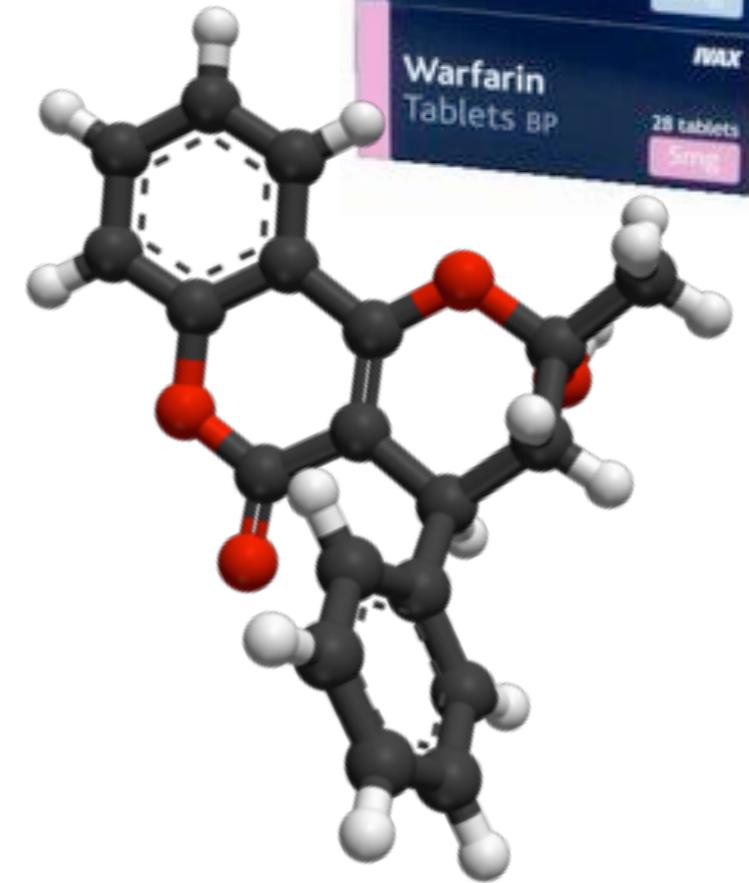
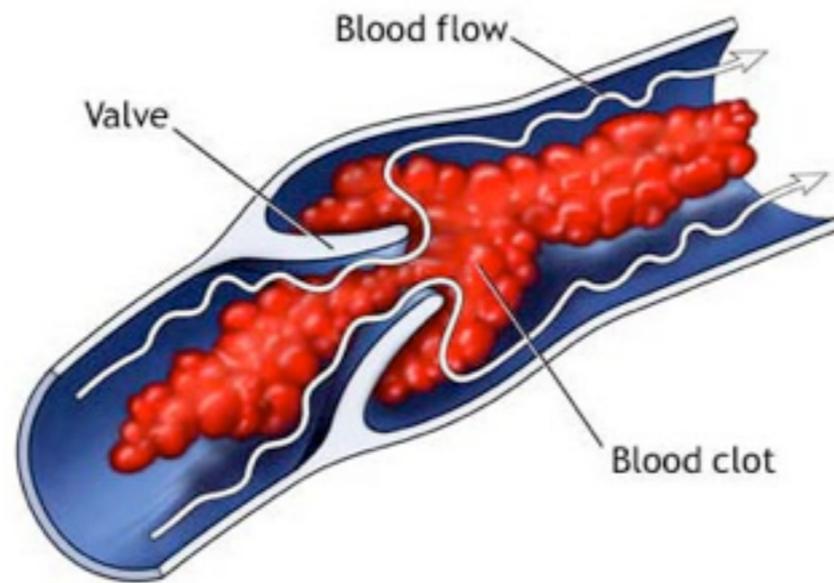


```
A G A G T T C T G C T C G
A G G G T T A T G C G C G
C G T T C G G G A A T C C
C G T T A G G A A A T C T
T C T T T G A C G A C T C
T C T T A G A G G A C T C
```

# Warfarin

First use: rat and mice killer

Anticoagulant. Prevents embolism and thrombosis



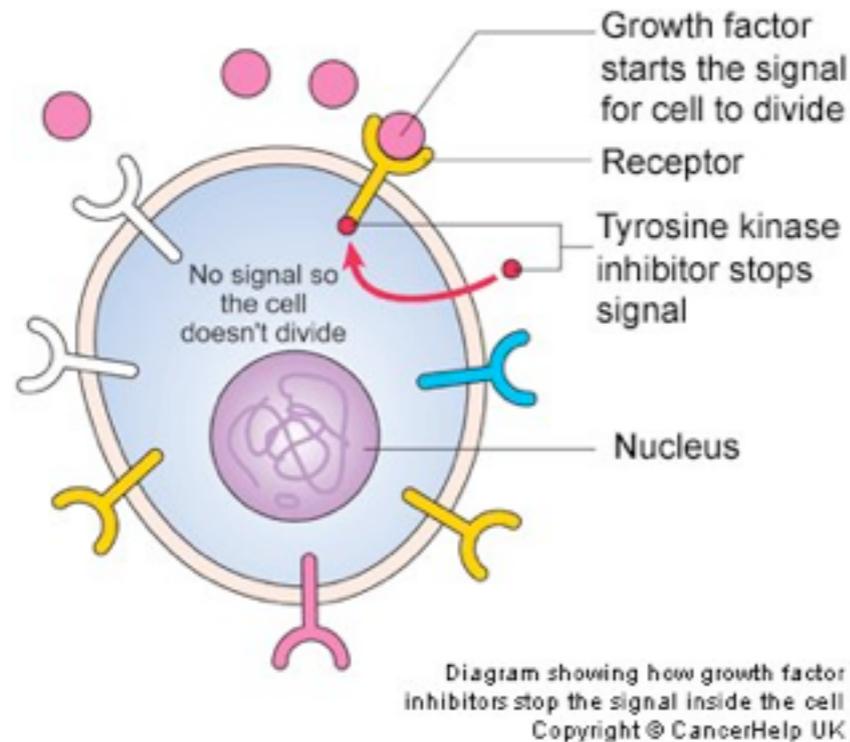
Dose requirement ~ clinical & demographic variables;  
VKORC1 (action)  
CYP2C9 (metabolism)

# Herceptin

Monoclonal antibody that interferes with the ERBB2 receptor.



# Tyrosin Kinase Inhibitors



- Erlotinib (Tarceva)
- Imatinib (Glivec)
- Gefitinib (Iressa)
- Dasatinib (Sprycel)
- Sunitinib (Sutent)
- Nilotinib (Tasigna)
- Lapatinib (Tyverb)
- Sorafenib (Nexavr)
- Temsirolimus (Torisel)

**NSCLC: resistance to TKI therapy  $\Leftarrow$  heterogeneity and mutational redundancy of the disease**

**Identify each patient's specific 'driver mutations'**

**E.g. Activation of EGFR by exon 19 deletion or exon 21 mutation  $\Rightarrow$  erlotinib and gefitinib**

**.... etc.**

# Genome-wide association studies

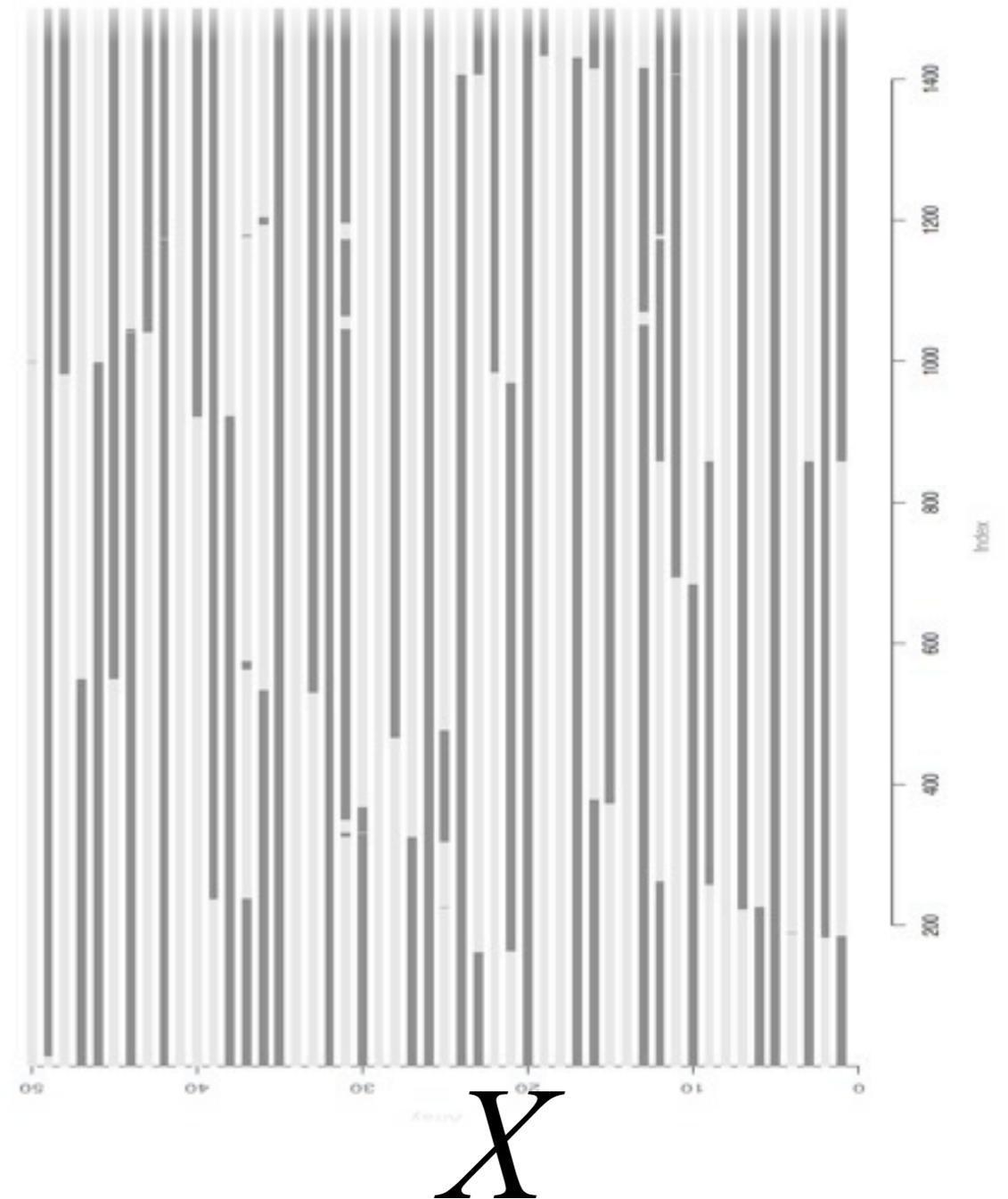


*Y*



← millions of genomic loci →

← thousands of people →



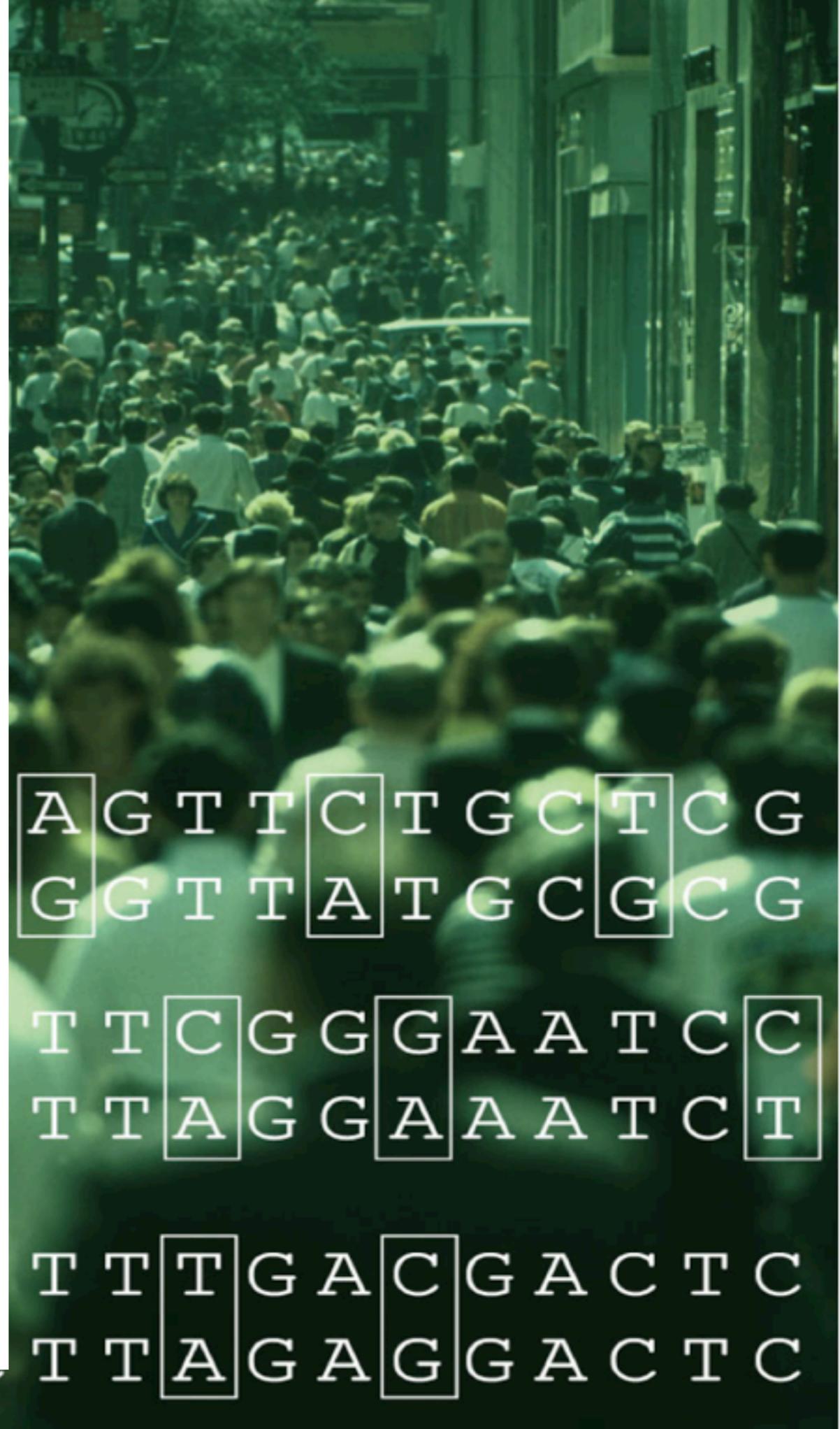
*X*

# Genome wide association studies:

**Identifiability** - additive model with no interactions

**Finding important variables (loci):**  
impressive

**Prediction** performance, effect sizes: poor



A G T T C T G C T C G  
G G T T A T G C G C G

T T C G G G A A T C C  
T T A G G A A A T C T

T T T G A C G A C T C  
T T A G A G G A C T C

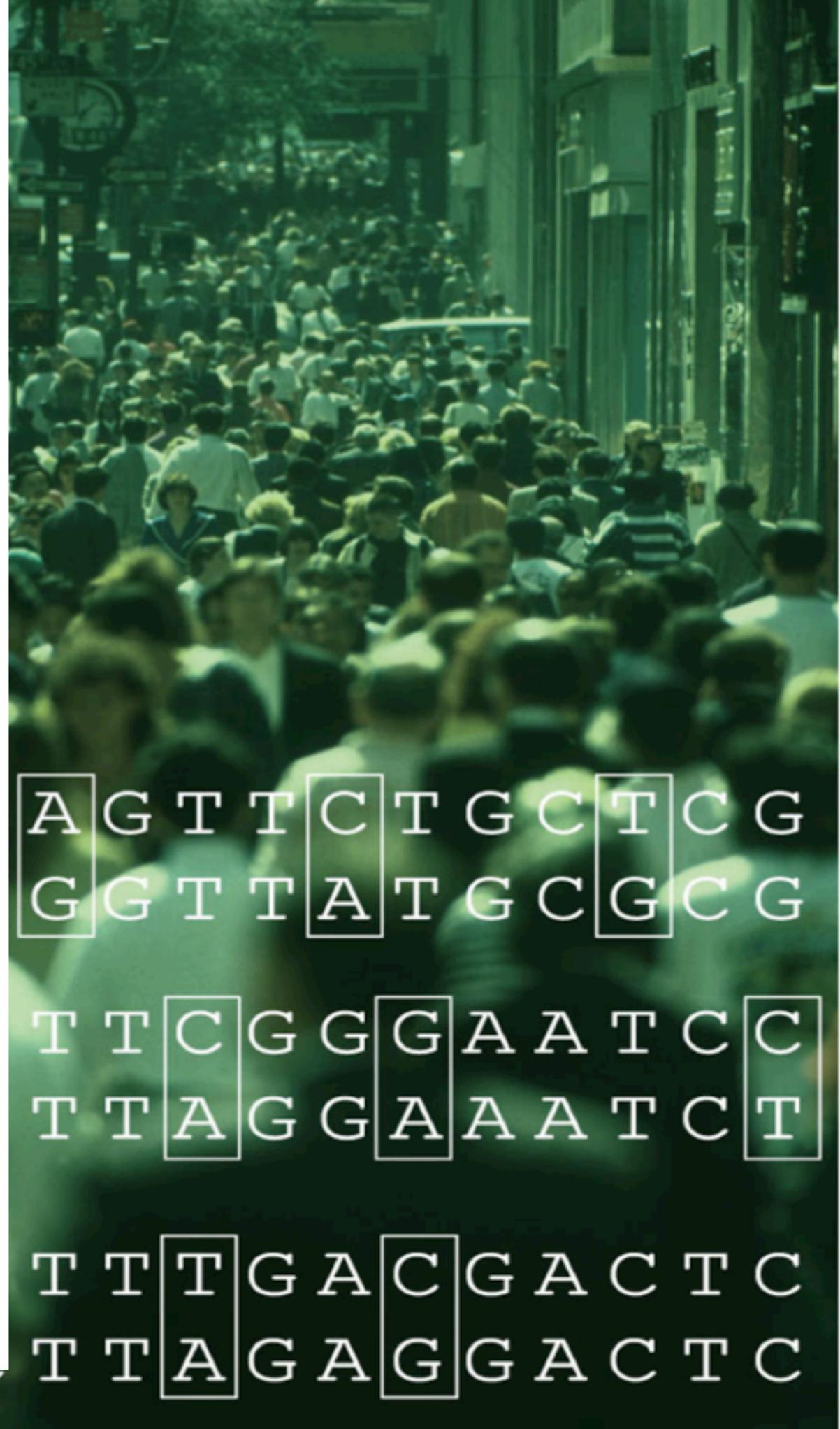
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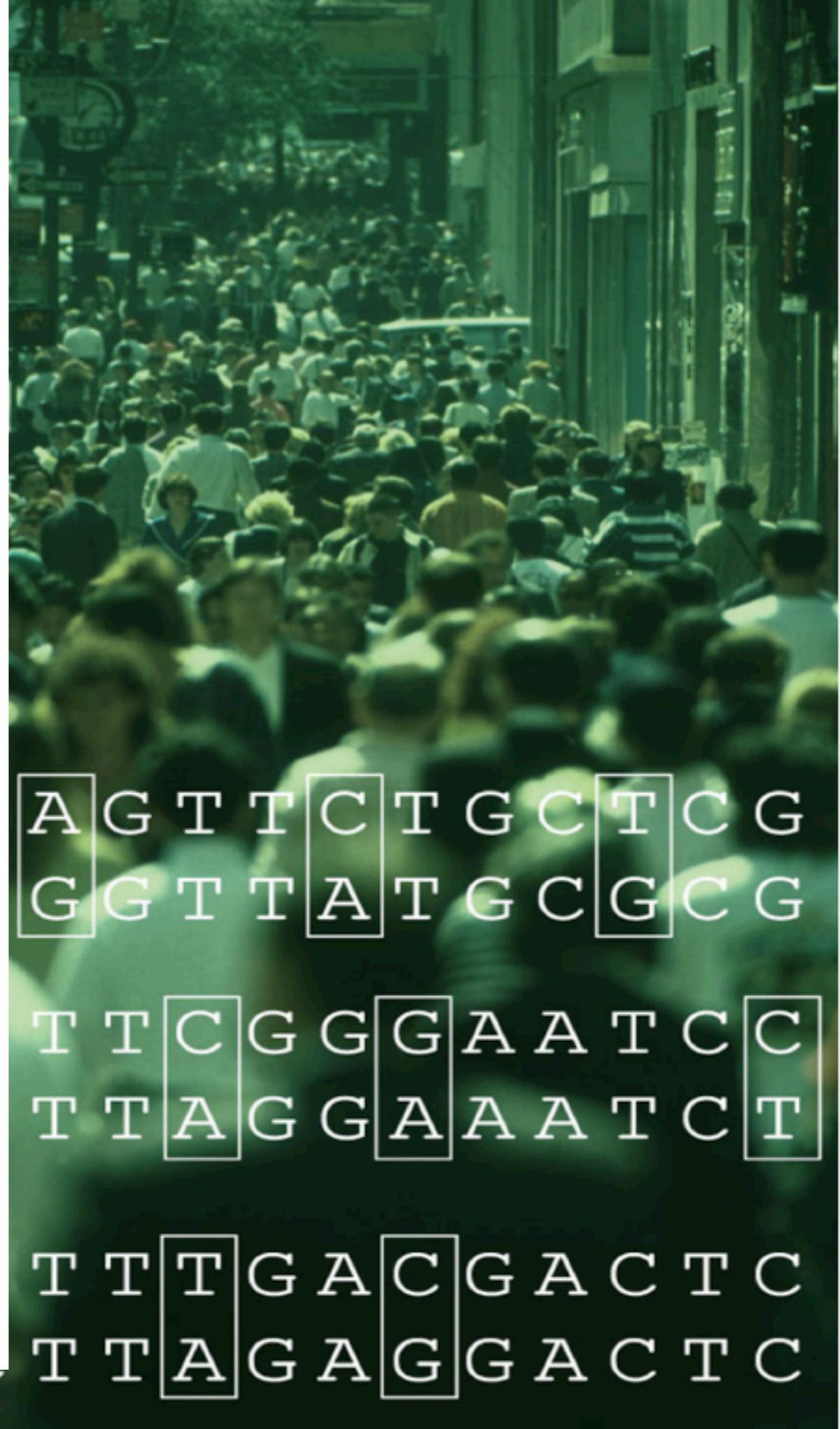
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strong effect (sufficient but not necessary)?



A G T T C T G C T C G  
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T T C G G G A A T C C  
T T A G G A A A T C T

T T T G A C G A C T C  
T T A G A G G A C T C

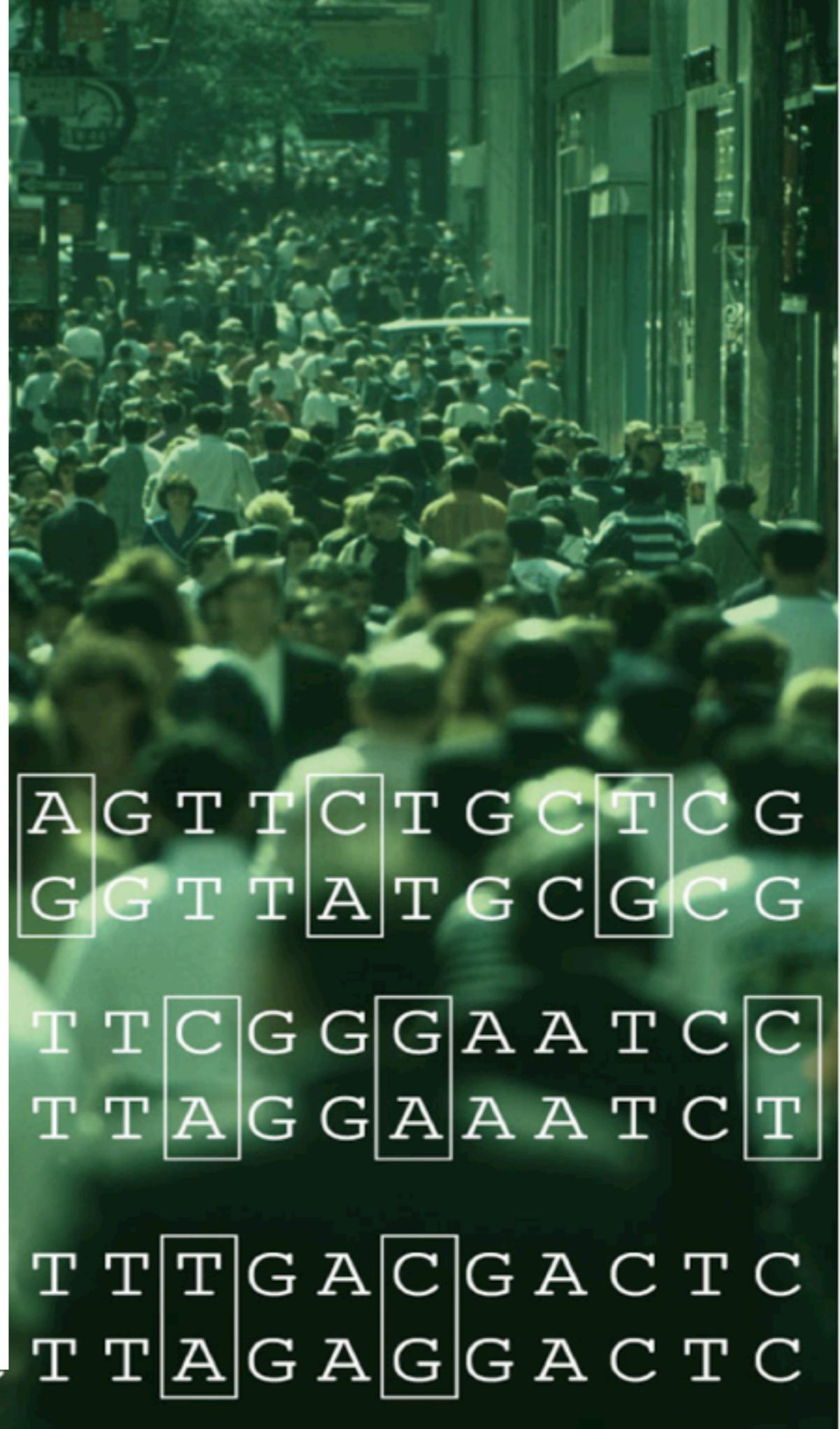
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- Interactions (epistasis)



A G T T C T G C T C G  
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T T C G G G A A T C C  
T T A G G A A A T C T

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T T A G A G G A C T C

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Association based approaches do not have enough power - we need **perturbation** experiments on model systems

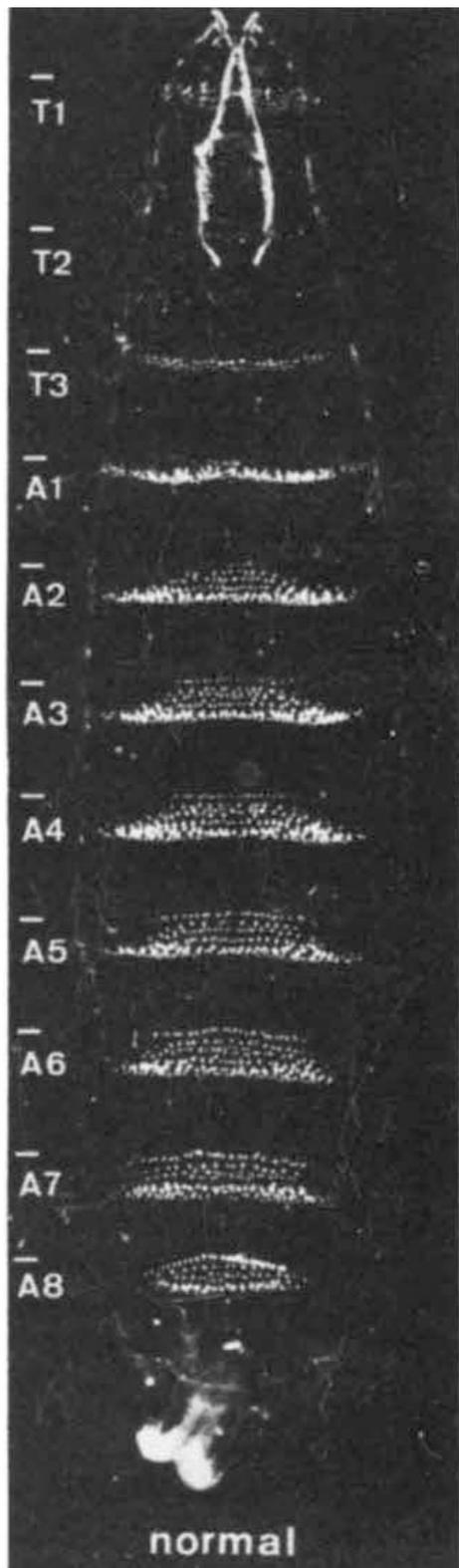


A G T T C T G C T C G  
G G T T A T G C G C G

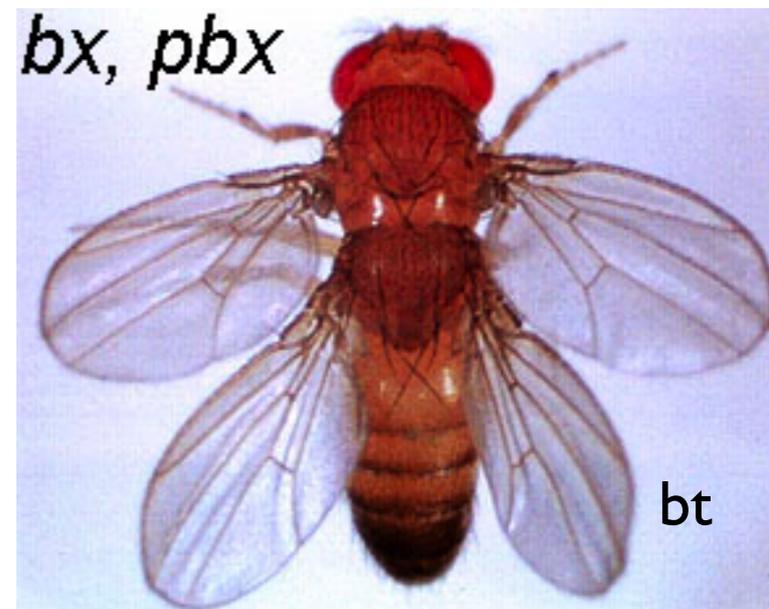
T T C G G G A A T C C  
T T A G G A A A T C T

T T T G A C G A C T C  
T T A G A G G A C T C

# Forward genetics



**Fig. 2** Ventral cuticular pattern of (from left to right) a normal *Drosophila* larva shortly after hatching, and larvae homozygous for *gooseberry*, *hedgehog* and *patch*. The mutant larvae were taken out of the egg case before fixation. All larvae were fixed, cleared and mounted as described in ref. 22. A, abdominal segment; T, thoracic segment. For further description see text and Fig. 3.  $\times 140$ .

