

Quantitative Analysis of Dynamic Contrast-Enhanced MRI using R

The **dcmriS4** package

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20 July 2010 for useR!2010



- 1 Motivation
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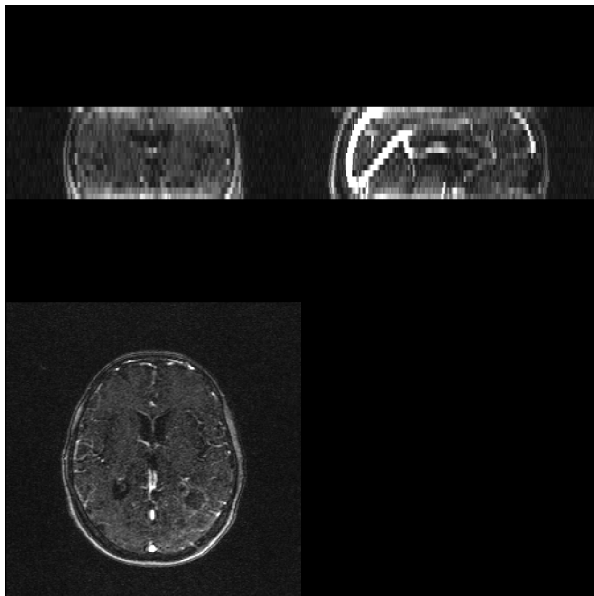


Introduction

- The quantitative analysis of DCE-MRI involves fitting pharmacokinetic (PK) models to the concentration of a contrast agent over time.
- Gadolinium-based contrast agents, involving a small molecular-weight substance, are injected after several baseline scans.
- Using T1-weighted sequences, the reduction in T1 relaxation time caused by the contrast agent is the signal dominant enhancement.
- T1-weighted kinetic curves have three major phases
 - the upslope
 - maximum enhancement
 - washout
- Dynamic acquisition
 - 5-10 minutes for oncology applications
 - 60-90 minutes for neurology (BBB) applications
- **dcmriS4** facilitates all stages of data analysis for DCE-MRI and diffusion-weighted imaging (DWI), using S4 nifti objects.



RIDER Neuro MRI



Data Acquisition

- 1 Localizer
- 2 Structural scans
- 3 Multiple flip angles
- 4 B1 characterization (3T or higher fields)
- 5 Dynamic (bolus injection + 30s)
- 6 Structural scans
- 7 DWI



Data Acquisition

- 1 Motion correction and co-registration
- 2 T1 relaxation
- 3 Gadolinium concentration
- 4 B1 mapping
 - For higher field strengths (3T or more)
- 5 Arterial input function
- 6 Kinetic parameter estimation
- 7 Statistical inference



Tips and Tricks

- Automation is the key to accurate and consistent results
- Quantitative analysis of DCE-MRI depends on several key acquisition parameters
- **oro.dicom** provides the facility of converting DICOM header information into a CSV file

```
path <- file.path(".")
subject <- 1086100996
dcm <- dicomSeparate(file.path(path, subject))
## Save DICOM header information to CSV file
dcm.csv <- dicomTable(dcm$hdr)
write.csv(dcm.csv, file=paste(subject, "csv", sep="."))
```



T1 Relaxation

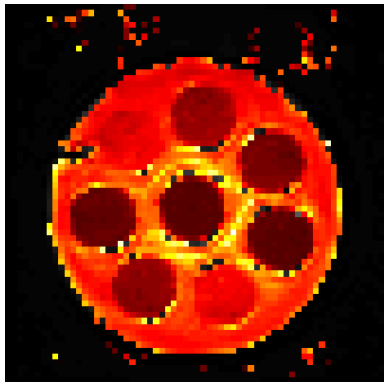


Figure: T1 Phantom

- Parametric form dictated by physics
- Multiple flip-angle acquisitions
- Nonlinear regression
 - Levenburg-Marquardt
 - **minpack.lm**
- B1 field correction is possible

R code for T1 Estimation

```
R> alpha <- c(5, 10, 20, 25, 15)
R> TR <- 4.22 / 1000 # seconds
R> R1 <- R1.fast(flip, mask, alpha, TR, verbose = TRUE)
  Deconstructing data...
  Calculating R10 and M0...
  Reconstructing results...
R> overlay(vibe, 1/R1$R10[, , 1:nsli(vibe)], z = 13,
+         zlim.x = c(0, 1024), zlim.y = c(0, 2.5),
+         plot.type = "single")
```

- Flip angles in degrees
- Repetition time in seconds
- Signal intensities = 4D array
- Mask = 3D array
- Visualization provided by `overlay()` in **oro.nifti**



Arterial Input Functions

$$C_p(t) = D [a_1 \exp(-m_1 t) + a_2 \exp(-m_2 t)]$$

Variables

- $D = 0.1 \text{mmol/kg}$, $a_1 = 3.99 \text{kg/l}$, $a_2 = 4.78 \text{kg/l}$, $m_1 = 0.144 \text{min}^{-1}$ and $m_2 = 0.0111 \text{min}^{-1}$
 - Weinmann *et al.* (1984); Tofts and Kermode (1984).
- $D = 0.1 \text{mmol/kg}$, $a_1 = 2.4 \text{kg/l}$, $a_2 = 0.62 \text{kg/l}$, $m_1 = 3.0 \text{min}^{-1}$ and $m_2 = 0.016 \text{min}^{-1}$
 - Fritz-Hansen *et al.* (1996).

$$C_p(t) = A_B t \exp(-\mu_B t) + A_G [\exp(-\mu_G t) - \exp(-\mu_B t)],$$

Variables

- Orton *et al.* (2008)