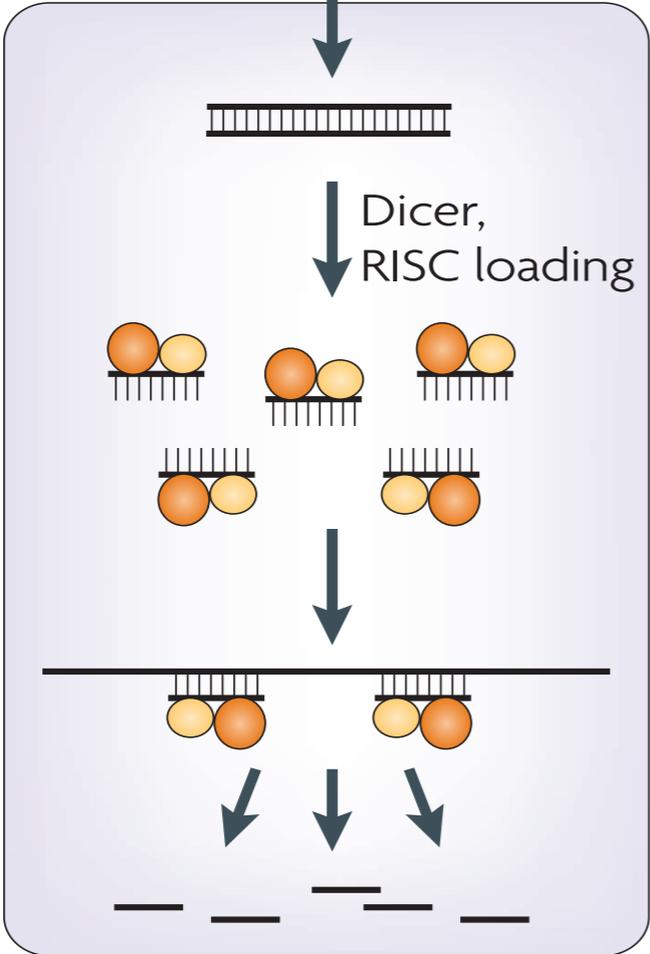


# RNAi: targeted depletion of a specific gene's products (mRNA)

*Drosophila*

Long dsRNA  
[diagram of long dsRNA]  
>100 bp

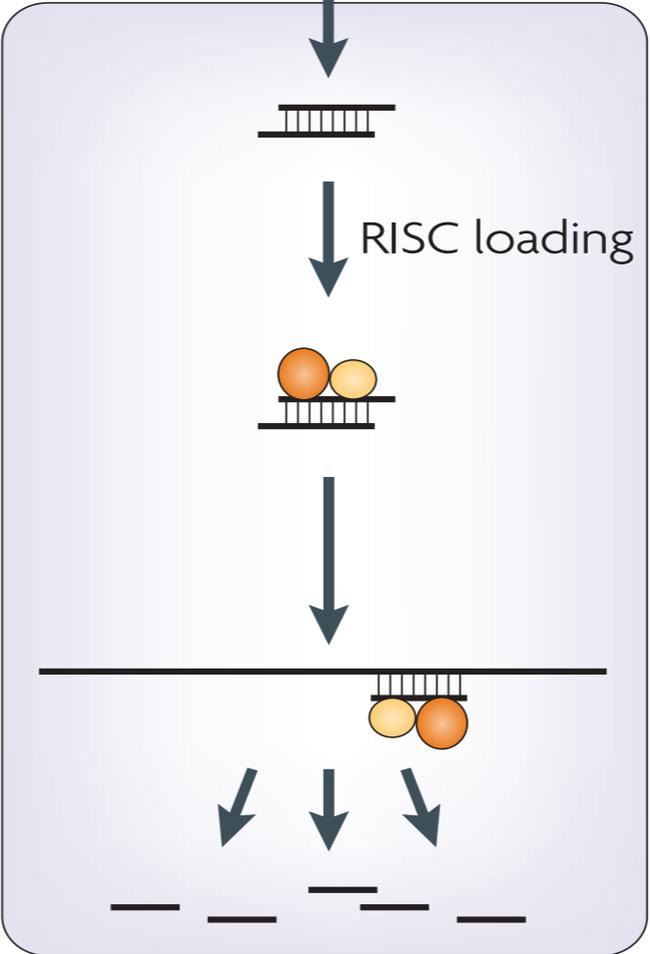
Bathing



Humans

siRNA  
[diagram of siRNA]  
21 bp

Transfection



Genome-wide  
“libraries”

Specificity  
Efficiency  
Reproducibility

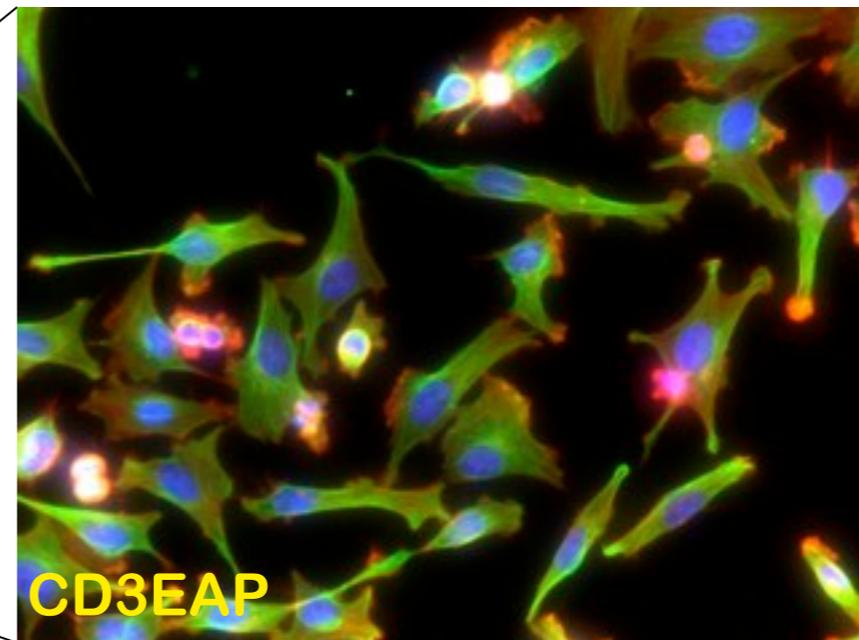
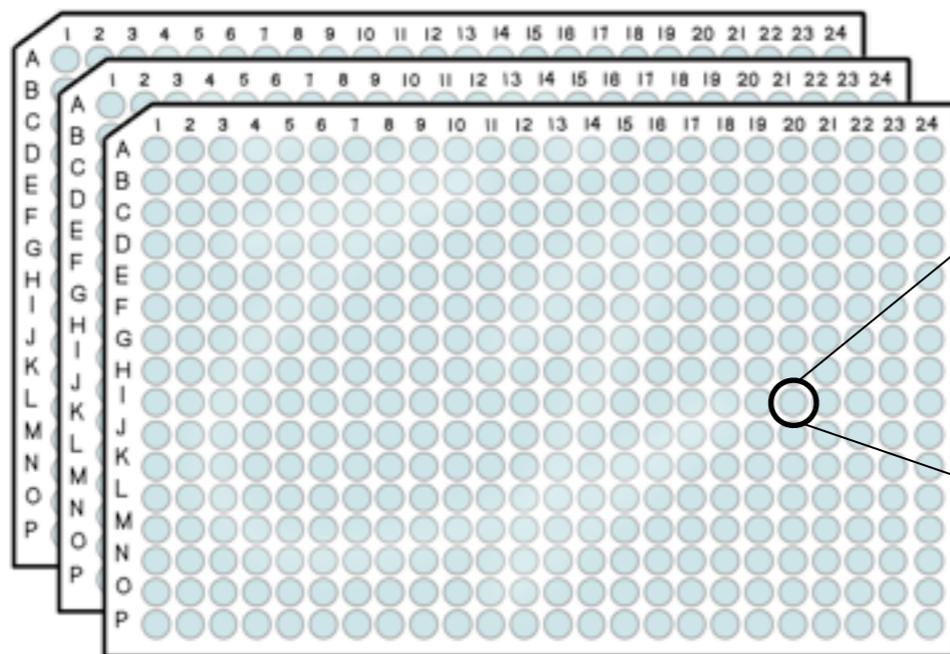
# What do human cells do when you knock down each gene in turn?

with F. Fuchs, C. Budjan, Michael Boutros (DKFZ)

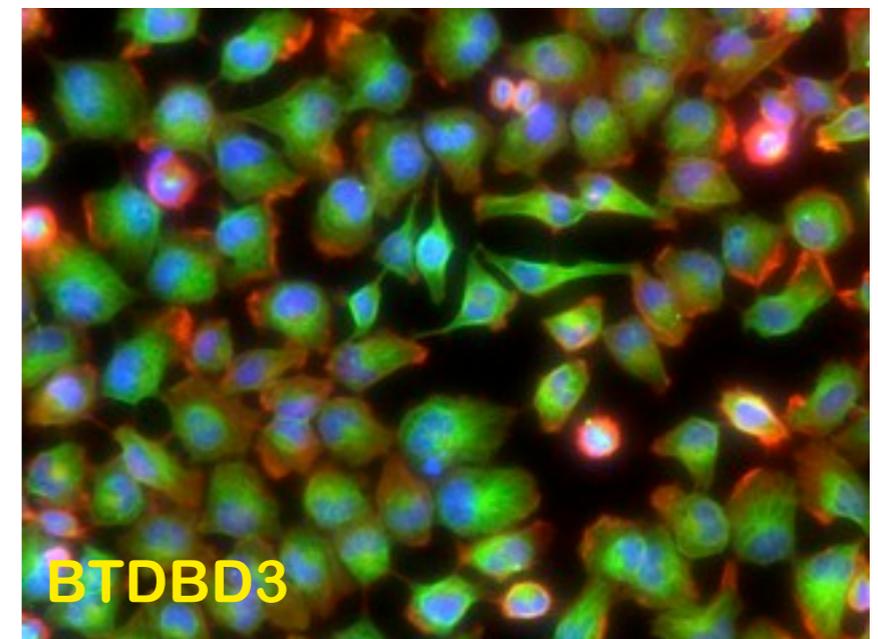
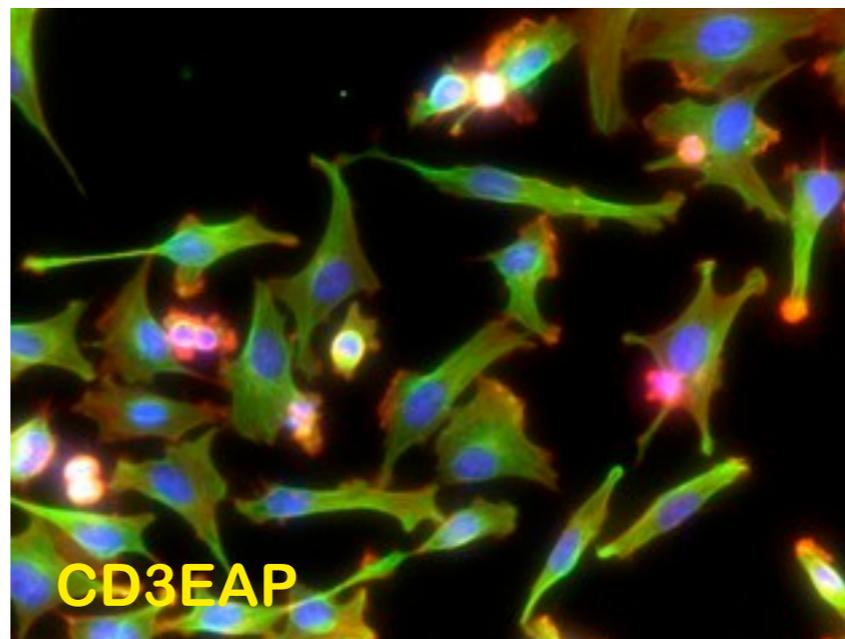
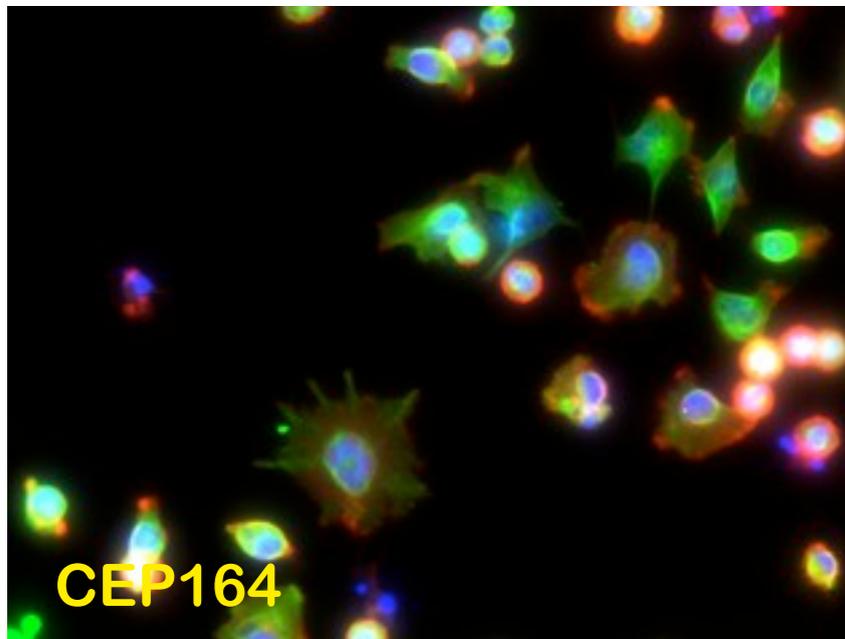
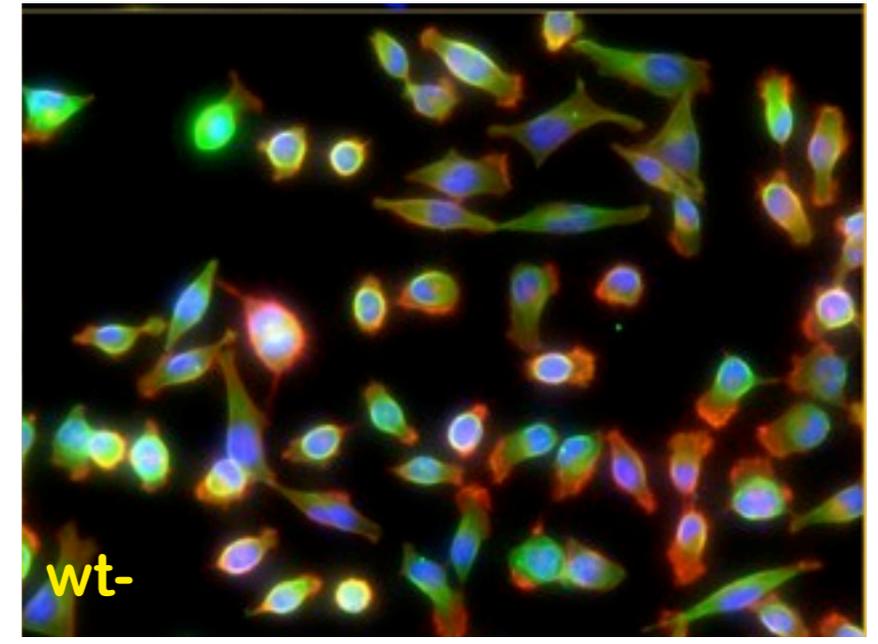
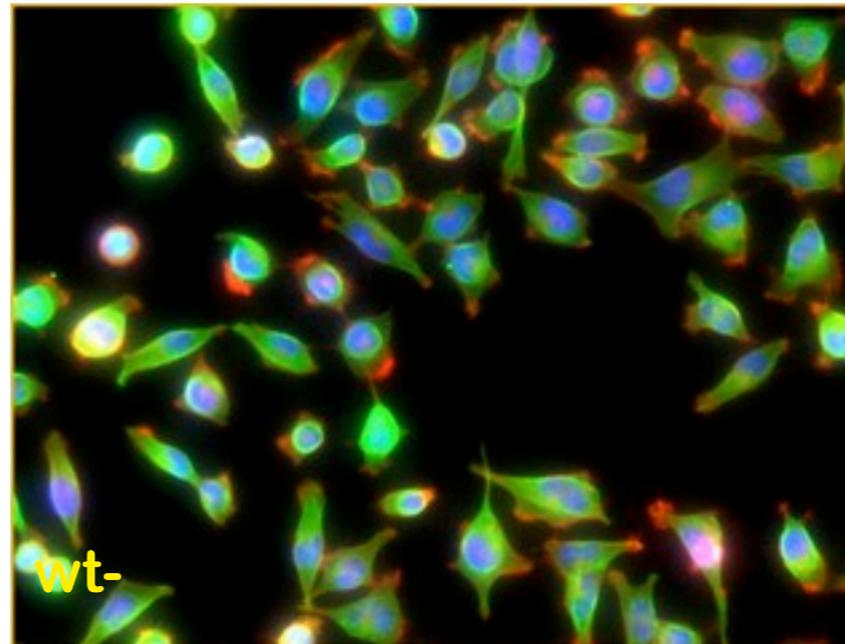
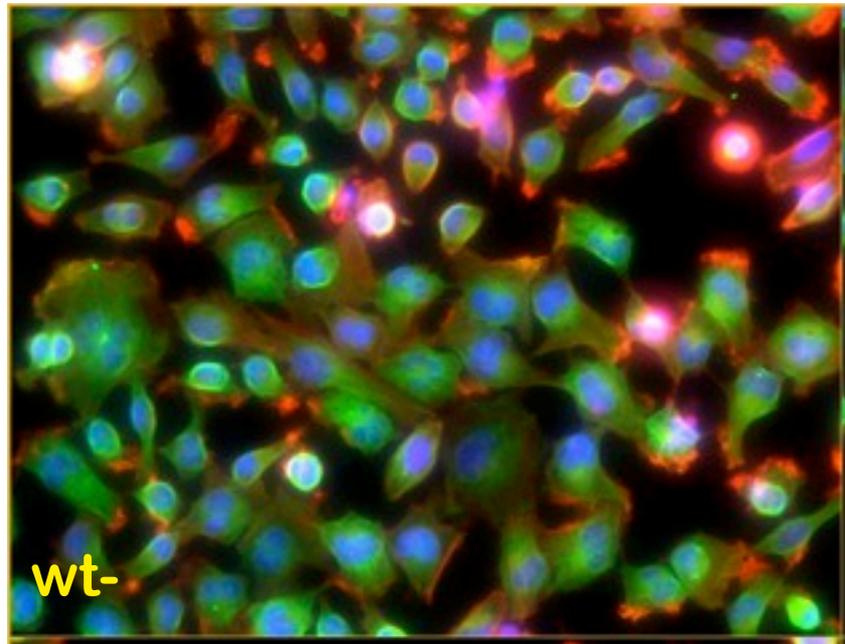
Genomewide RNAi library (Dharmacon, 22k siRNA-pools)

HeLa cells, incubated 48h, then fixed and stained

Microscopy readout: **DNA (DAPI)**, **tubulin (Alexa)**, **actin (TRITC)**



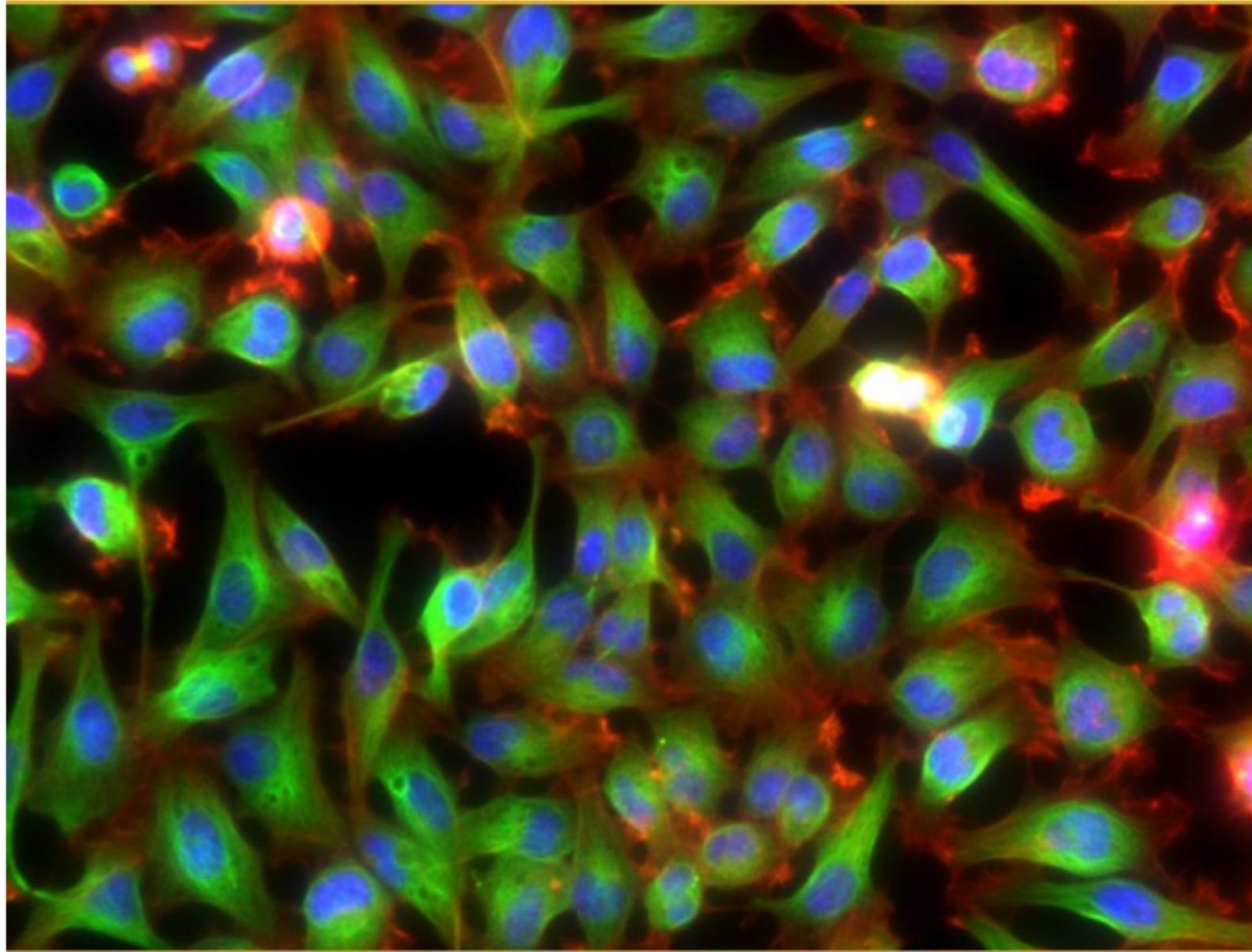
# RNAi perturbation phenotypes are observed by automated microscopy



22839 wells, 4 images per well

each with **DNA**, **tubulin**, **actin** (1344 x 1024 pixel at 3 x 12 bit)

# Segmentation

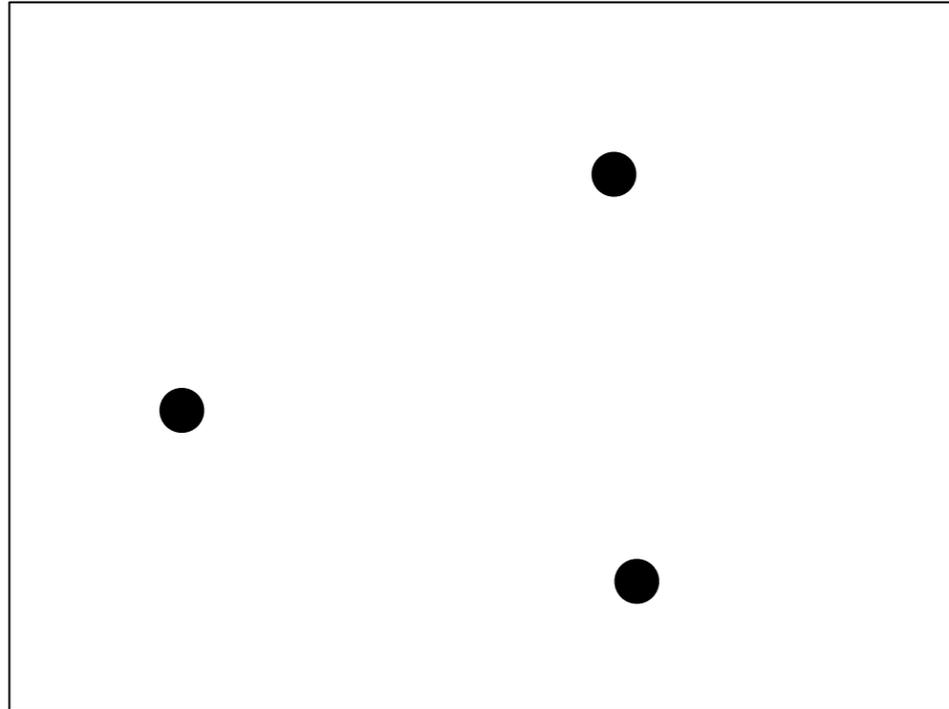


**Nuclei are easy (e.g. locally adaptive threshold)**

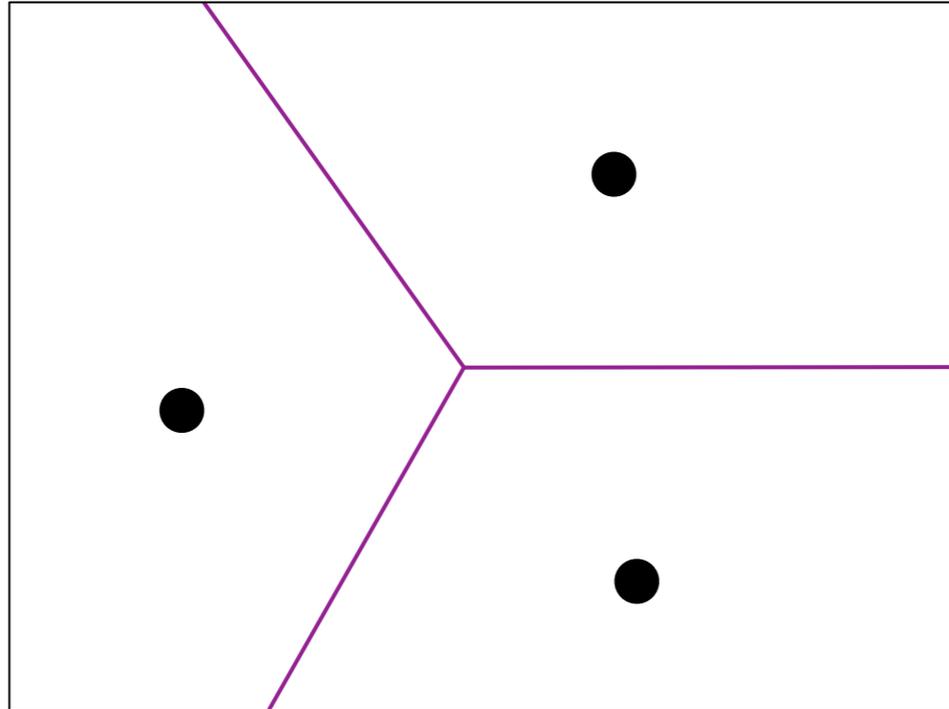
**But cells touch.**

**How do you draw reasonable boundaries between cells?**

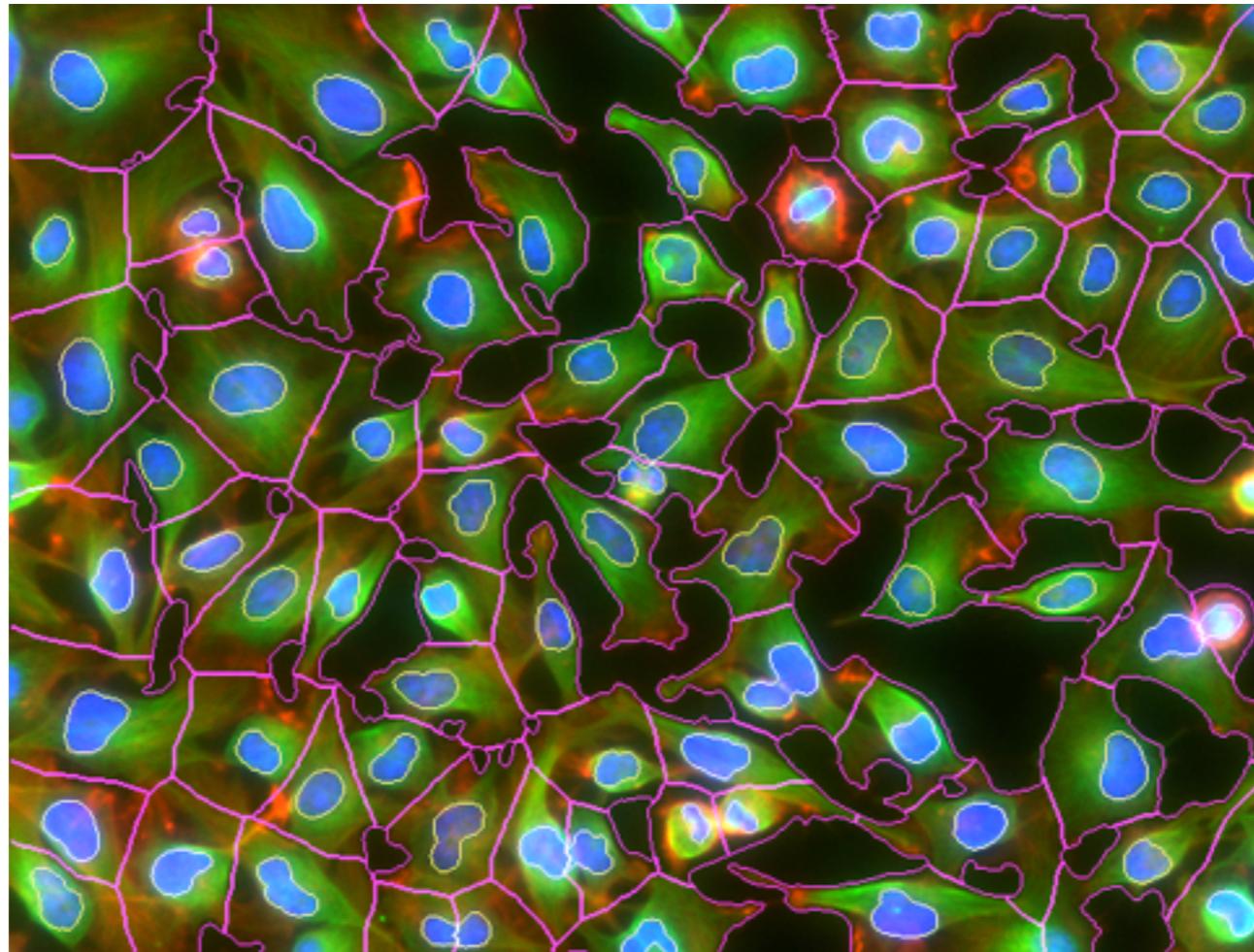
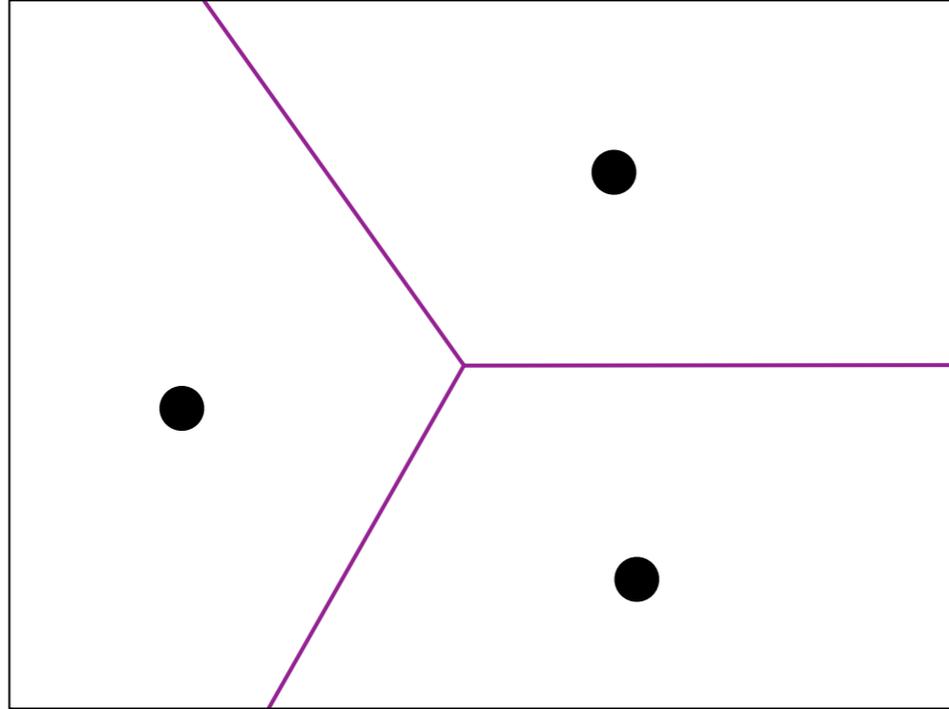
# Voronoi segmentation



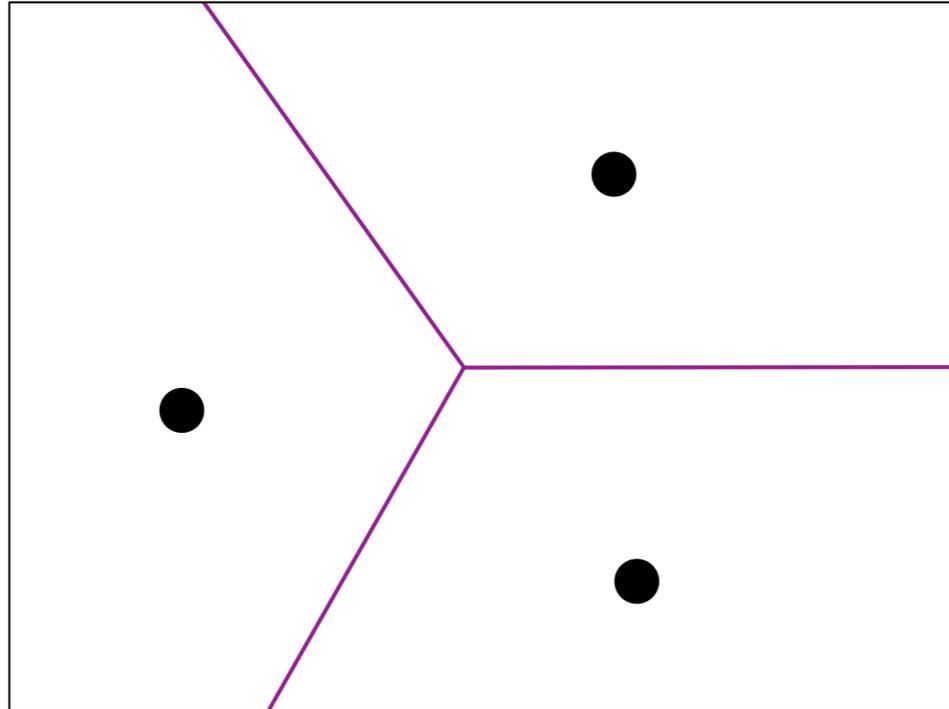
# Voronoi segmentation



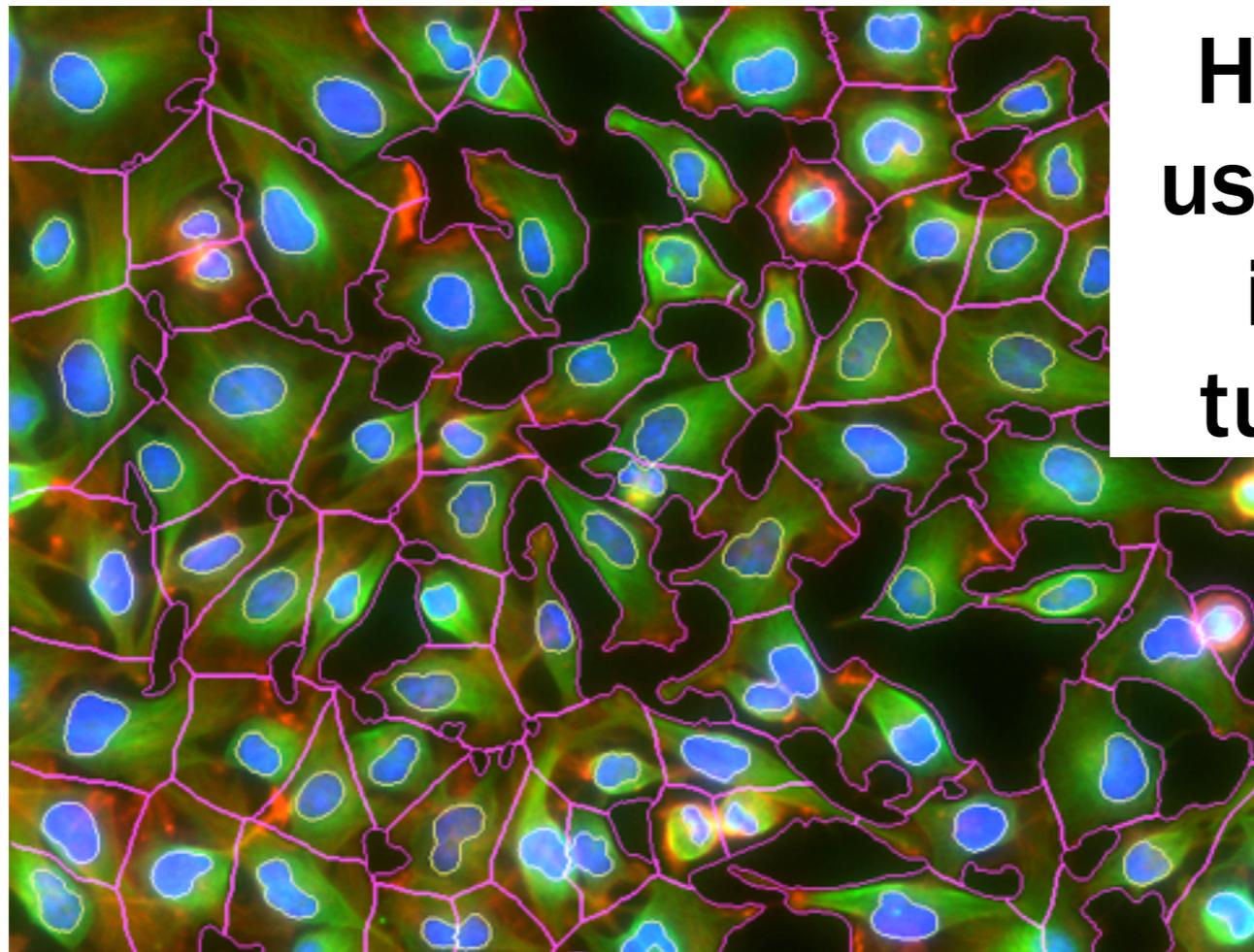
# Voronoi segmentation



# Voronoi segmentation



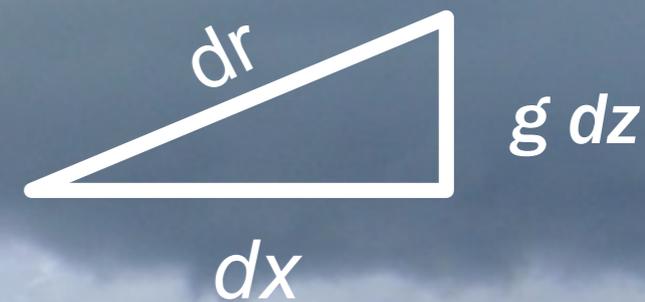
**But we only used the nuclei. The boundaries are artificially straight.**



**How can we better use the information in the actin and tubulin channels?**

Riemann metric on the  
topographic surface  
(*'manifold'*)

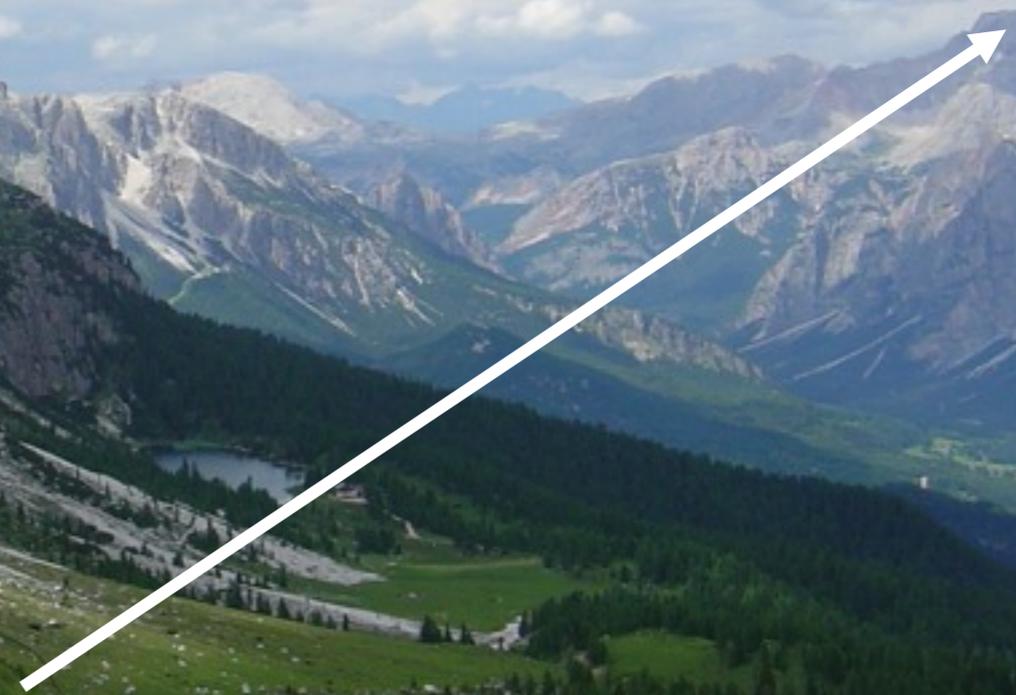
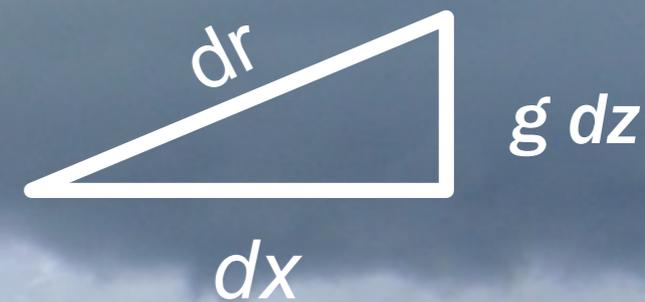
$$dr^2 = dx^2 + dy^2 + g dz^2$$



EBImage::propagate

Riemann metric on the  
topographic surface  
(*'manifold'*)

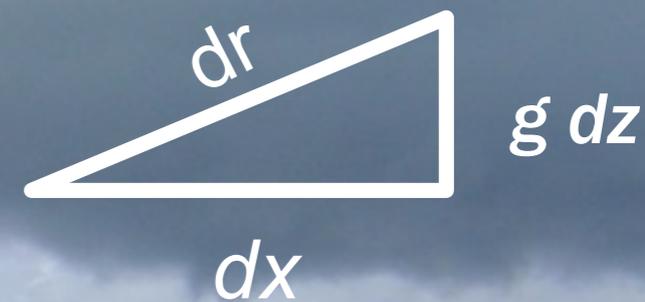
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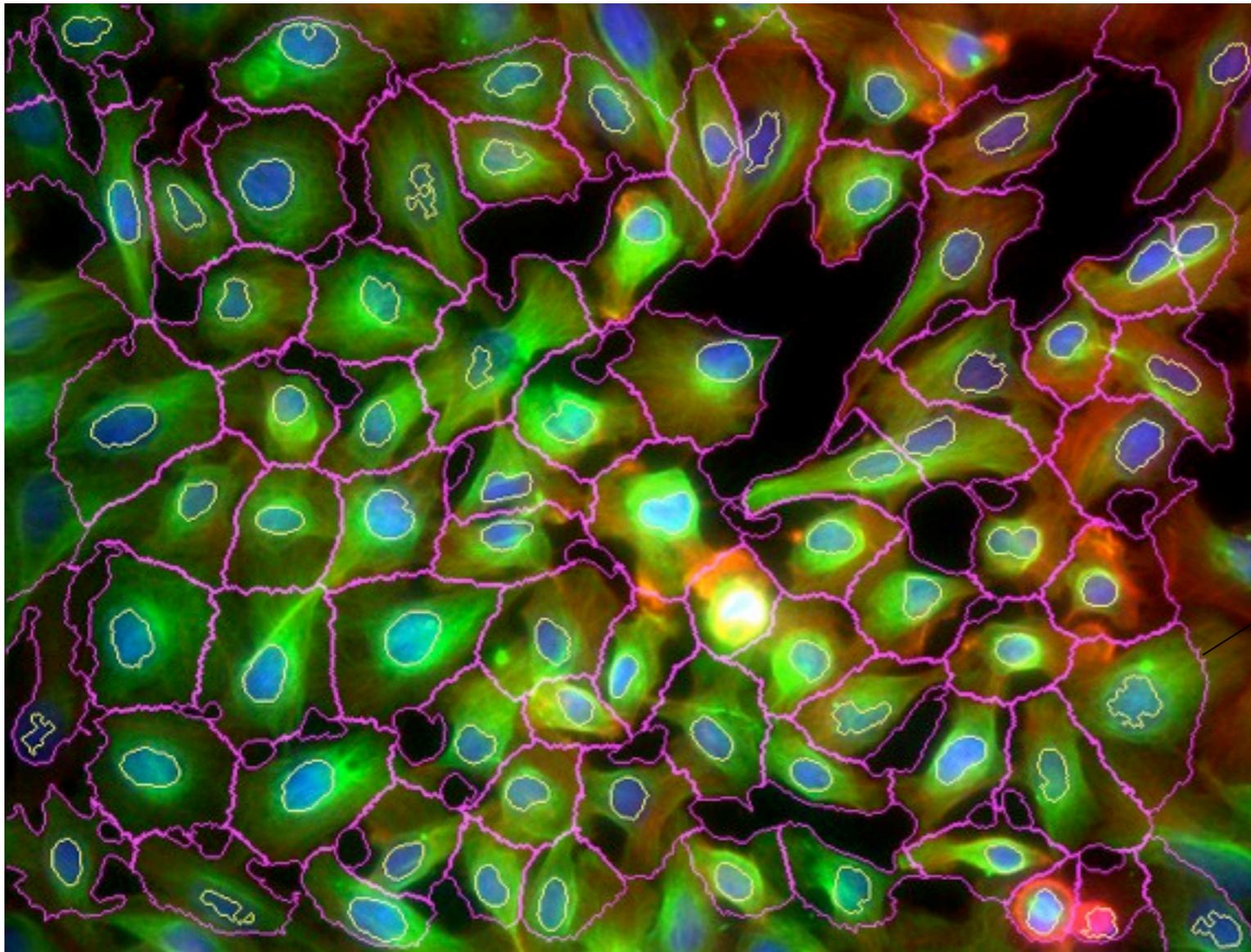
Riemann metric on the  
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$$dr^2 = dx^2 + dy^2 + g dz^2$$



EBImage::propagate

# Converting images into quantitative features



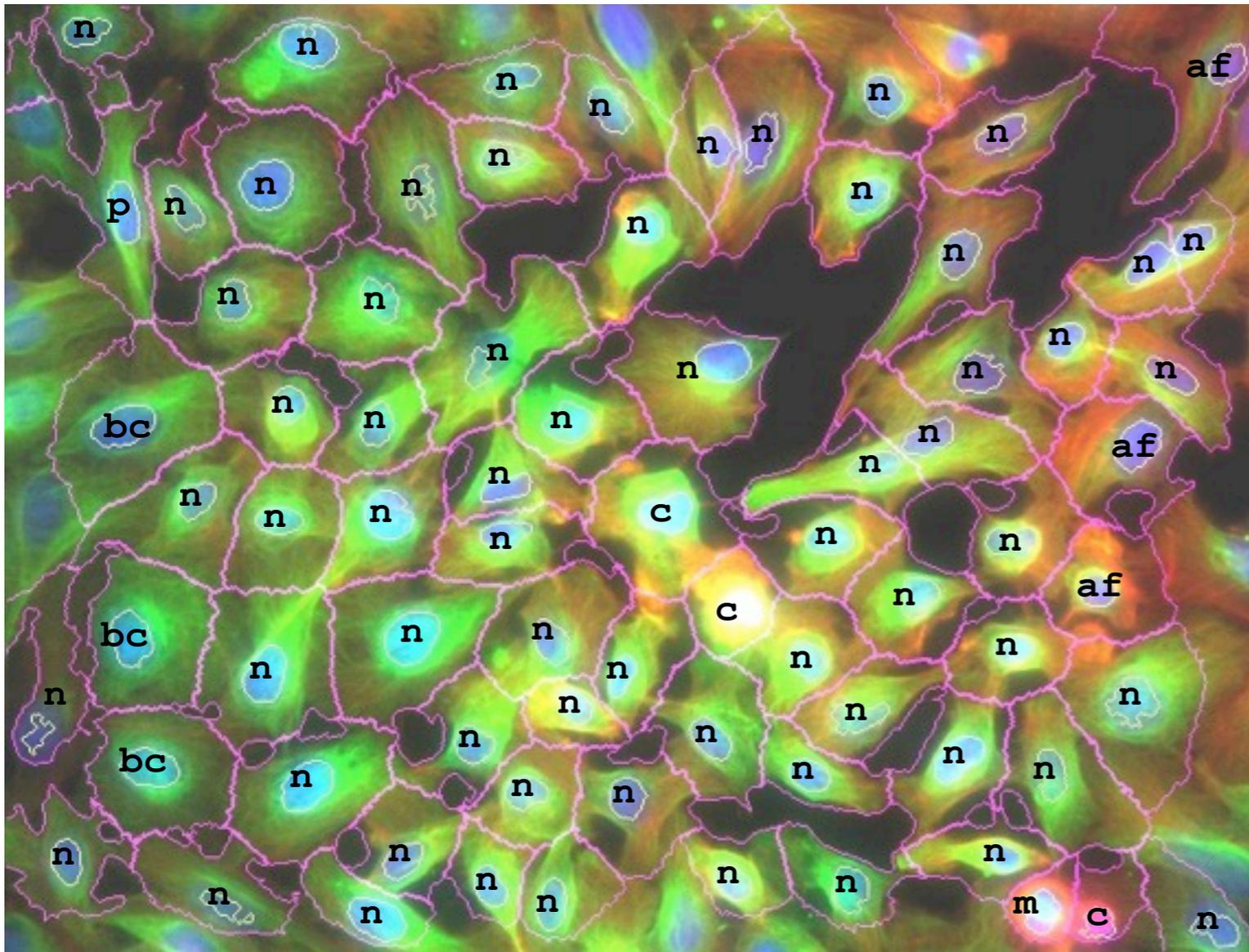
cell size	289
cell intensity	34.33118
eccentricity	0.472934
nucleus size	2857.356
DNA content	485.2710
actin content	0.828876
tubulin content	0.098647
actin F11	0.049594
actin F12	0.081746
actin F21	0.158817
actin F22	0.179339
tubulin F11	0.009249
tubulin F12	0.219697

... ..