Seed-based algorithm in `extract.aif()`



Kinetic Parameter Estimation

- The standard Kety (1951) model, a single-compartment model, or the extended Kety model, forms the basis for **dcmriS4**.

$$C_t(t) = K^{\text{trans}} [C_p(t) \otimes \exp(-k_{\text{ep}}t)]$$

$$C_t(t) = v_p C_p(t) + K^{\text{trans}} [C_p(t) \otimes \exp(-k_{\text{ep}}t)]$$

- Estimation techniques include:
 - Nonlinear regression (Levenburg-Marquardt, **minpack.lm**)
 - Bayesian estimation via MCMC (Schmid *et al.* 2006)
 - Bayesian estimation via MAP (adaptation of Schmid *et al.* 2006)
 - Bayesian estimation via penalized B-splines (Schmid *et al.* 2009a)
- Hierarchical Bayesian methods are not available at this time in **dcmriS4**, but will be in **PILFER**.
 - <http://pilfer.sourceforge.net>

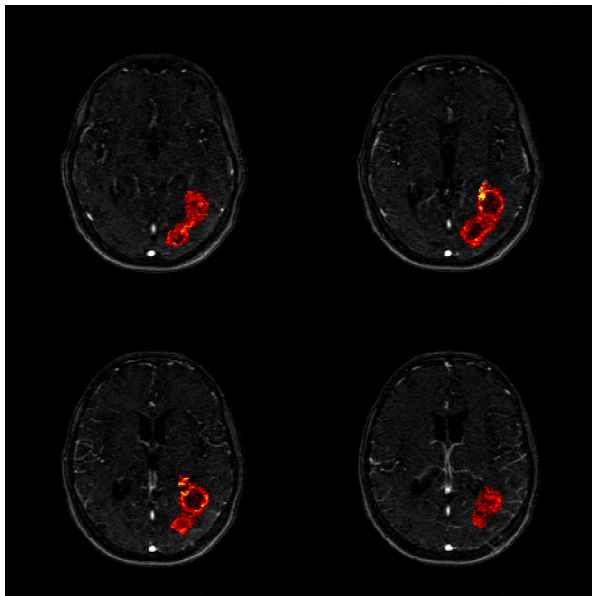


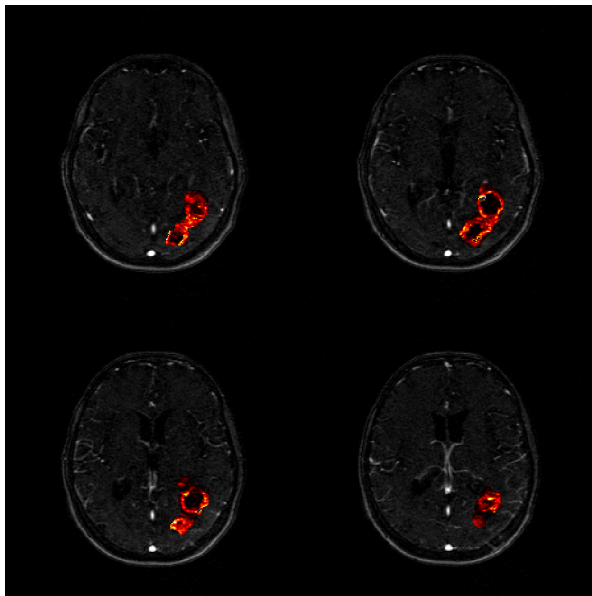
Kinetic Parameter Estimation

```
acqtimes <- str2time(unique(sort(scan("rawtimes.txt", quiet=TRUE))))$time
conc <- readNIfTI(paste(s, d, "perfusion", "gdconc", sep="_"))
mask <- readANALYZE(paste(s, d, "perfusion", "mask2", sep="_"))
fit <- dcmri.lm(conc, (acqtimes - acqtimes[8]) / 60,
               ifelse(mask > 0, TRUE, FALSE), model="extended",
               aif="fritz.hansen", verbose=TRUE)
writeNIfTI(fit$ktrans, paste(s, d, "perfusion", "ktrans", sep="_"))
overlay(dyn[, ,6:9,],
       ifelse(fit$ktrans[, ,6:9] < 0.25, fit$ktrans[, ,6:9], NA),
       w=11, zlim.x=c(32,512), col.y=hotmetal(), zlim.y=c(0,.1))
```

- Acquisition times are found in the DICOM data
 - DICOM header fields are vendor dependent
 - Zero must be defined as time of gadolinium injection
- Mask was created in FSLView
- Visualization provided by `overlay()` in **oro.nifti**
- Time conversion provided by `str2time()` in **oro.dicom**



RIDER Neuro MRI: K^{trans} 

RIDER Neuro MRI: k_{ep} 

Statistical Inference

- Methodology for statistical inference is not included in the **dcmriS4** package.
- Please use the models/tests in **R** to perform hypothesis tests.
- Hierarchical Bayesian models are available, but not using **R**.
 - See Schmid *et al.* (2009b) for more information.



Conclusions

- The package **dcmriS4** attempts to provide quantitative methods for DCE-MRI
 - Vendor software (GE, Siemens, Philips, etc.)
 - Proprietary software (JIM, etc.)
 - Home-grown solutions
- Future directions
 - Multi-compartment models (Buckley *et al.*)
 - Parallelization (e.g., **multicore**)
 - Semi-parametric procedures (AUC, T_{max} , C_{max} , etc.)
- Feedback
 - Please provide feedback (pos/neg) on the SourceForge forum or mailing list.



References I

- Fritz-Hansen, T., E. Rostrup, *et al.* (1996). Measurement of the arterial concentration of Gd-DTPA using MRI: A step toward quantitative perfusion imaging. *Magnetic Resonance in Medicine*, **36**, 225–231.
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References II

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