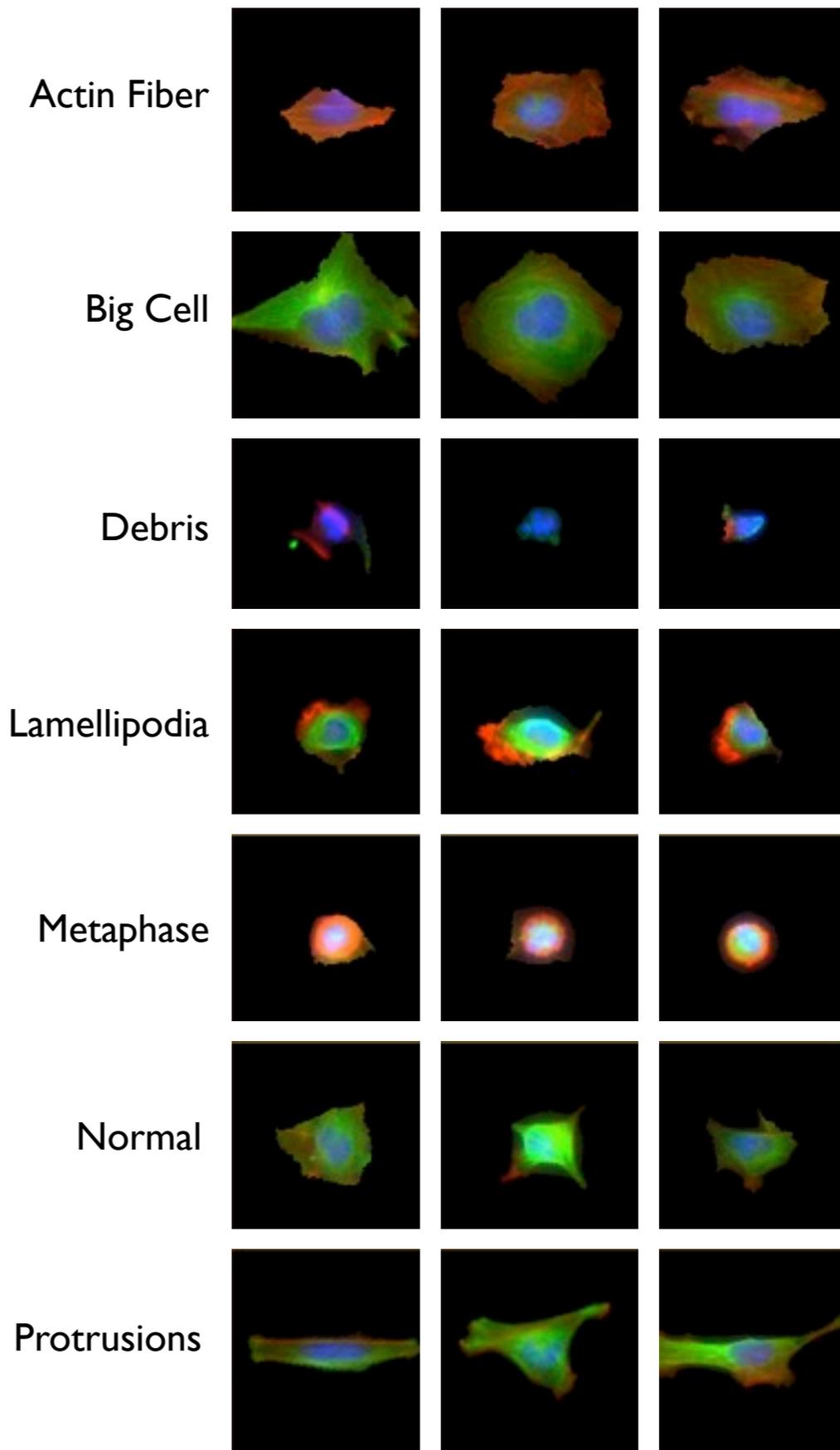
**178 features per cell**

**EBImage::computeFeatures**

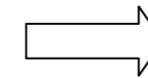
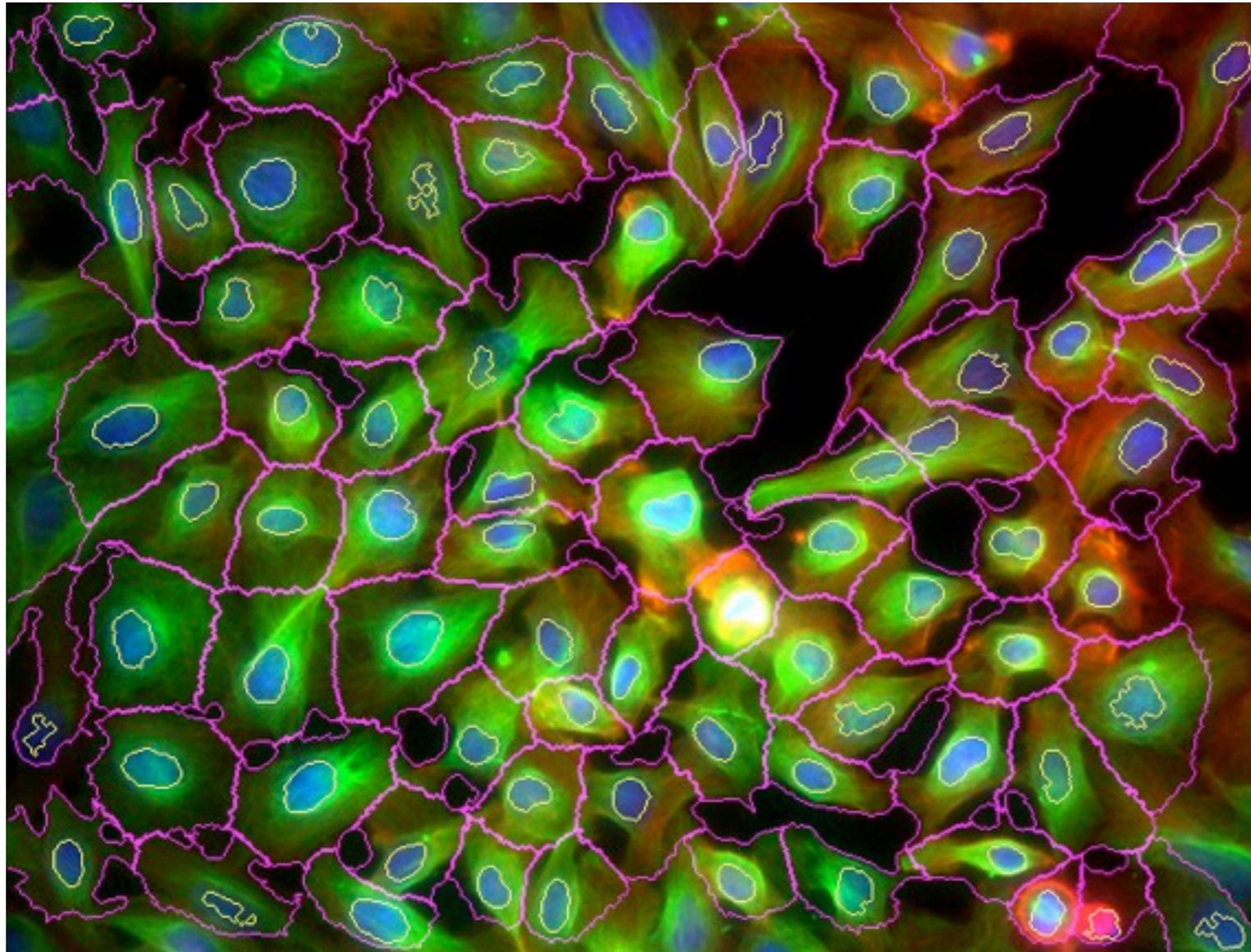
# Cells are classified into predefined classes



178 features per cell  
Radial-kernel SVM  
Manually annotated training set of ~3000 cells  
Accuracy: ~ 90 %



# The image is now represented by a 13-dim vector: “phenotypic profile”



n	289
ext	34.33118
ecc	0.472934
Next	2857.356
Nint	485.2710
AtoTint	0.828876
NtoATsz	0.098647
AF %	0.049594
BC %	0.081746
C %	0.158817
M %	0.179339
LA %	0.009249
P %	0.219697

**How do you measure  
distance and similarity  
in a 13-dimensional phenotypic  
profile space?**

# Similarity depends on the choice and weighting of descriptors



# Distance metric learning

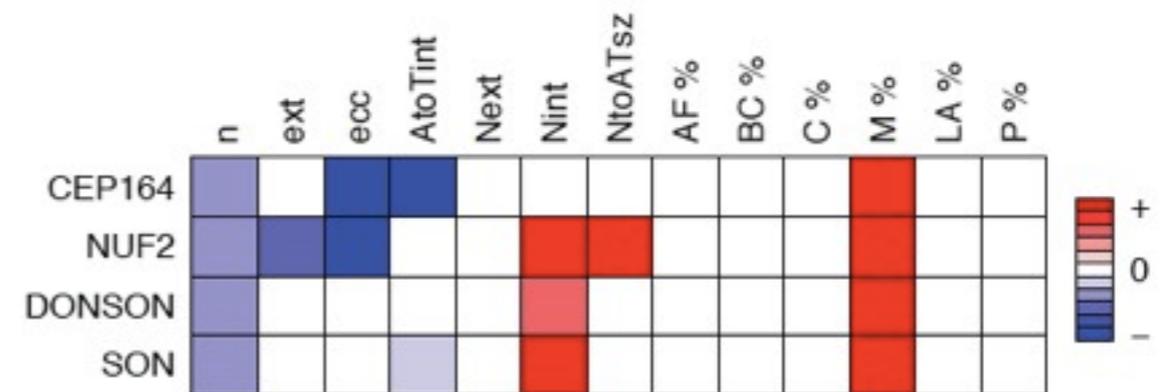
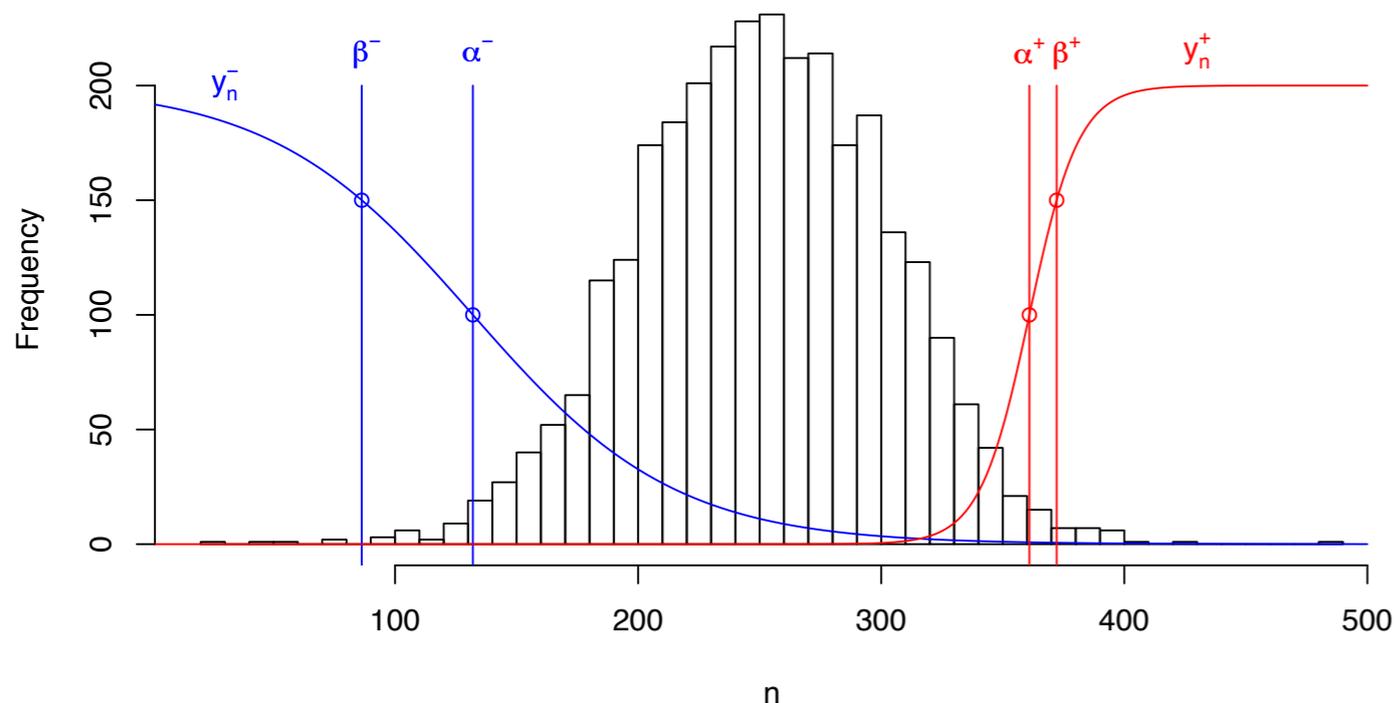
$$d(x, y) = \sum_k |f_k(x_k) - f_k(y_k)|$$

$$f_k(x) = \frac{1}{1 + \exp(-\eta_k(x - \alpha_k))}$$

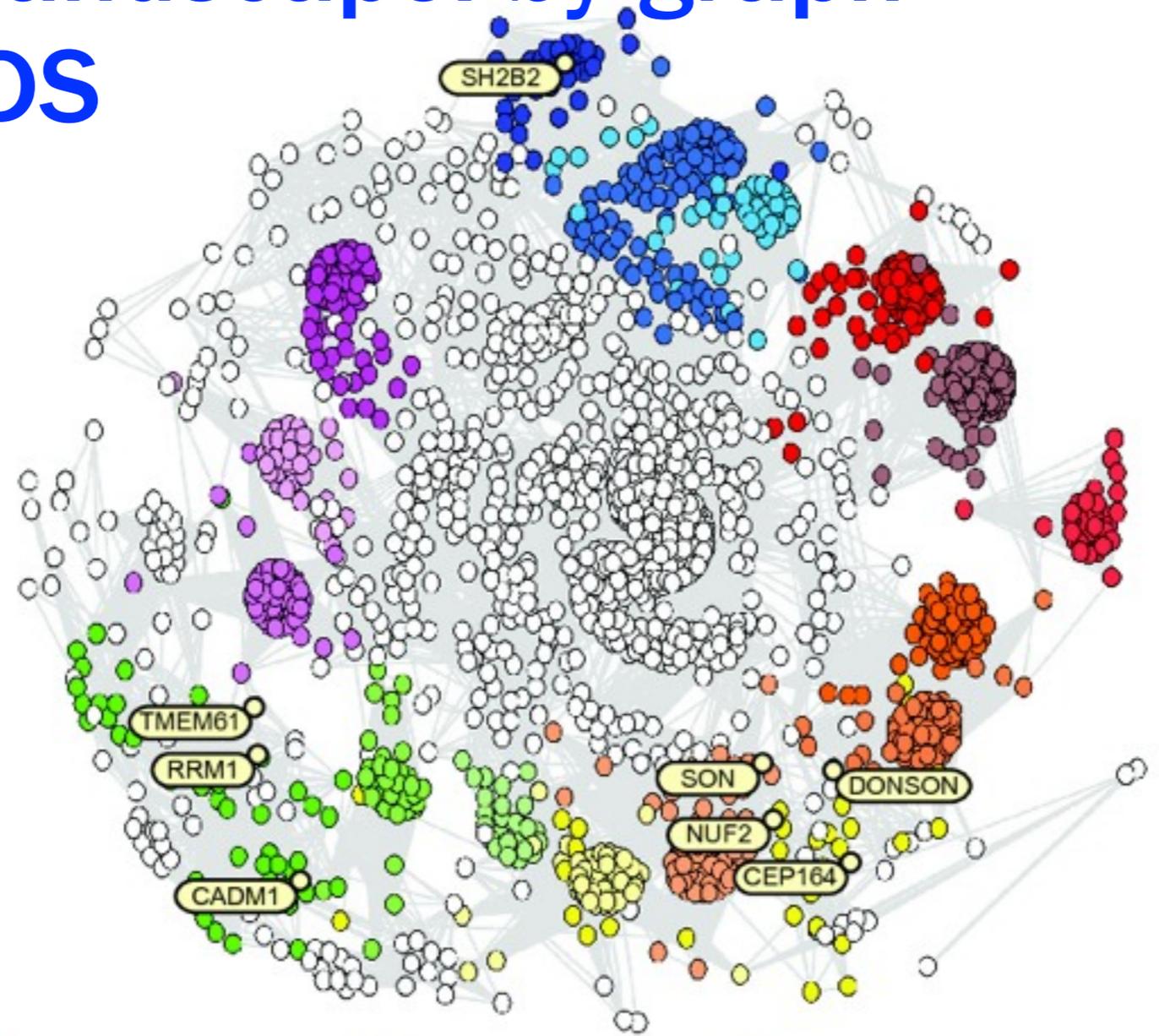
x =

n	289
ext	34.33118
ecc	0.472934
Next	2857.356
Nint	485.2710
a2i	0.828876
Next2	0.098647
AF %	0.049594
BC %	0.081746
C %	0.158817
M %	0.179339
LA %	0.009249
P %	0.219697

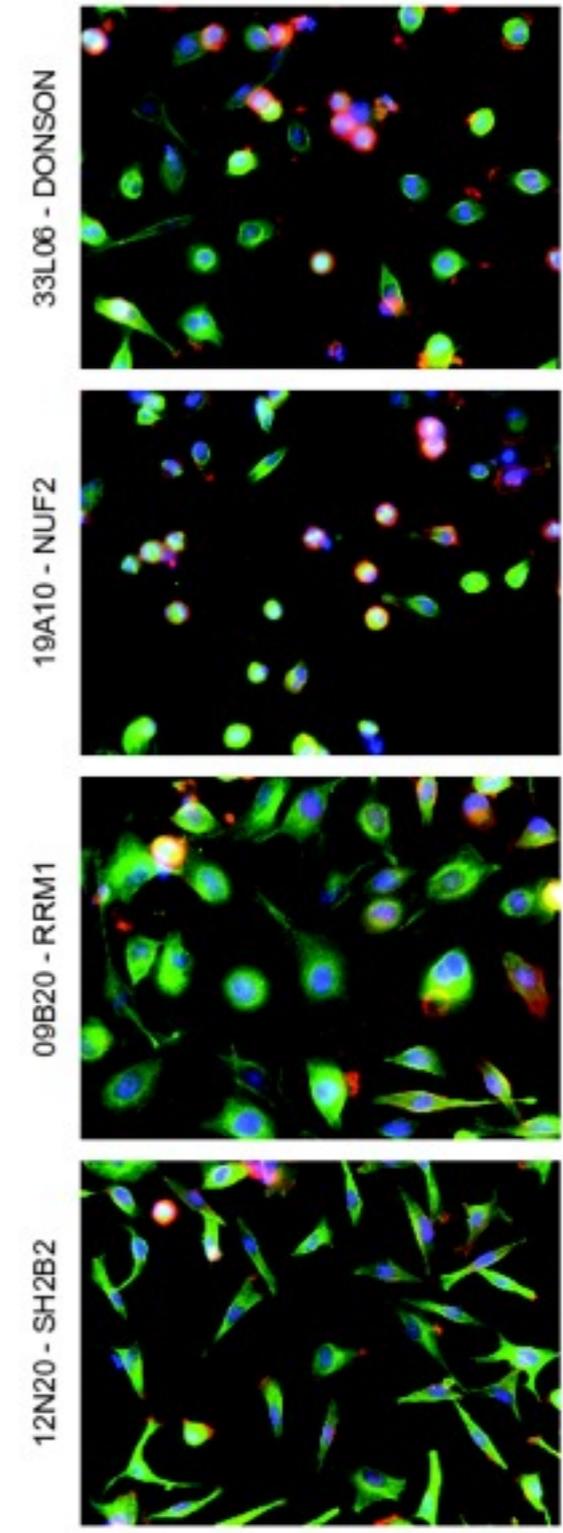
**Training set: pairs of genes that are somehow ‘related’: EMBL STRING**  
**Get  $(\eta, \alpha)$  by minimizing average distance between training set genes, keeping average distance of all genes fixed.**



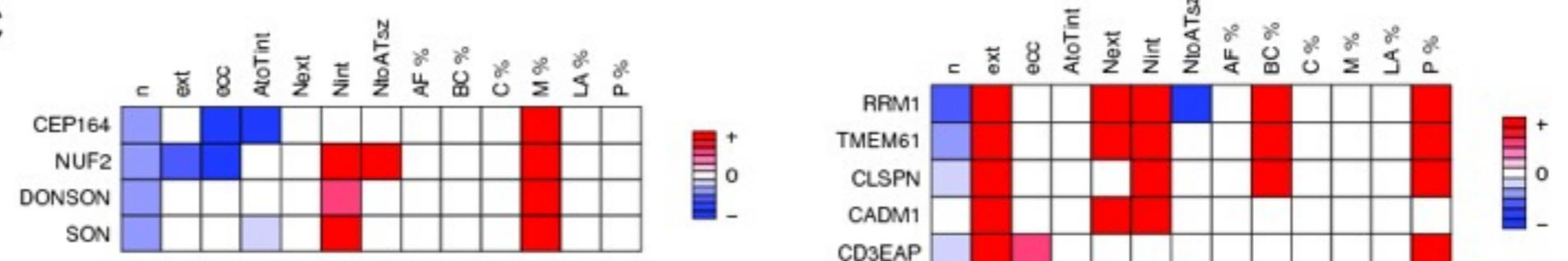
# Phenotype landscape: by graph layout or MDS



- BL phenotype
- SM phenotype
- Actin fiber cells
- Bright nuclei
- Small cells
- Big cells
- Large nuclei
- Low eccentricity cells
- Large cells
- Cells with protrusions
- High actin ratio cells
- Lamellipodia cells
- Elongated cells
- Metaphase cells
- Lamell. + high actin ratio cells
- Elong. cells with protrusions
- Other phenotype
- Proliferating cells



C



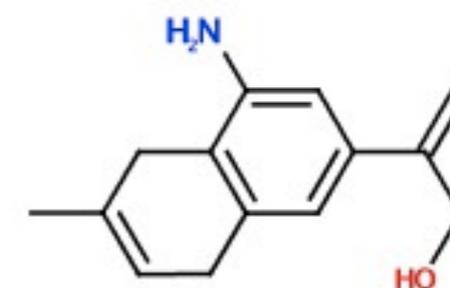
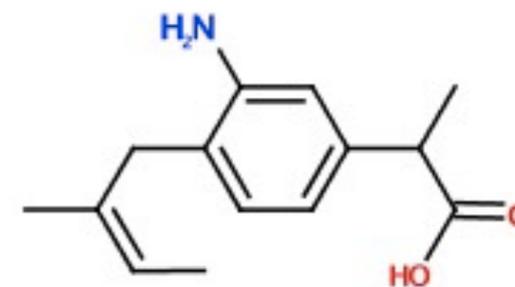
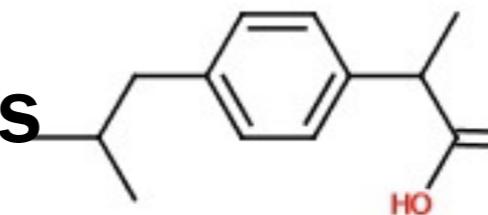
# Summary

**Automated phenotyping of cells upon genetic perturbations by microscopy and image analysis**

**Segmentation, feature extraction, classification, distance metric learning, multi-dimensional scaling, clustering.**

**“Phenotypic map” is useful to biologists**

**Method is also being applied to drugs**



# Collaboration with Michael Boutros, German Cancer Research Centre (Heidelberg)

Fuchs, Pau et al., Molecular Systems Biology (2010)

All data and software available at

<http://www.cellmorph.org>

packages EBImage and imageHTS

Gregoire Pau





**Focus on the analysis of genomic data**

**Based on R and CRAN**

**Six-monthly release cycle, in sync with R**

**Releases:**

- **1.0 in March 2003 (15 packages), ...,**
- **2.8 in April 2011 (466 software packages)**



## *What's the added value?*

Complex **data containers** (S4 classes) for new experimental technologies (microarrays, sequencing) shared between packages - even from different authors.

**metadata** packages: gene annotation, pathways, genomes

**experiment data** packages: landmark datasets

stronger emphasis on **vignette**-style documentation

stricter submission review (much more could be done)

more package interdependence → **releases**

**training courses**

**mailing list** is amenable to software and domain (bio) questions

Push **new technologies**: S4, vignettes, string handling, computations with ranges, out-of-RAM objects

# Interactive Reports

## Distinguish

- **interactive exploration by data analyst**
- **reports (presentation graphics)**

# Interactive Reports

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- interactive exploration by data analyst
- reports (presentation graphics)

Everybody has a PDF reader.

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**Everybody has a PDF reader.**

**Everybody has a web browser.**

# Interactive Reports

## Distinguish

- interactive exploration by data analyst
- reports (presentation graphics)

**Everybody has a PDF reader.**

**Everybody has a web browser.**

**Web browsers are turning into an operating system.**

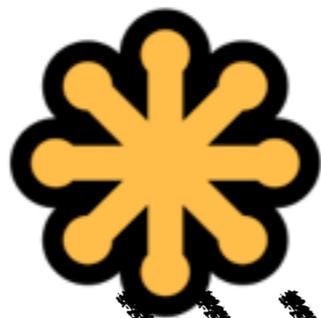


PDF viewer



LATEX





**HTML**



**PDF viewer**



**LATEX**



# arrayQualityMetrics

**Reports on Quality of Microarray Datasets**

**effort to collect all extant, useful quality metrics for microarrays**

**funding by EU FP7 and by Genentech**

**used by public databases (EBI::ArrayExpress) to annotate their data**

**offerings for users**

**Example report**

# arrayQualityMetrics

## Reports

effort to  
funding  
used by  
offerings

- **mouseover** → **tooltip** (rendered as an **HTML table** next to the plot)
- **click** → **select & highlight** (propagated to several plots, tables)
- **expand, collapse sections**
- **use HTML elements (checkboxes) to control plots**

# Comments and outlook

SVG is part of HTML 5:

- linked plots and brushing
- HTML widgets as controllers (checkboxes, wheels)

SVG/HTML post-processing via the XML package

Callback processing currently in JavaScript.

Use R? On server: googleVis talk by Markus Gesmann, Diego de Castillo; locally: browser plugin

Duncan Temple Lang's [SVGAnnotation](#) package: works for any R graphic (incl. base), but depends on undocumented / changeable behavior of cairo.

Paul Murrell's [gridSVG](#) package: cleaner and more durable approach, based on grid graphics.

# Generalisation?

`arrayQualityMetrics` is for microarrays

Software sees:

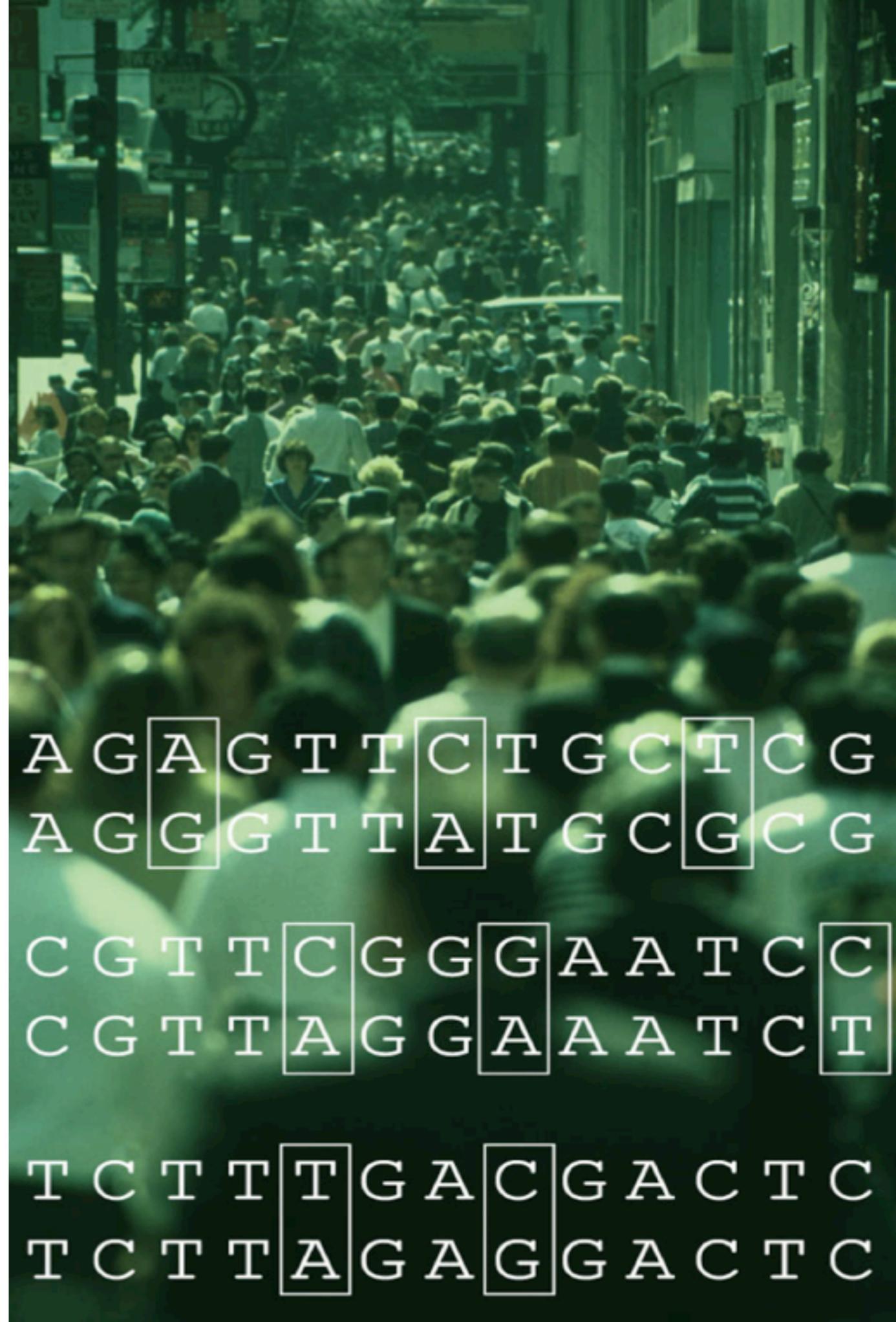
- a set of **items** (arrays)
- a set of **modules** that compute the sections of the report (PCA, boxplots, scatterplots)

This could be generalised to reports on very different types of subject matter - I will be happy to discuss this.

# What makes us different?

From Genome Wide  
Association Studies, ~400  
variants that contribute to  
common traits and diseases  
are known

Individual and the cumulative  
effects are disappointingly  
small



A G A G T T C T G C T C G  
A G G G T T A T G C G C G  
C G T T C G G G A A T C C  
C G T T A G G A A A T C T  
T C T T T G A C G A C T C  
T C T T A G A G G A C T C

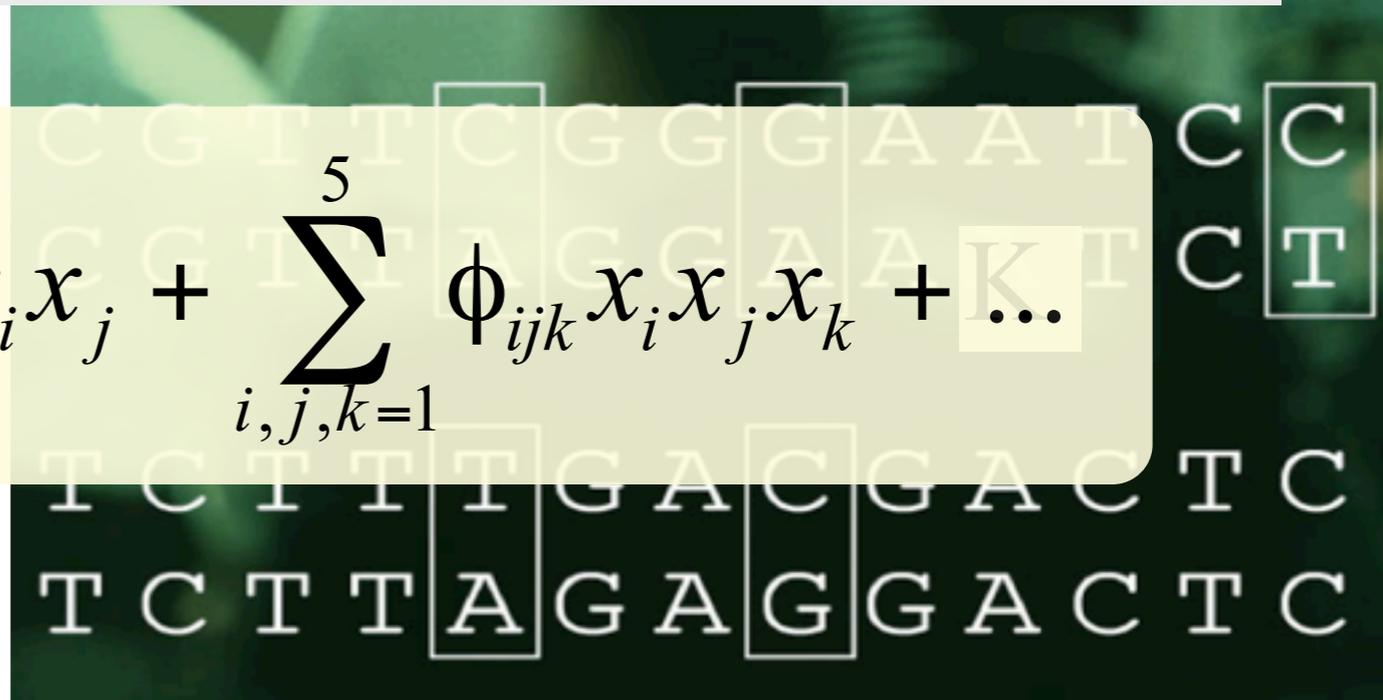
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Individual and the cumulative  
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small

Epistasis, interactions

$$\phi = \phi_0 + \sum_{i=1}^5 \phi_i x_i + \sum_{i,j=1}^5 \phi_{ij} x_i x_j + \sum_{i,j,k=1}^5 \phi_{ijk} x_i x_j x_k + \dots$$

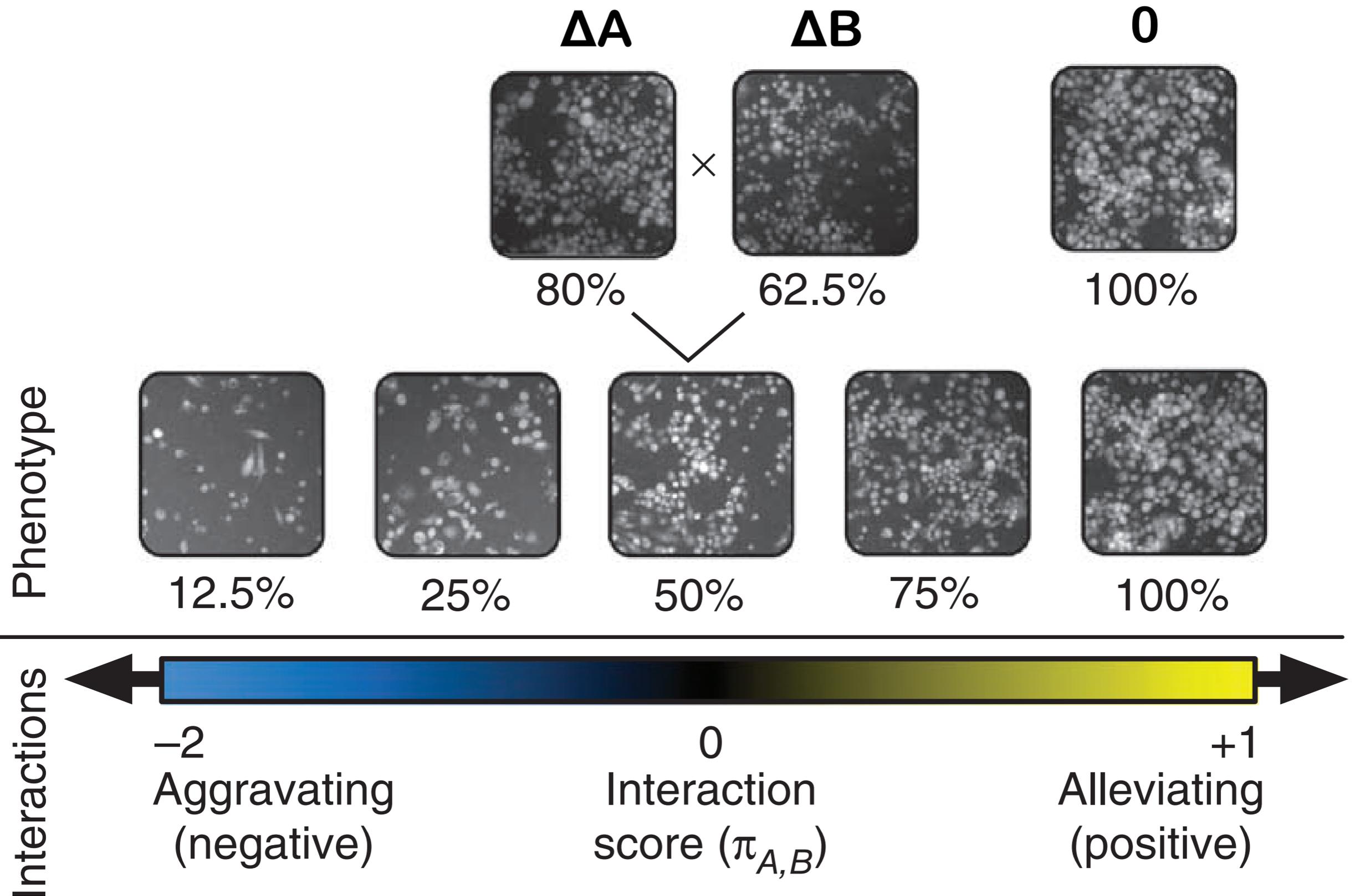


# Take a step back...

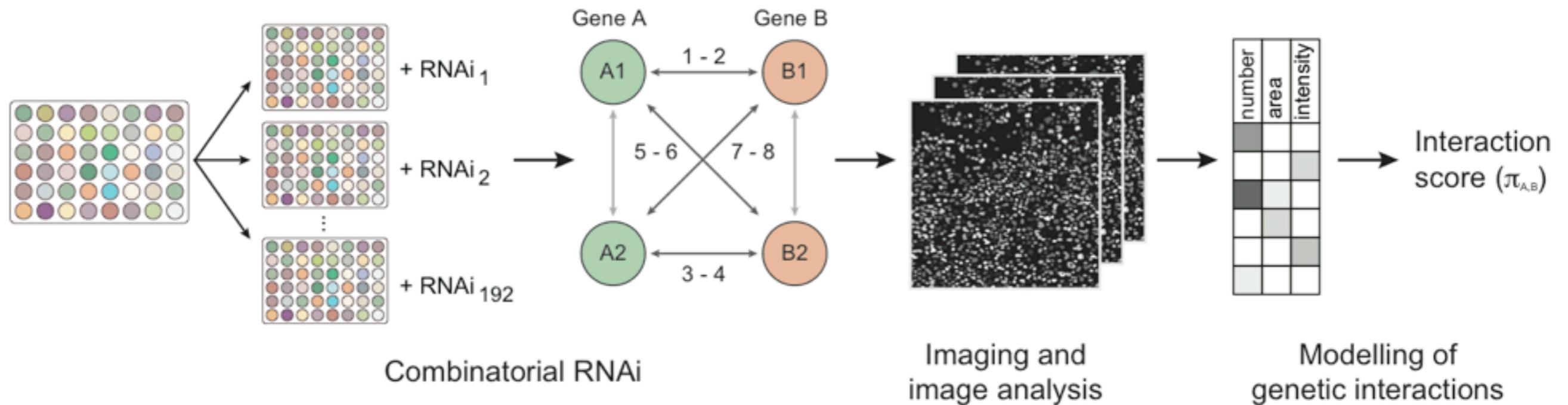
## Genetic interactions

- **only pairwise**
- **for a simple phenotype**
- **in a simple model system**

# Simplest “model system”: pairwise gene knock-down interactions and a scalar phenotype



# A combinatorial RNAi screen



- 93 Dm kinases and phosphatases
- Each targeted by two independent dsRNA designs
- Validation of knock-down by qPCR
- 96 plates (~37.000 wells)
- 4.600 distinct gene pairs

with Bernd Fischer (EMBL) and  
M. Boutros, Thomas Sandmann,  
Thomas Horn (DKFZ )

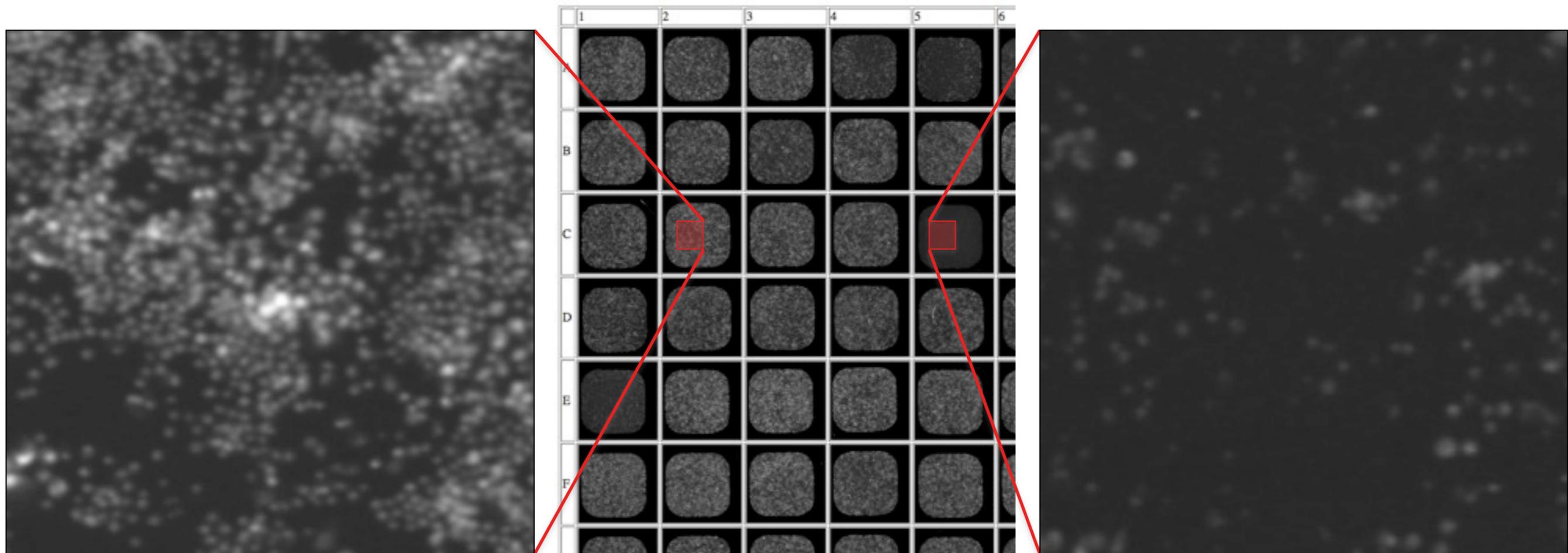
Nature Methods 4/2011



# Image analysis and feature extraction

(version of 2010)

- number of cells
- DAPI intensity for each cell
- DAPI area for each cell



# Modelling Genetic Interactions

For many phenotypes, the perturbation effects combine multiplicatively for non-interacting genes  $i, j$ :

$$d_{ij} = \omega \mu_i \mu_j$$

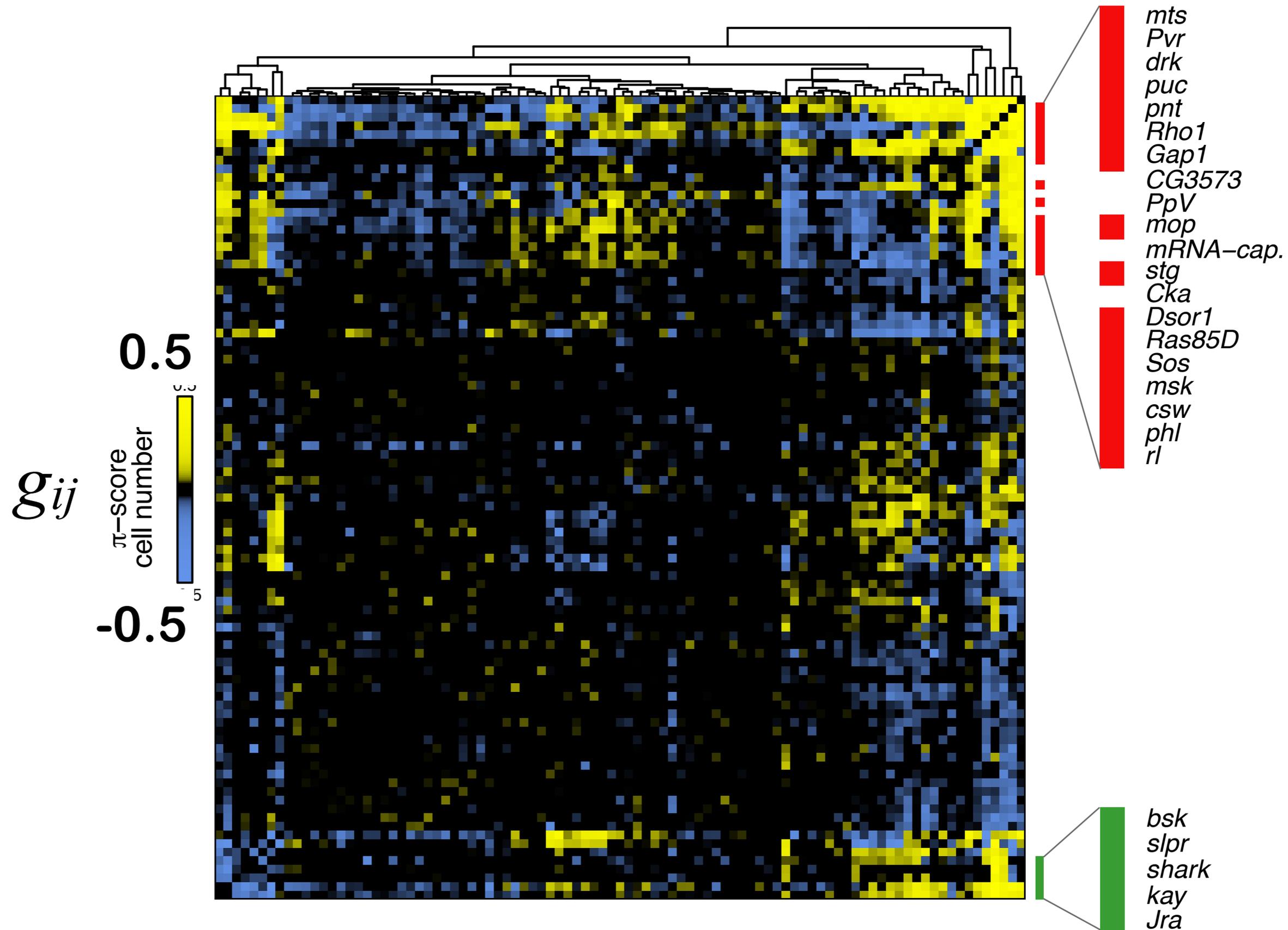
... i.e. additive on a logarithmic scale

$$\log d_{ijk} = w + m_i + m'_j + g_{ij} + \varepsilon_{ijk}$$

The diagram illustrates the additive components of the logarithmic model. Green arrows point from the following labels to their corresponding terms in the equation above:

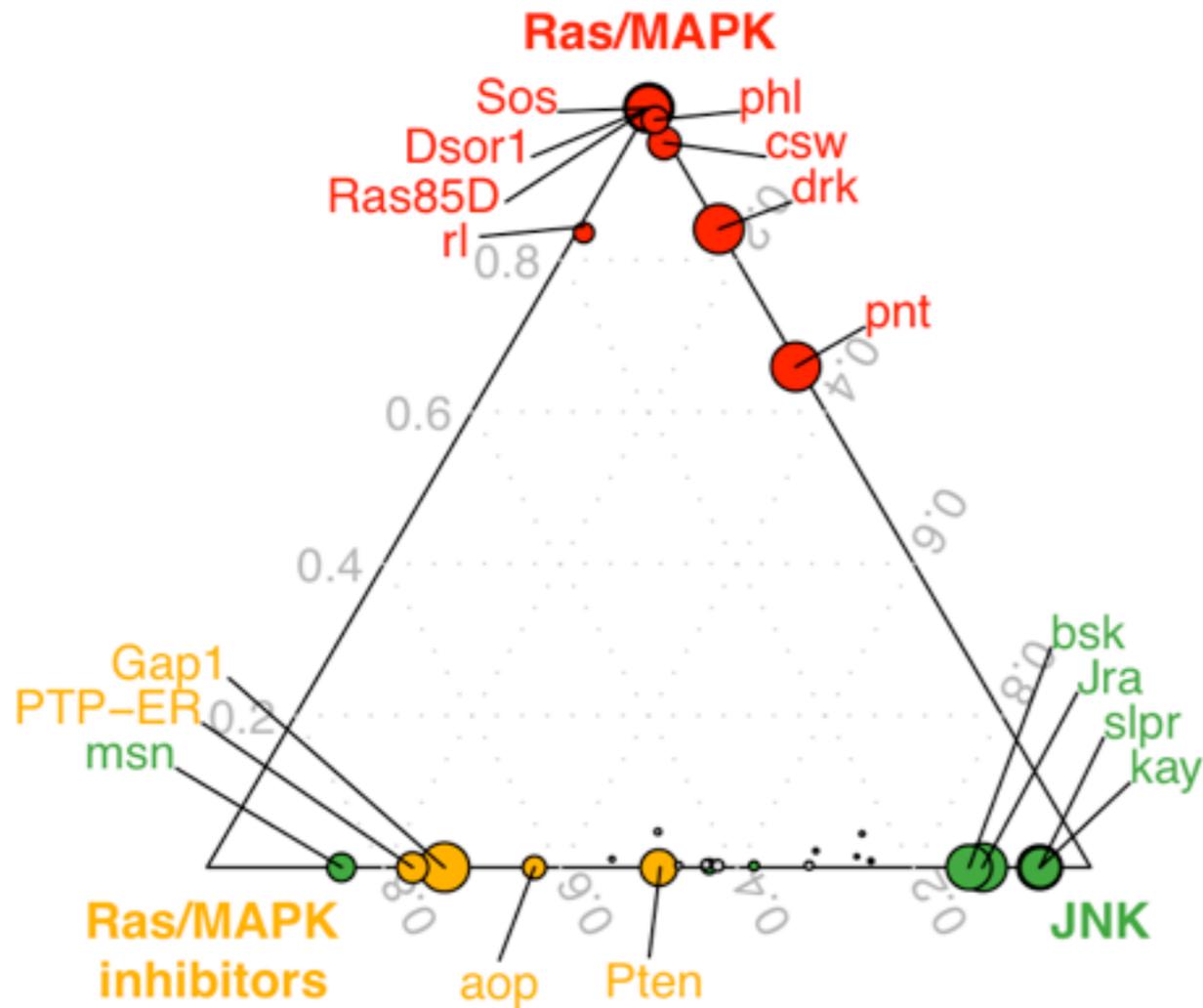
- measurement (nr cells, growth rate, ...)** points to  $\log d_{ijk}$
- baseline** points to  $w$
- main effect of dsRNA  $i$**  points to  $m_i$
- main effect of dsRNA  $j$**  points to  $m'_j$
- interaction** points to  $g_{ij}$
- measurement error** points to  $\varepsilon_{ijk}$

Thus we get a matrix of interaction parameters:  
profile clustering reflects functional modules

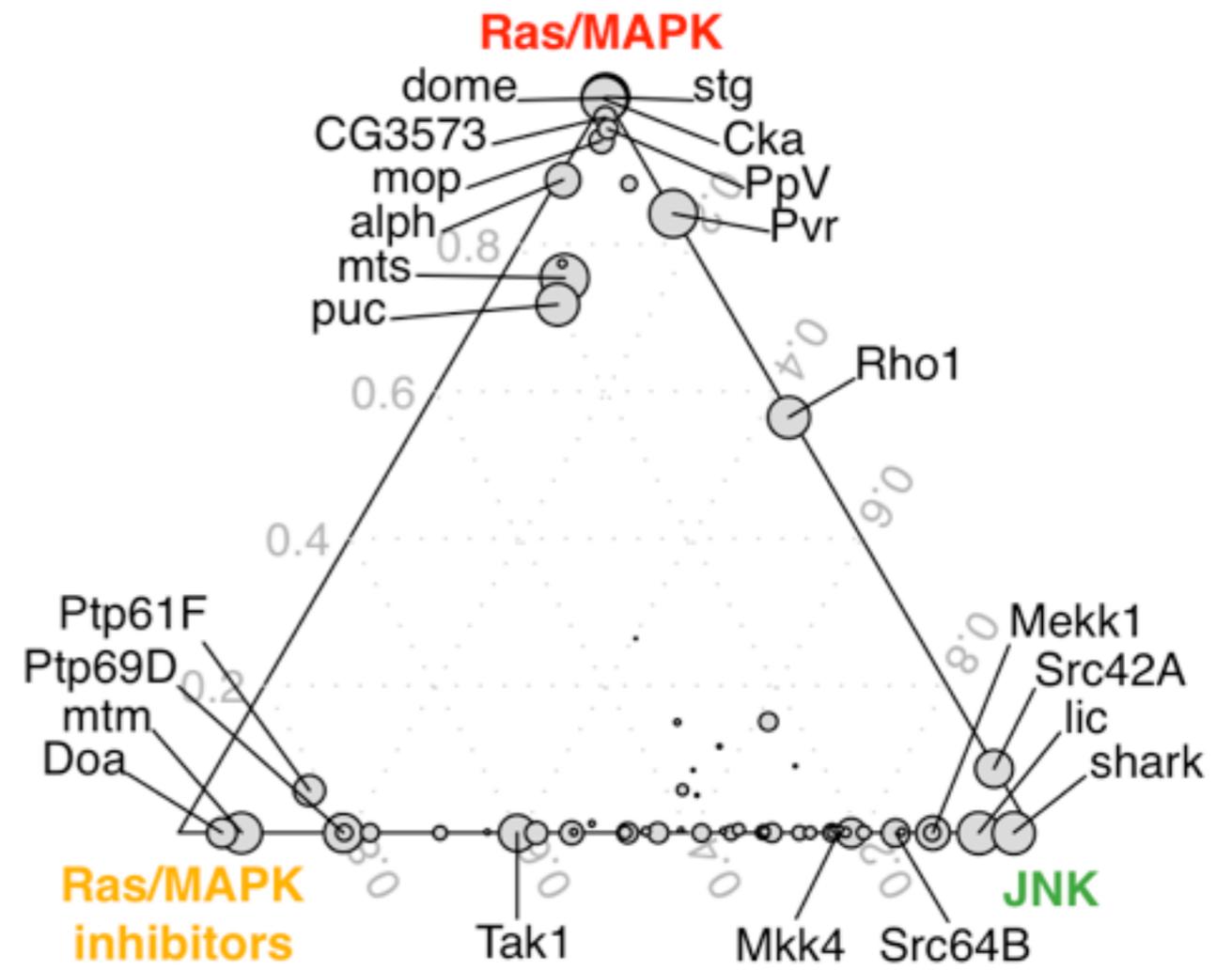


# Classification of genes by function through their interaction profiles

cross-validated performance on training set



functional prediction applied to new genes

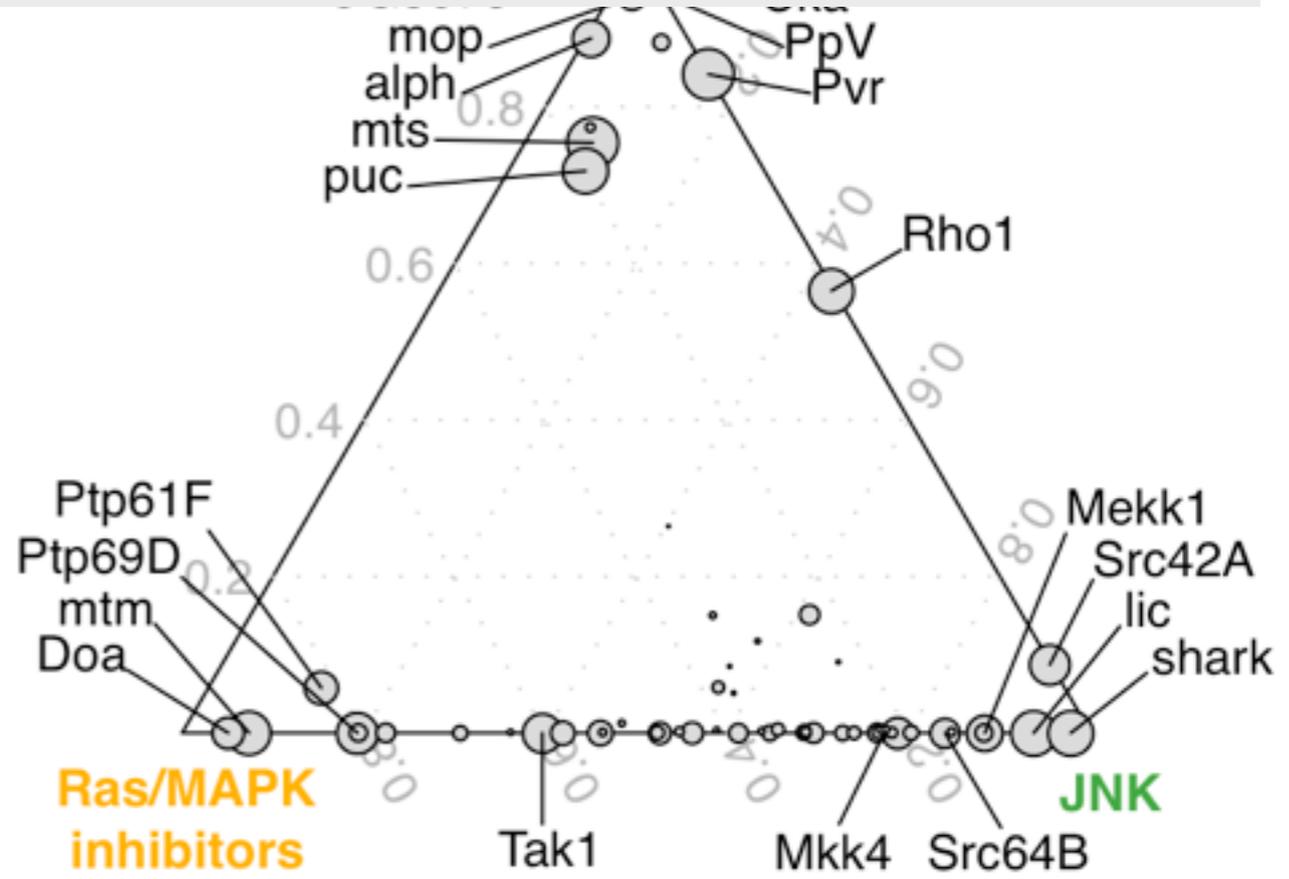
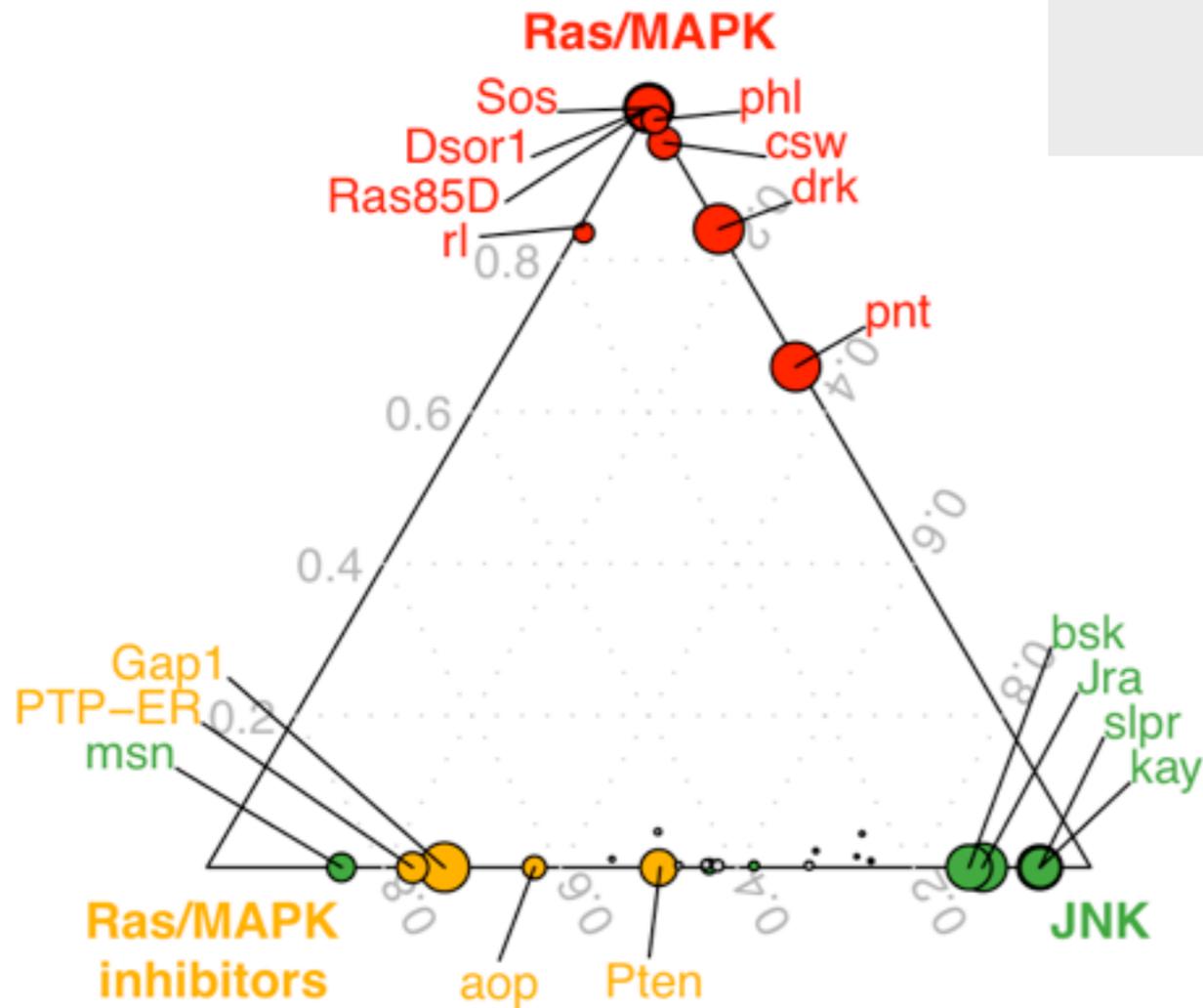


circle sizes ~ cross-validated posterior probabilities of the classifier

# Classification of genes based on their interactions

Show Me Your Friends and I'll Tell You Who You Are

cross-validated performance on training set

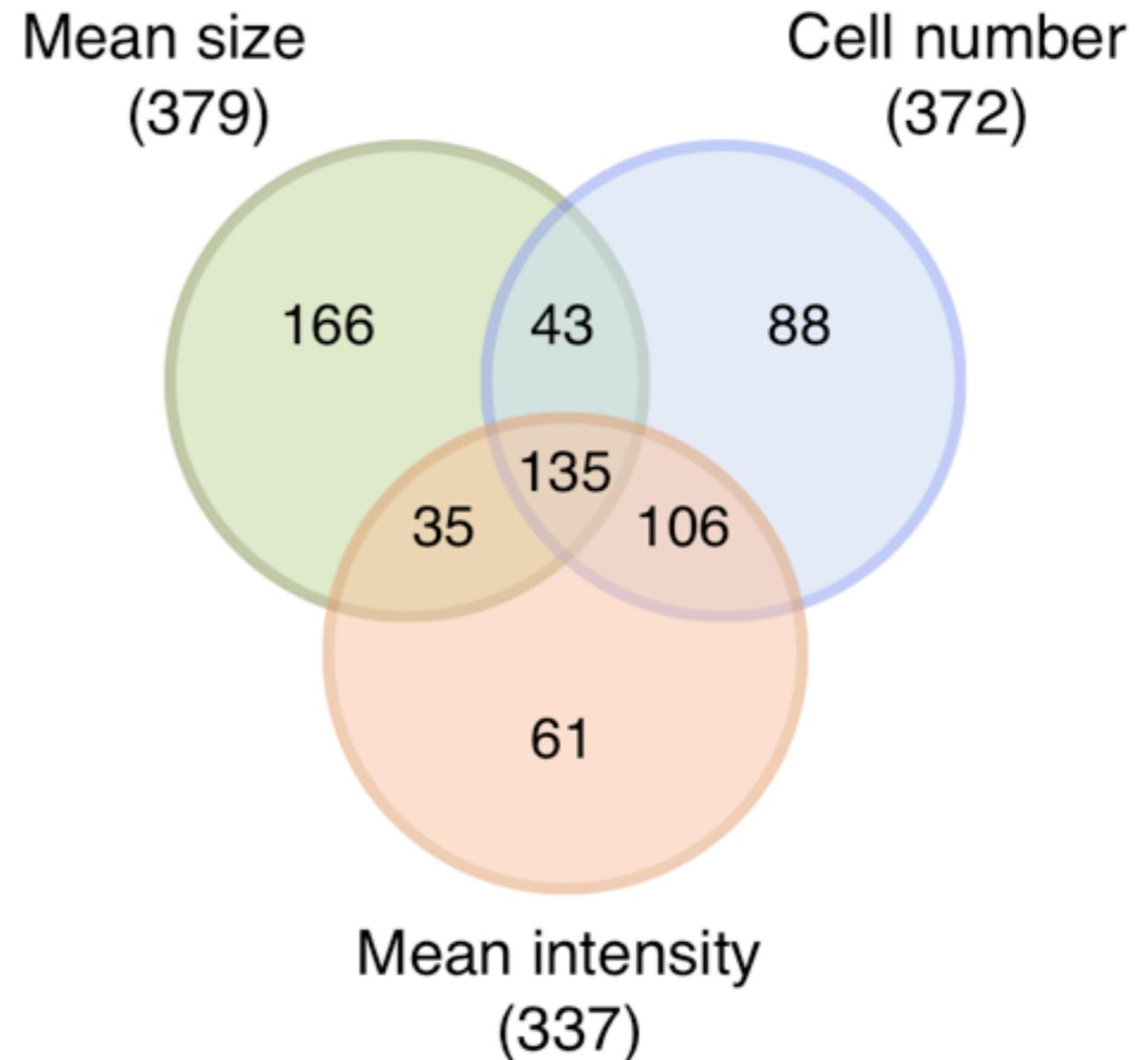


circle sizes ~ cross-validated posterior probabilities of the classifier

# Genetic interactions in 3 dimensions

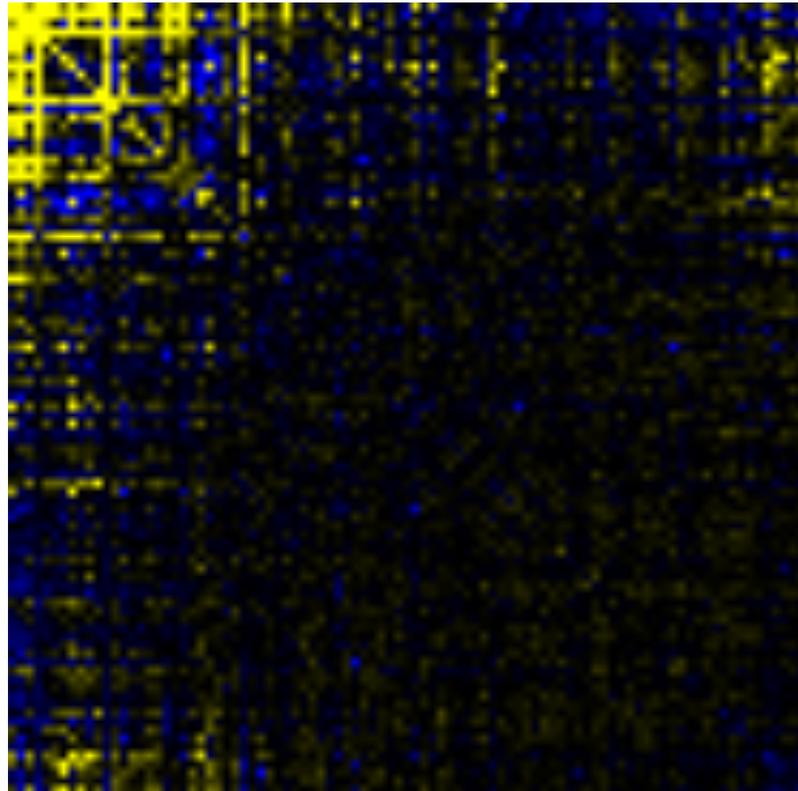
**Different phenotypes produce different sets of interactions**

**For each set, significant overlap with known genetic interactions and with human interologs**

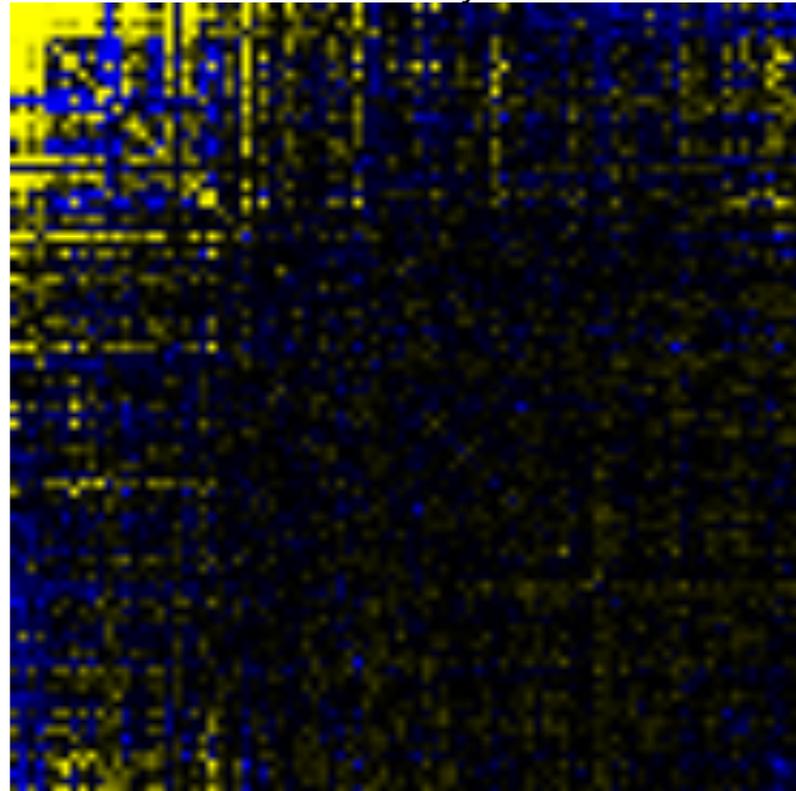


# Interaction matrices

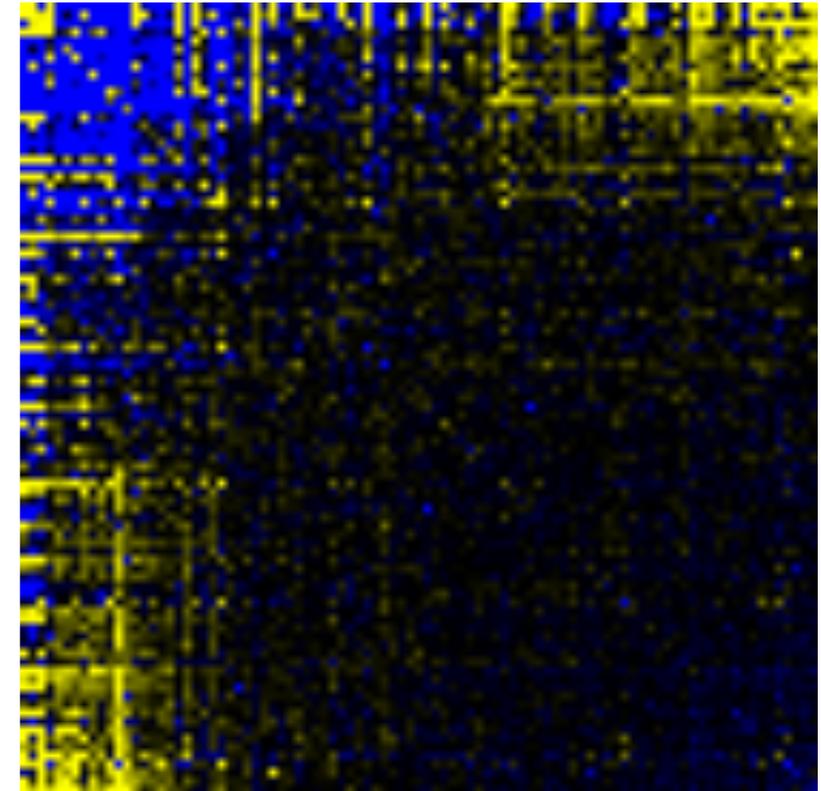
number of cells  
nrCells



intensity  
intensity

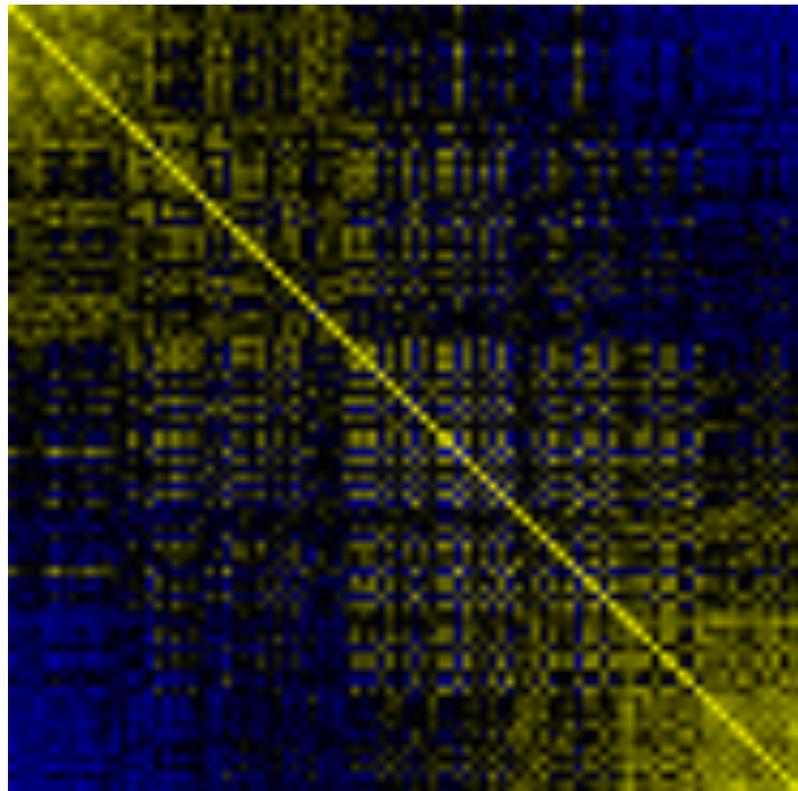


area  
area

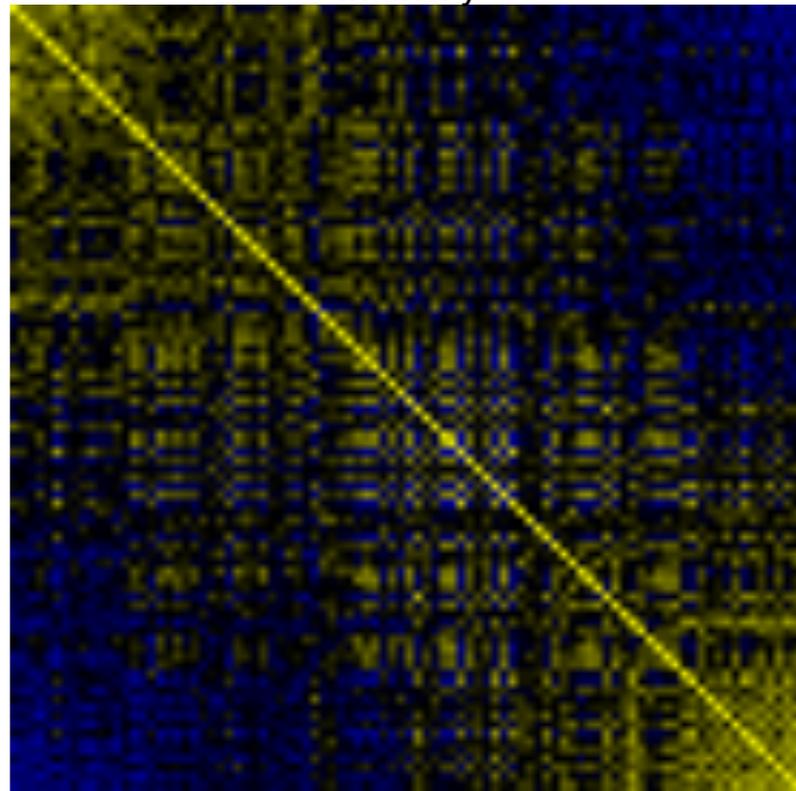


# Correlation matrices

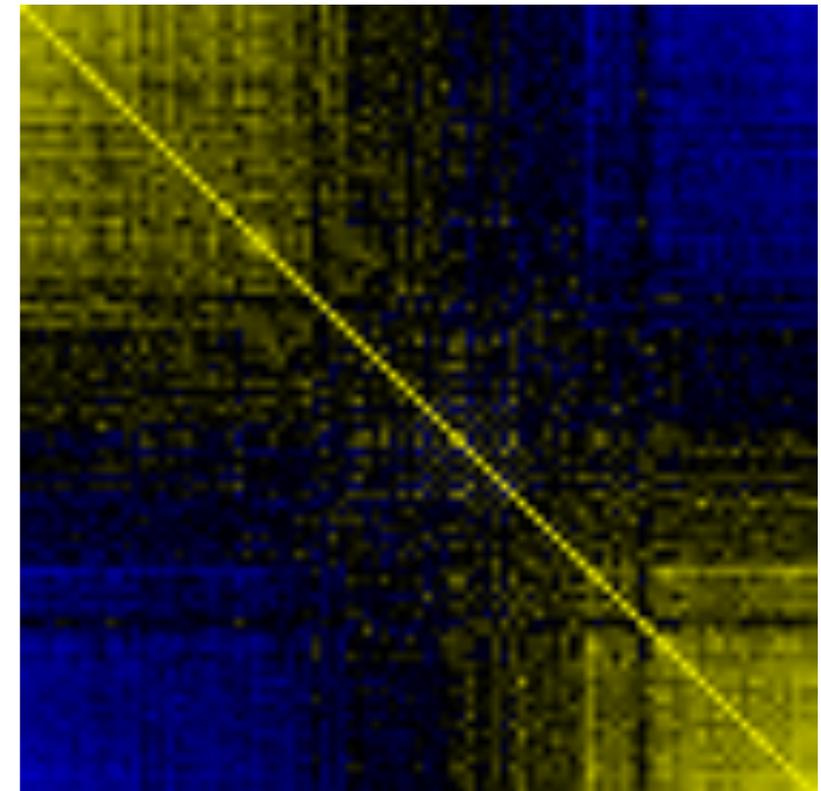
nrCells



intensity

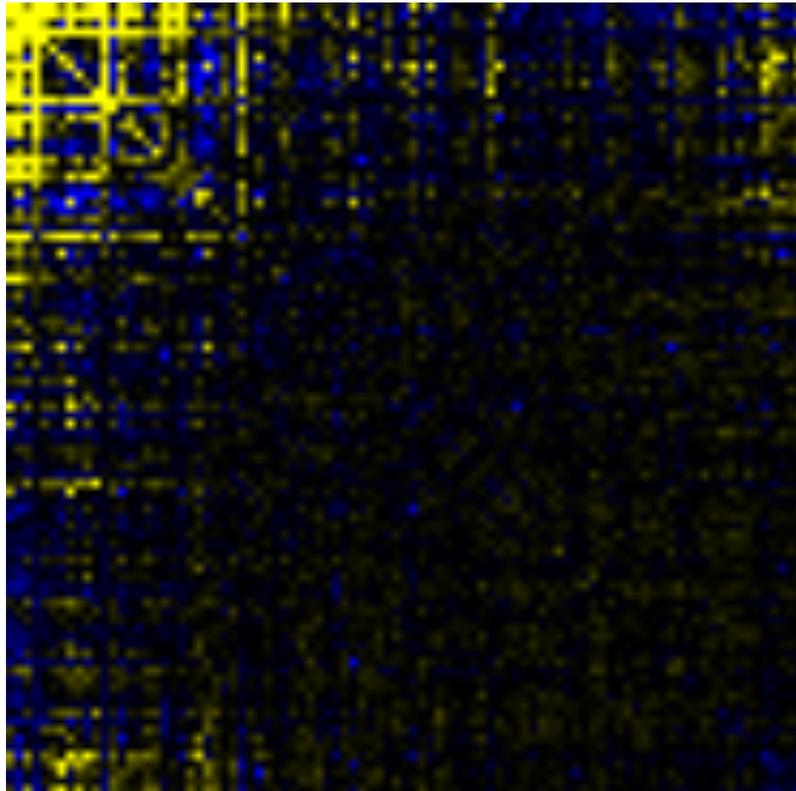


area

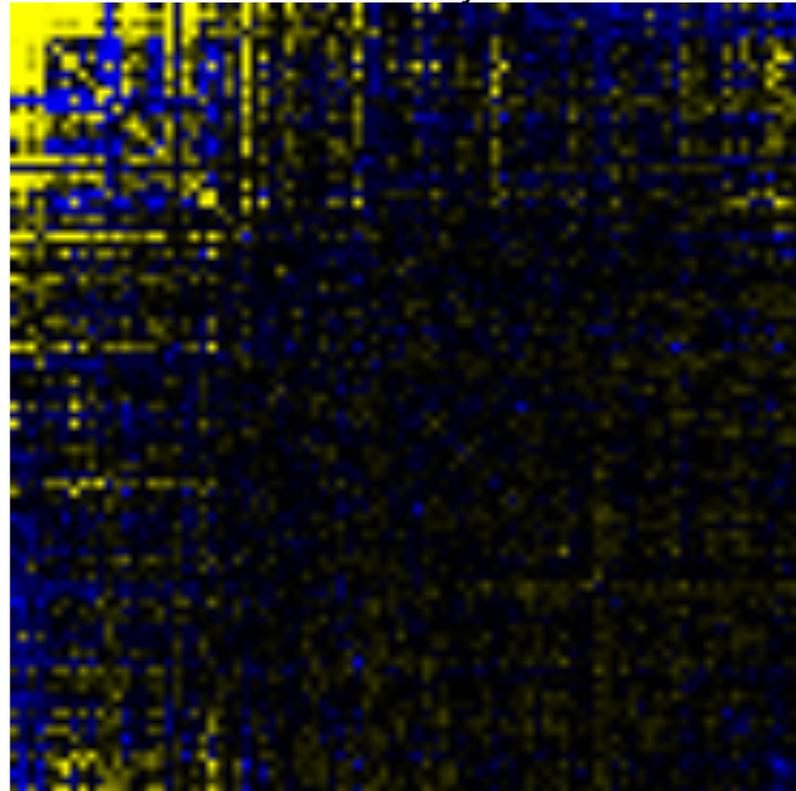


# Interaction matrices

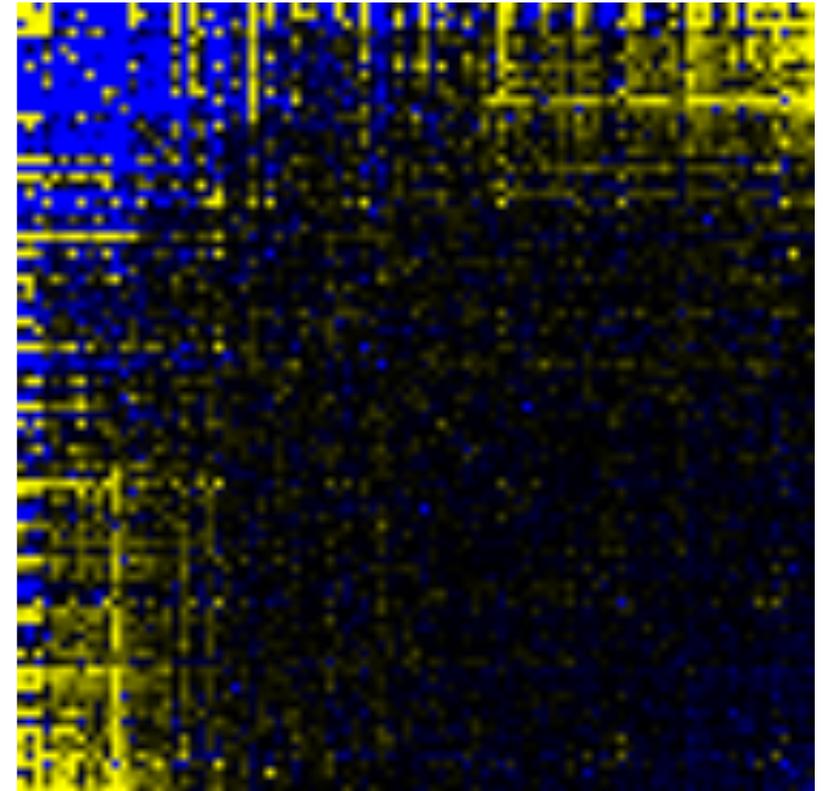
number of cells  
nrCells



intensity  
intensity

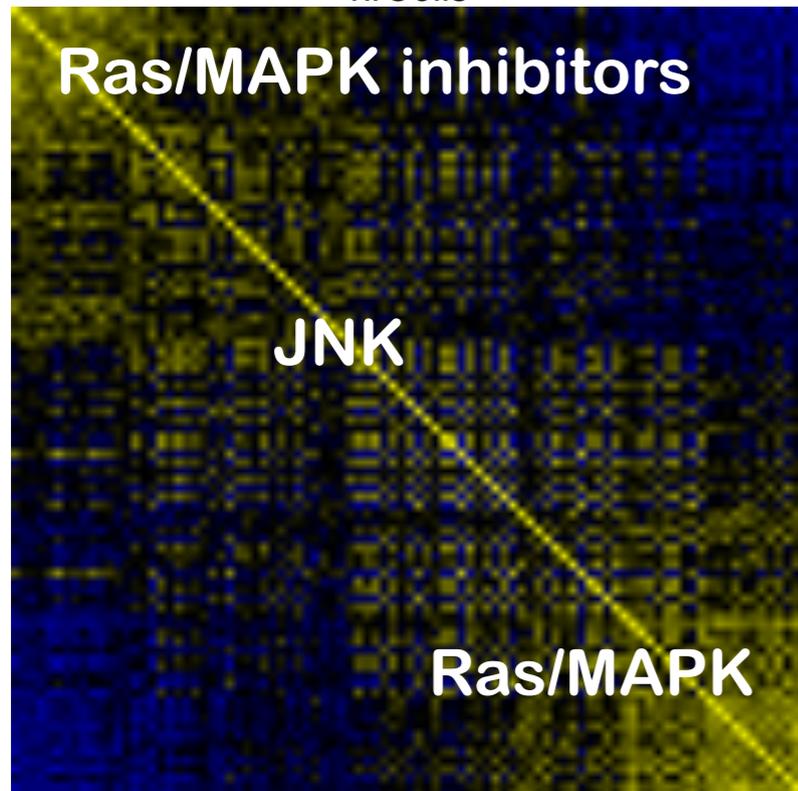


area  
area

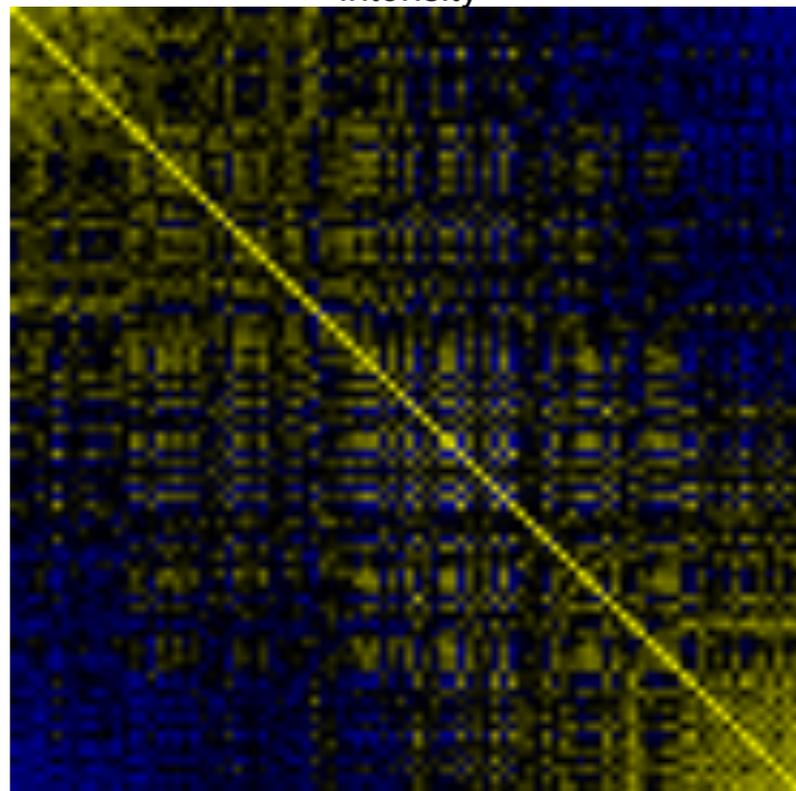


# Correlation matrices

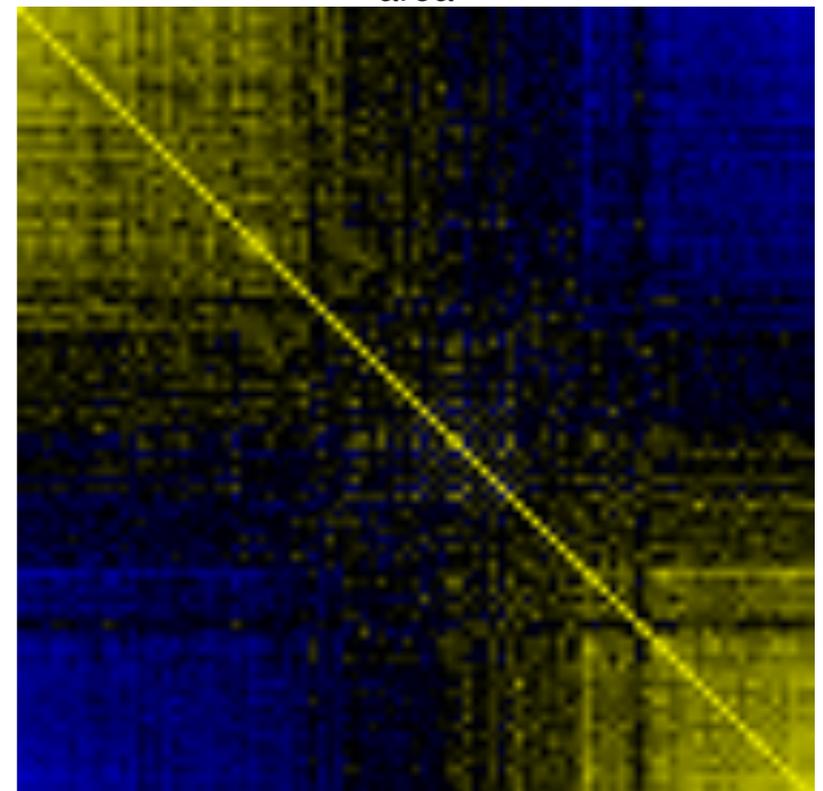
nrCells



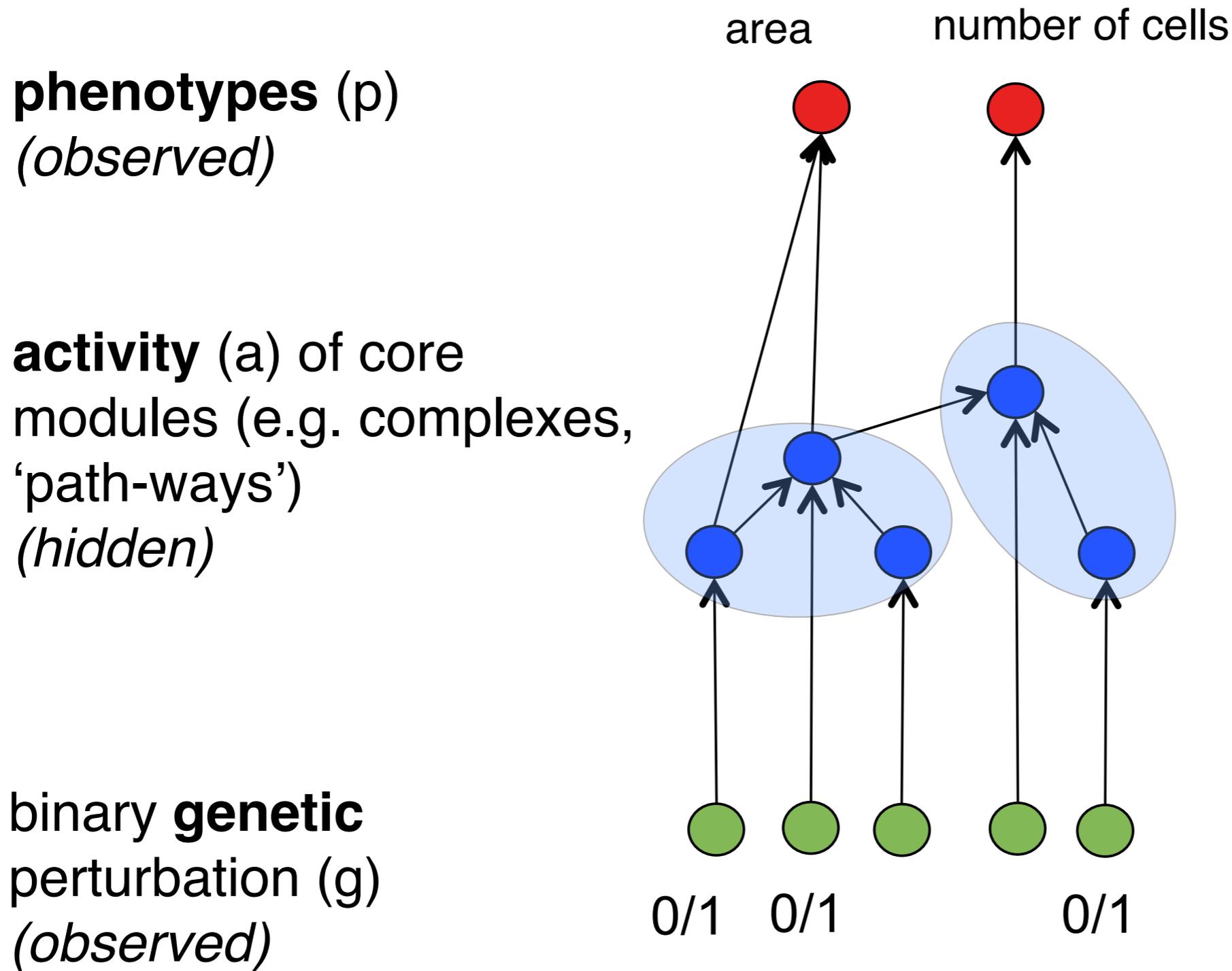
intensity



area



# Network learning - identify the underlying molecular modules



$$P(p | g; \alpha, \beta, \gamma) = \sum_a P(p | a; \alpha) \prod_{i=1}^N P(a_i | a_{pa(i)}, g_{pa(i)}; \beta, \gamma)$$

# Ongoing: a much bigger matrix

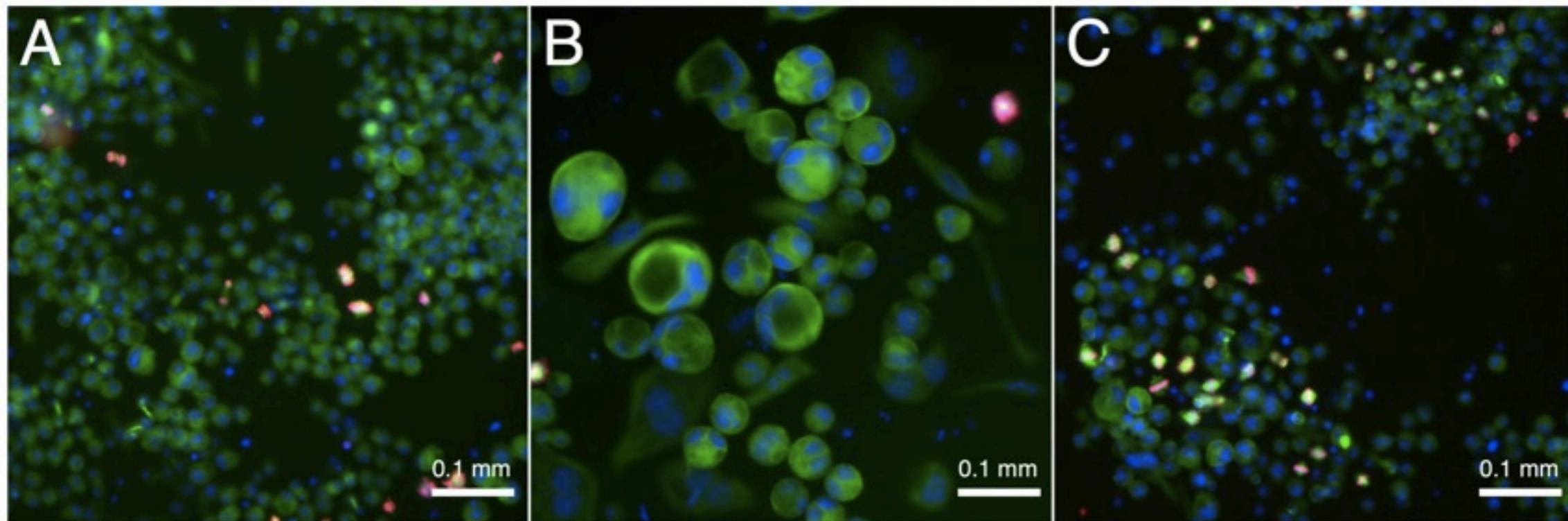
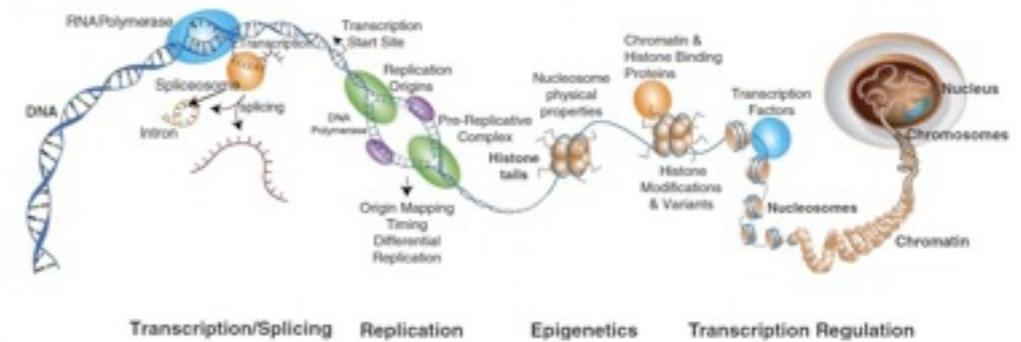
- Larger matrix, again Dmel2 cells
- ~1500 chromatin-related genes x 100 query genes
- full microscopic readout (4x and 20x), 3 channels:

- \* DAPI

- \* phospho-His3 (mitosis marker)

- \* aTubulin (for spindle phenotypes)

- 1600 384-well plates, ~ 300.000 measurements

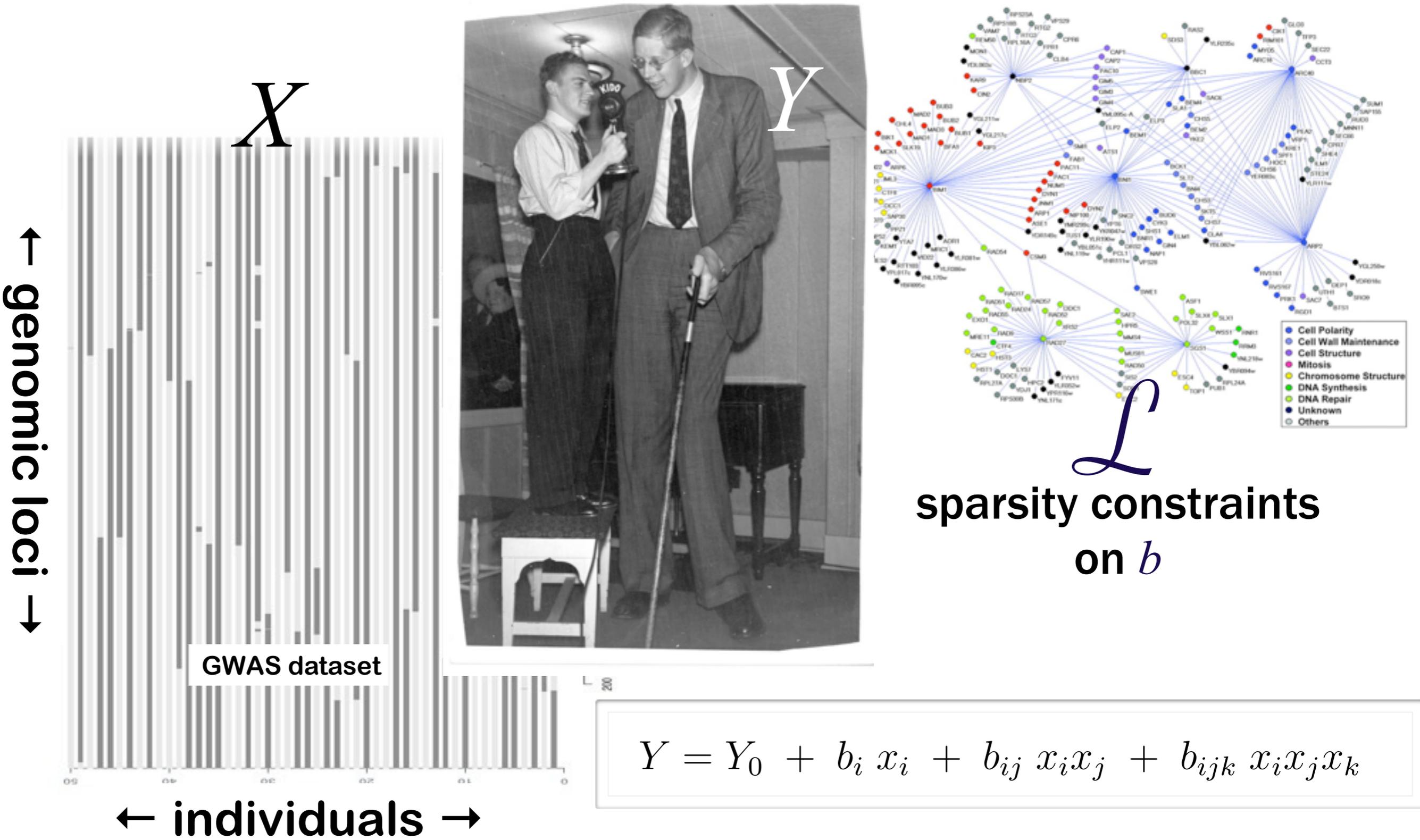


ctrl dsRNA

Rho1 dsRNA

Dynein light chain dsRNA

# Outlook: genetic interactions from model system experiments as regularisation/priors for the identification of genetic interactions in observational studies



# Summary

Quantitative, combinatorial RNAi works in metazoan cells.  
Technological tour de force; data exploration, QA/QC,  
normalisation and transformation....

Individual genetic interactions vs interaction profiles.

Data are high-dimensional and complicated:

- dose effects,
- different / multivariate phenotypes
- relative timing

reveal non-redundant interactions.

All data & code available from  **BIOCONDUCTOR**

**Bernd Fischer,**

Thomas Horn, Thomas Sandmann, Michael Boutros  
Nature Methods 2011(4)





**Simon Anders**  
**Joseph Barry**  
**Bernd Fischer**  
**Ishaan Gupta**  
**Felix Klein**  
**Gregoire Pau**  
**Aleksandra Pekowska**  
**Paul-Theodor Pyl**  
**Alejandro Reyes**

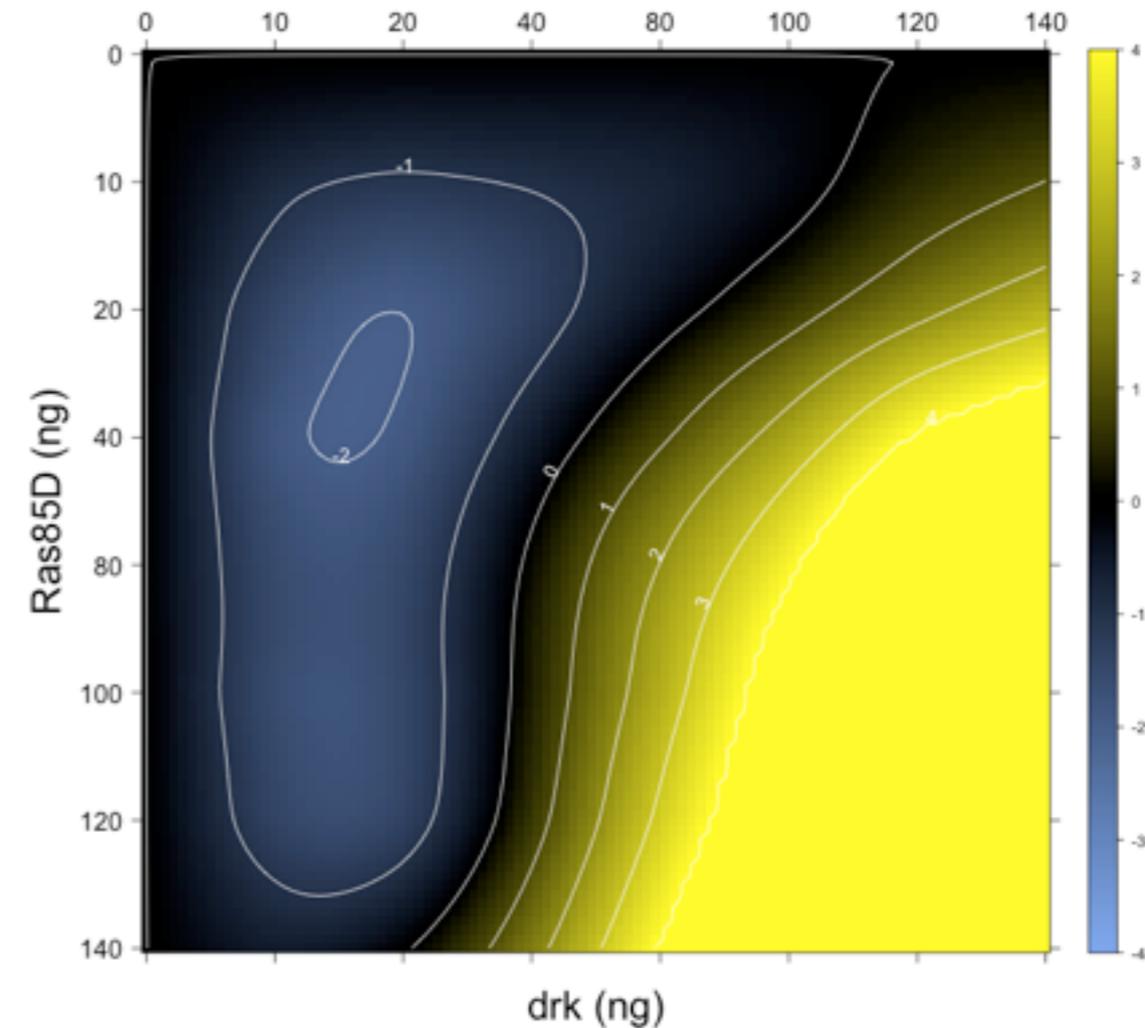
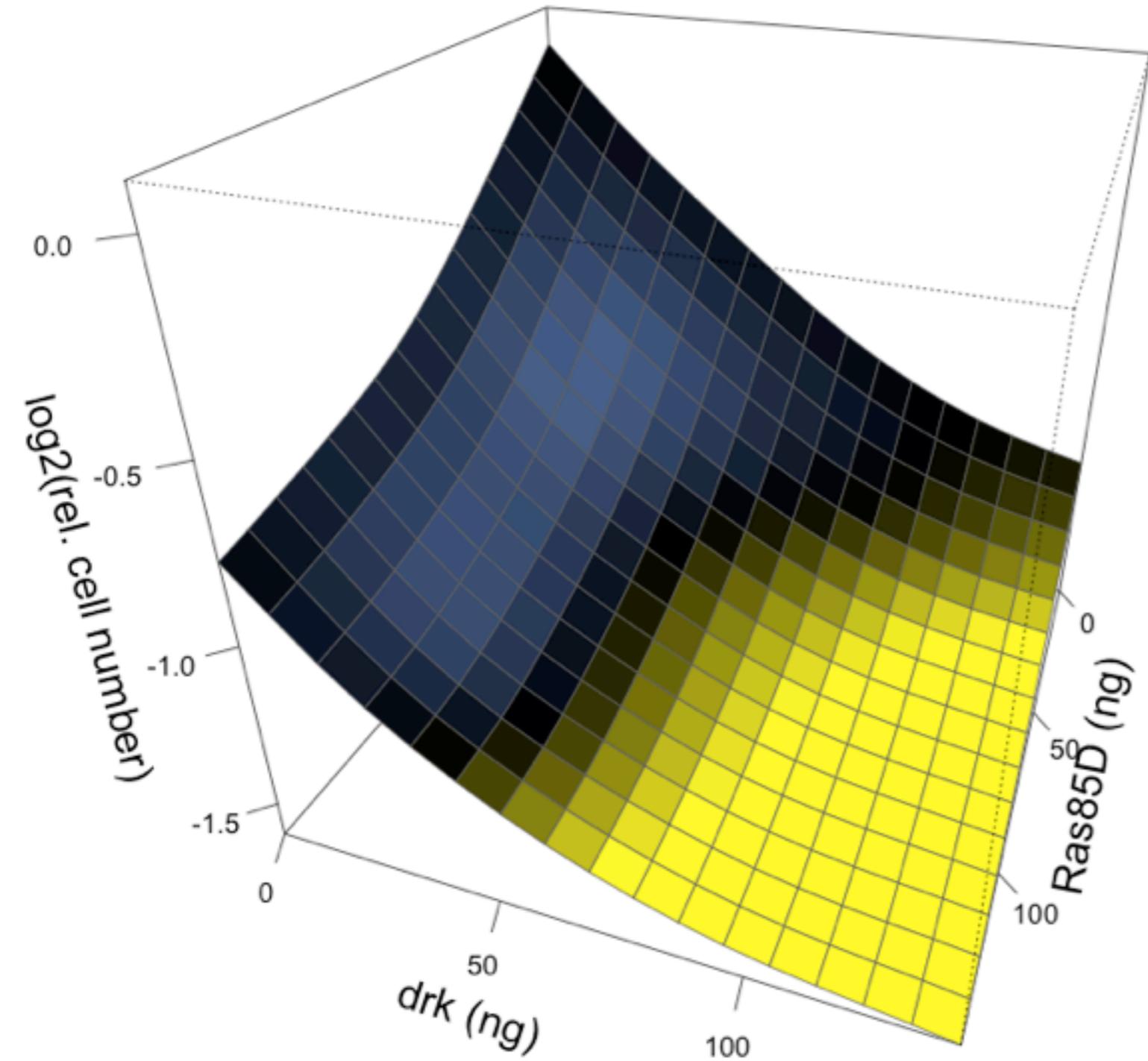


**Collaborators**  
**Lars Steinmetz**  
**Michael Boutros (DKFZ)**  
**Robert Gentleman (Genentech)**  
**Jan Korbel**

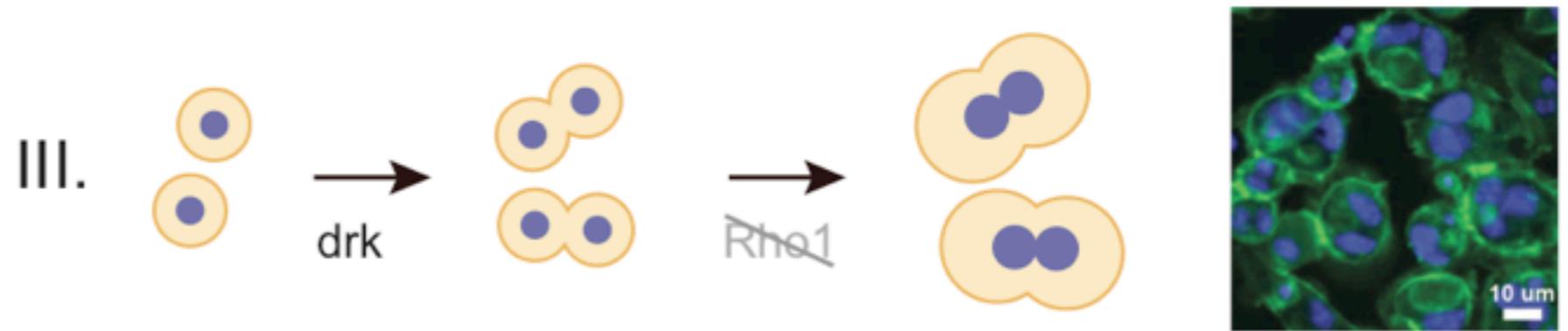
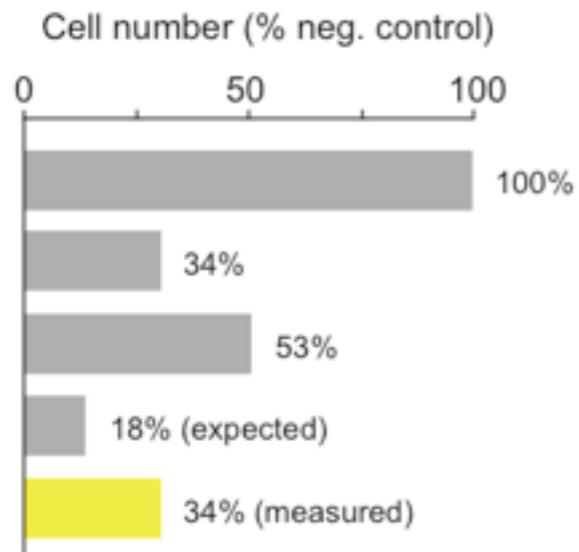
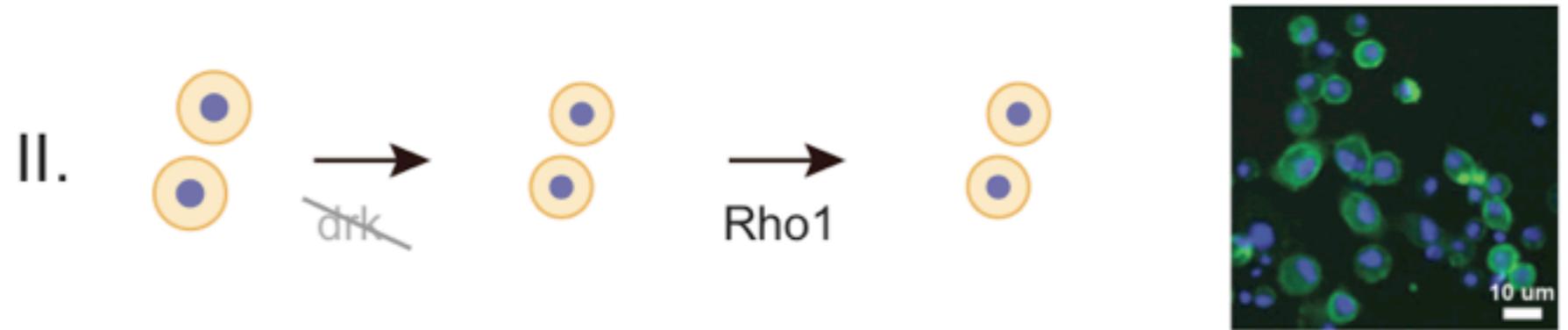
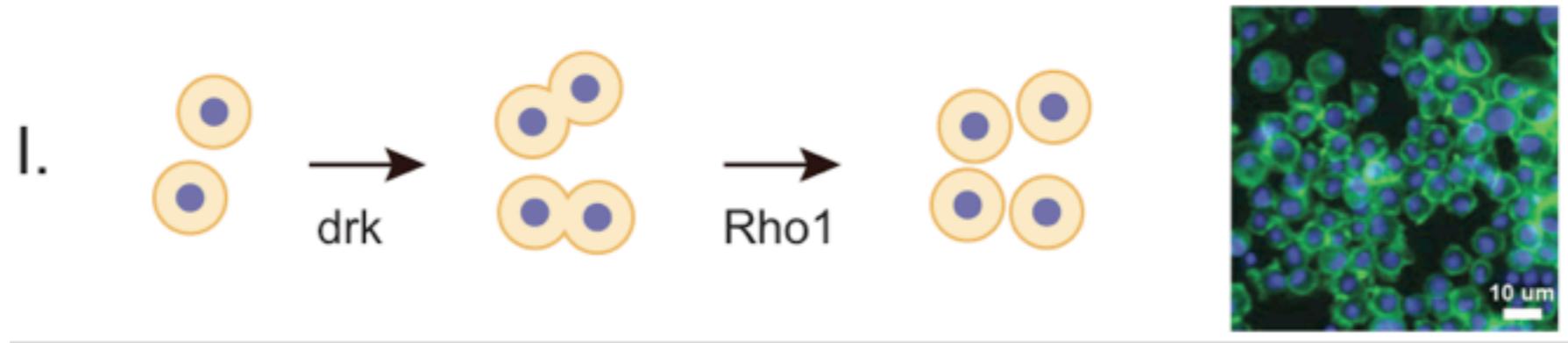
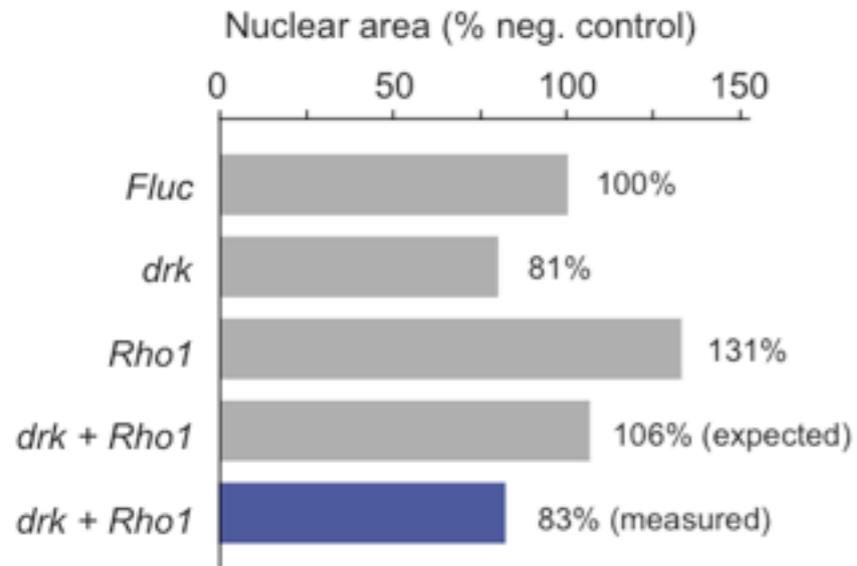
**Michael Knop (Uni HD)**  
**Jan Ellenberg**  
**Kathryn Lilley (Cambridge)**  
**Anne-Claude Gavin**  
**Alvis Brazma (EBI)**  
**Paul Bertone (EBI)**  
**Ewan Birney (EBI)**

# Ras85D and drk: concentration dependence

strength, presence and direction of an interaction can depend on reagent concentration (cf. drug-drug interactions)



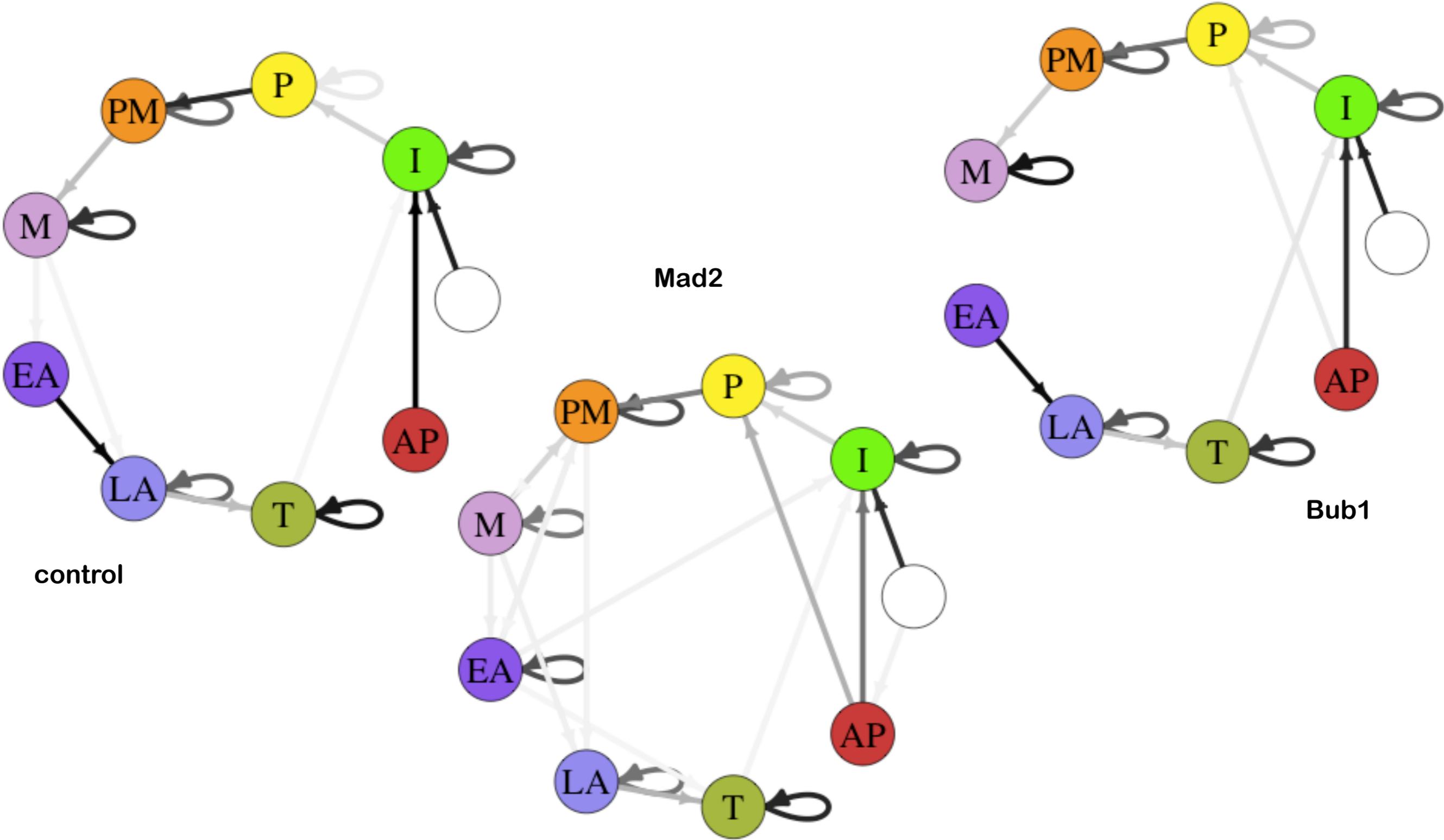
# Sign inversion for different phenotypes

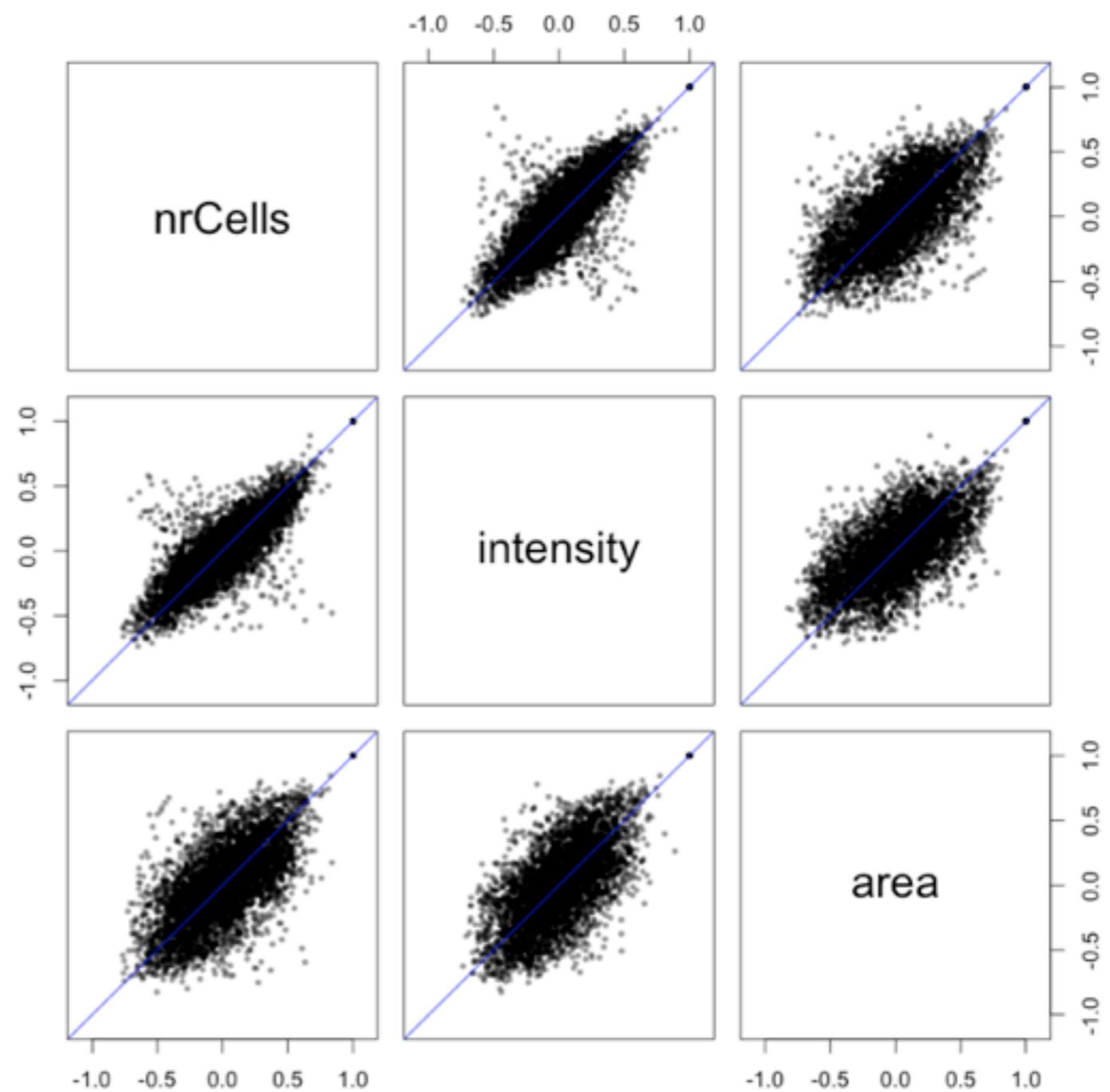
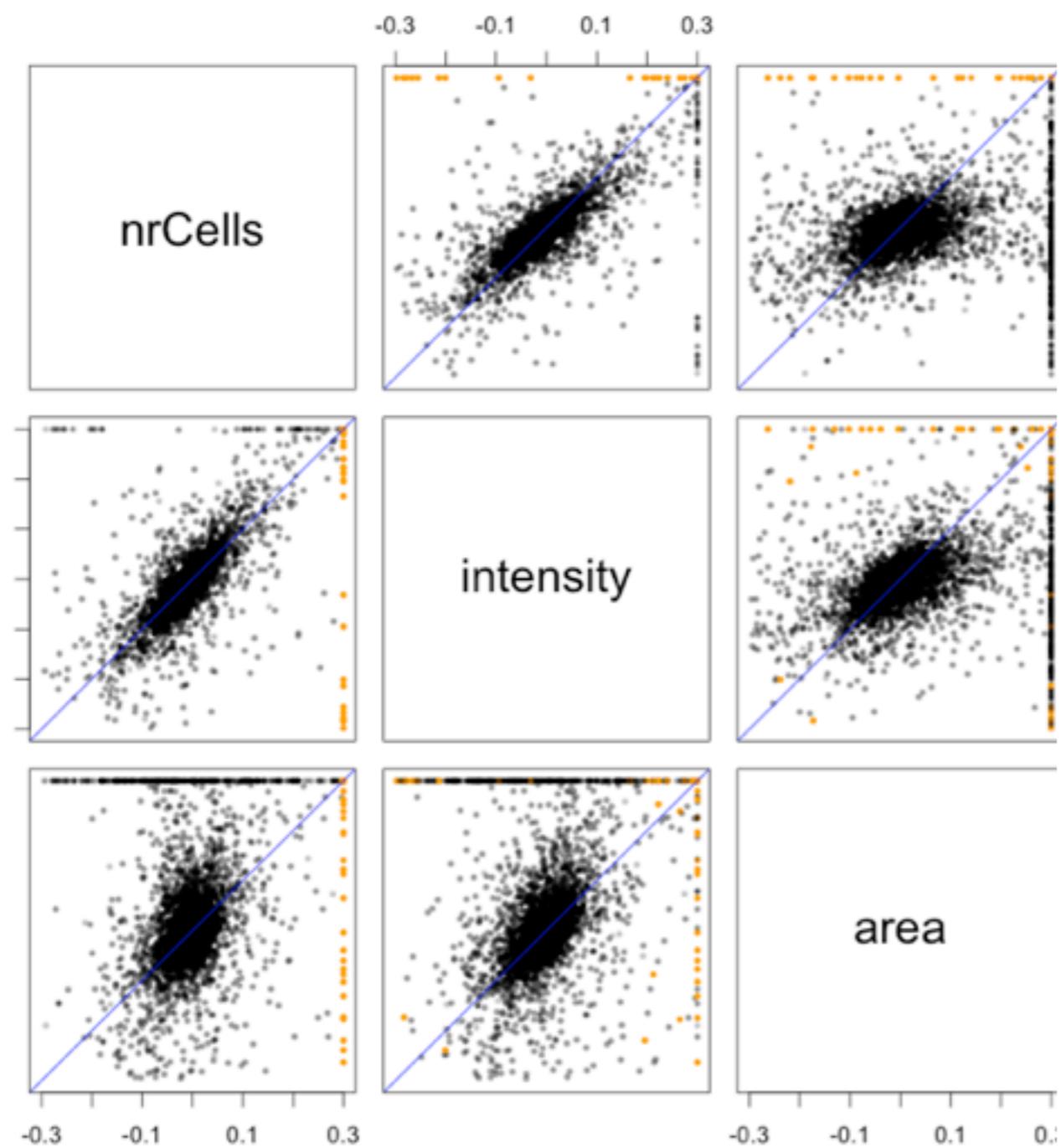


— DNA  
—  $\alpha$ -Tubulin

# Hidden Markov Model on class labels: parameters summarise the data

Learn HMM on class labels

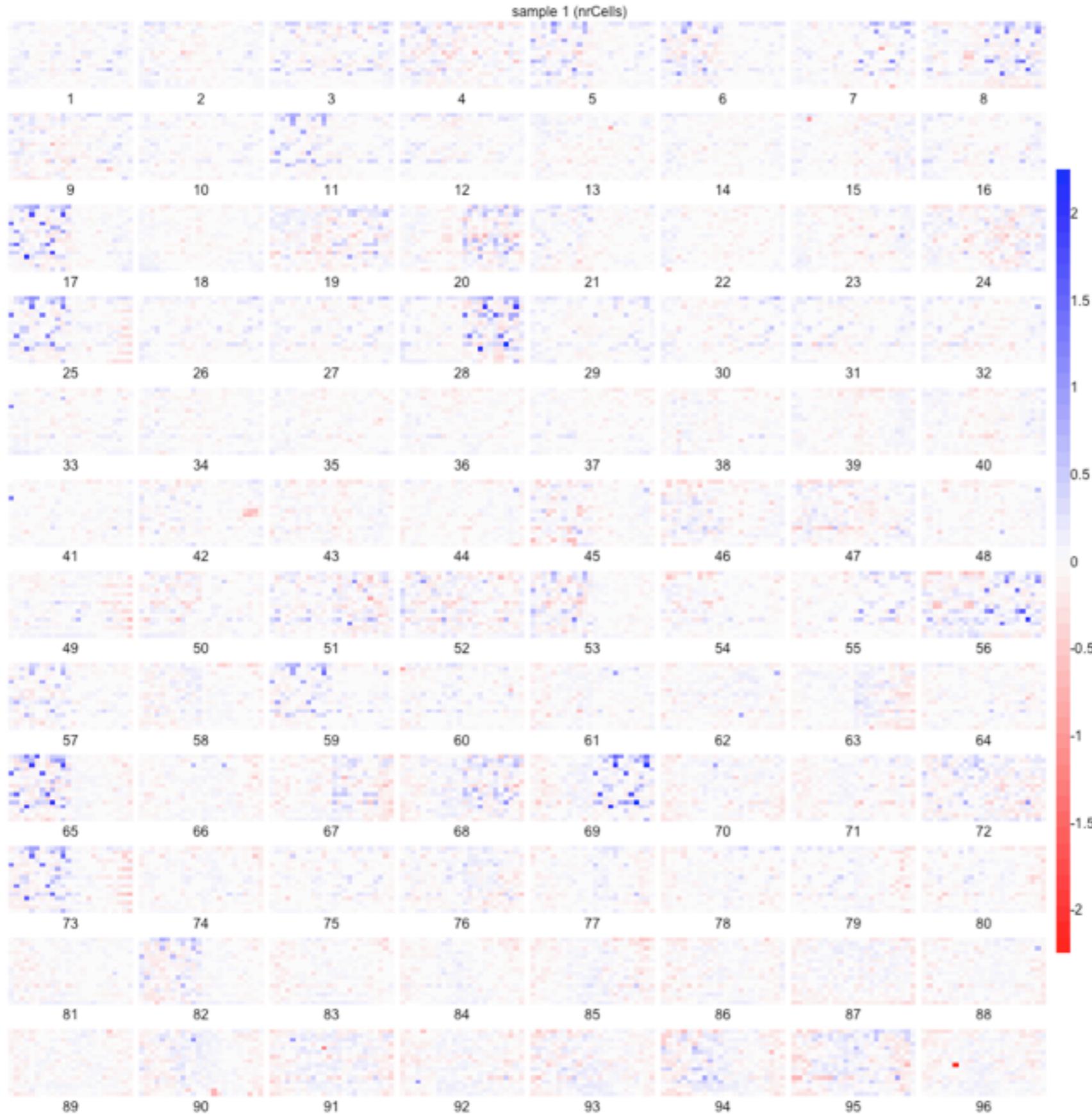




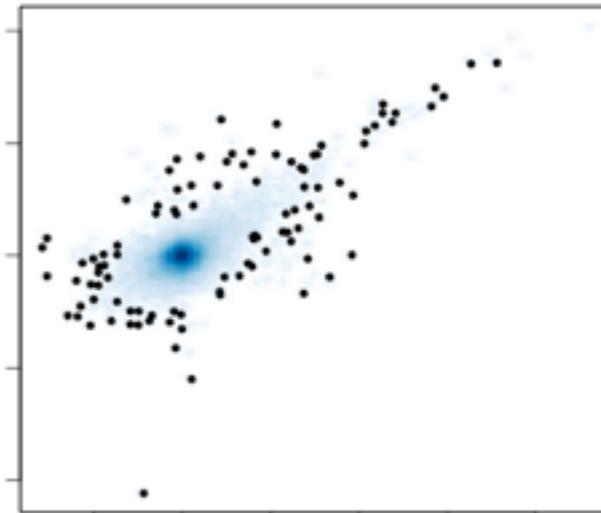
Interaction scores

Correlations

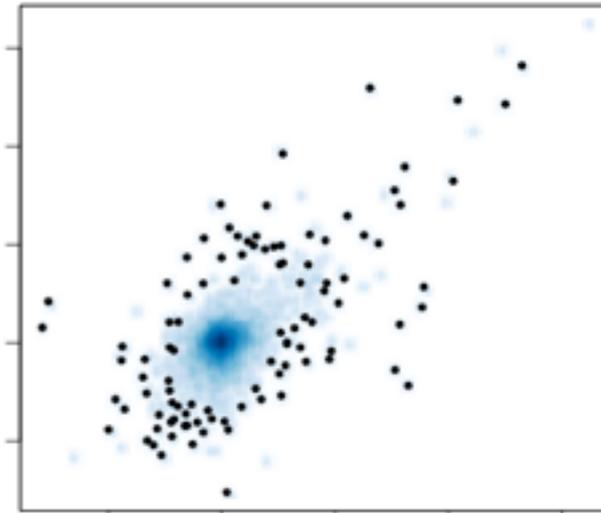
# Screen Plot of Interaction Score (#cells)



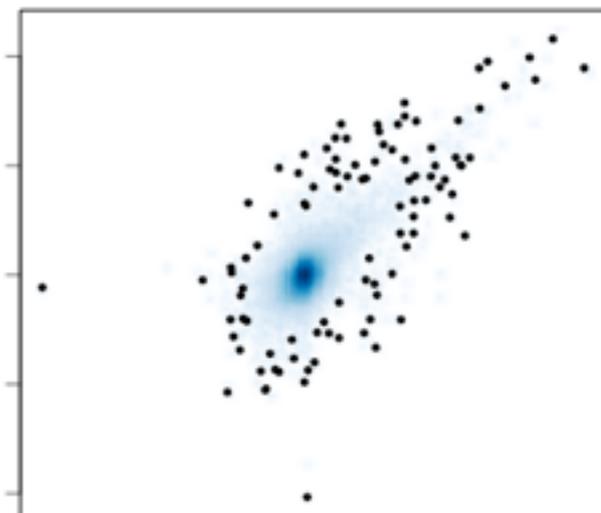
within  
screen  
replicates  
(cor=0.968)



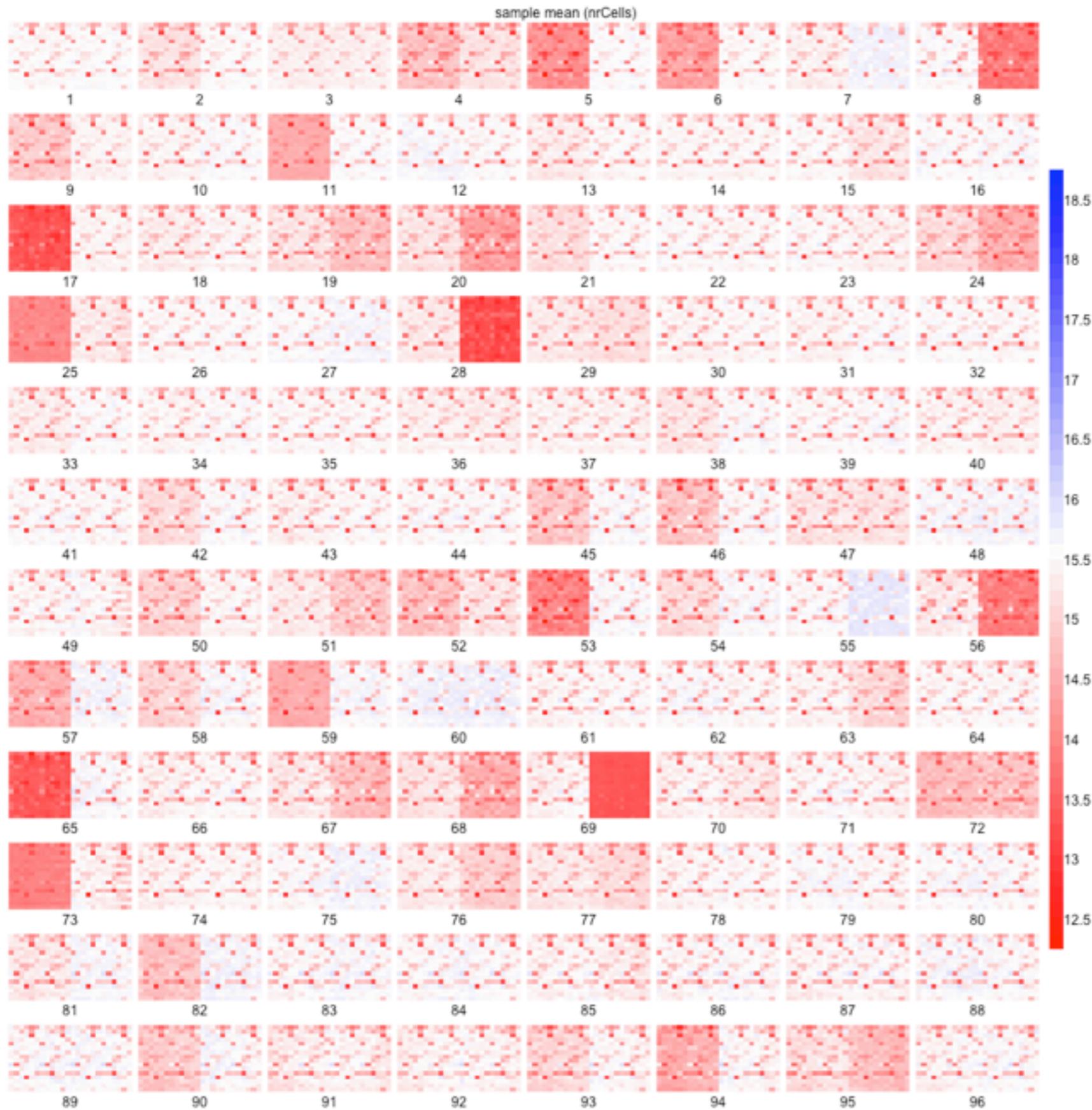
independent  
daRNA  
designs  
(cor=0.902)



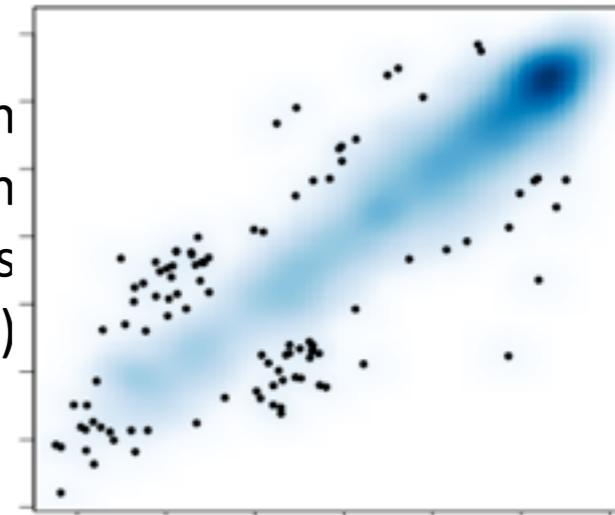
between  
screen  
replicates  
(cor=0.948)



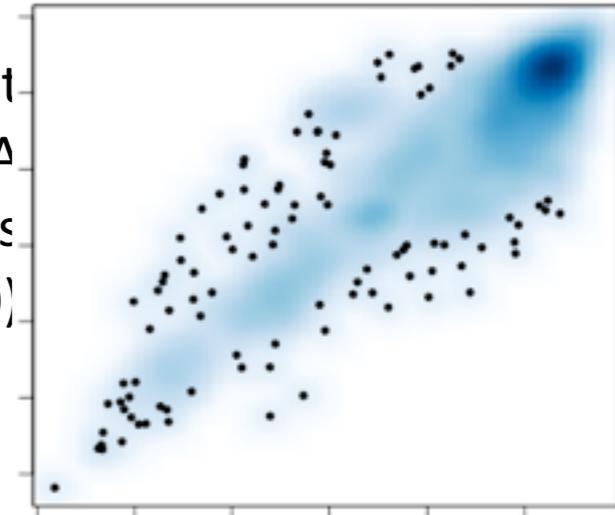
# Screen Plot of Read-out (Number of Cells)



within  
screen  
replicates  
(cor=0.97)



independent  
dsRNA  
designs  
(cor=0.90)



between  
screen  
replicates  
(cor=0.95)

